

PALEOCEANOGRAPHICAL PROXIES BASED ON DEEP-SEA BENTHIC FORAMINIFERAL ASSEMBLAGE CHARACTERISTICS

Frans J. Jorissen*, Christophe Fontanier *and* Ellen Thomas

Contents

| | |
|---|-----|
| 1. Introduction | 263 |
| 1.1. General introduction | 263 |
| 1.2. Historical overview of the use of benthic foraminiferal assemblages | 266 |
| 1.3. Recent advances in benthic foraminiferal ecology | 267 |
| 2. Benthic Foraminiferal Proxies: A State of the Art | 271 |
| 2.1. Overview of proxy methods based on benthic foraminiferal assemblage data | 271 |
| 2.2. Proxies of bottom water oxygenation | 273 |
| 2.3. Paleoproductivity proxies | 285 |
| 2.4. The water mass concept | 301 |
| 2.5. Benthic foraminiferal faunas as indicators of current velocity | 304 |
| 3. Conclusions | 306 |
| Acknowledgements | 308 |
| 4. Appendix 1 | 308 |
| References | 313 |

1. INTRODUCTION

1.1. General Introduction

The most popular proxies based on microfossil assemblage data produce a quantitative estimate of a physico-chemical target parameter, usually by applying a transfer function, calibrated on the basis of a large dataset of recent or core-top samples. Examples are planktonic foraminiferal estimates of sea surface temperature (Imbrie & Kipp, 1971) and reconstructions of sea ice coverage based on radiolarian (Lozano & Hays, 1976) or diatom assemblages (Crosta, Pichon, & Burckle, 1998). These

* Corresponding author.

methods are easy to use, apply empirical relationships that do not require a precise knowledge of the ecology of the organisms, and produce quantitative estimates that can be directly applied to reconstruct paleo-environments, and to test and tune global climate models.

Assemblage-based proxy methods that do not yield fully quantitative results have become less popular over the last decennia, mainly because semi-quantitative or qualitative proxy methods implicitly admit a considerable degree of uncertainty. In many allegedly quantitative proxies, however, the error may be as large, but is concealed by the numerical aspect of the estimates, commonly expressed with of 1 or 2 numbers behind the decimal point, suggesting a highly precise and trustworthy reconstruction of the target parameter.

The decreased popularity of assemblage-based proxy methods affects proxies based on benthic foraminifera, although they have been used to reconstruct a wide range of oceanographic parameters, including water depth, water mass properties, bottom water oxygen content, and the extent and/or seasonality of the organic flux to the ocean floor. Although the use of benthic foraminiferal assemblage compositions has become less common, the use of benthic foraminiferal tests as carriers of geochemical proxy methods (stable isotopes, Mg/Ca, Sr/Ca, etc.) has never been so widespread (e.g., Wefer, Berger, Bijma, & Fischer, 1999; Lea, 2004; Lynch-Stieglitz, 2004; Ravizza & Zachos, 2004; Sigman & Haug, 2004).

All geochemical proxies based on the remains of micro-organisms rely on a thorough knowledge of their ecology that determines when, where, and exactly under which conditions the proxy value is fixed in the microfossil test. In the case of benthic foraminifera, their strongly increased application in geochemical studies has led to a renewed interest in ecological studies in field situations, but also under controlled laboratory conditions (e.g., Heinz, Schmiedl, Kitazato, & Hemleben, 2001; Ernst & Van der Zwaan, 2004; Geslin, Heinz, Hemleben, & Jorissen, 2004). Due to these studies and to studies affiliated with the Joint Global Ocean Flux Studies (JGOFS), benthic foraminiferal ecology is much better known than 20 years ago, and it has become clear that the composition of benthic foraminiferal faunas is controlled by a limited number of closely interrelated environmental parameters.

In spite of the significant advances in knowledge of foraminiferal ecology, only hesitant attempts to develop or improve proxy methods based on benthic foraminiferal assemblage composition have been undertaken in recent years. The further elaboration of these proxy methods is hampered by a number of problems, most of which are not unique to benthic foraminifera, and concern most commonly used proxy methods:

- 1) Many of the controlling parameters that the proxy methods aim to reconstruct are strongly interdependent in the present oceans, making the reconstruction of individual parameters particularly difficult.
- 2) Recent ecosystems cover only a fraction of the environmental conditions encountered in past oceans. The successful reproduction of non-analog conditions is a critical point for all proxy methods, particularly because usually we cannot test the validity of the proxy estimates.

- 3) Laboratory experiments may remedy these two problems by observing faunal responses to changes in single environmental parameters, and to environmental conditions not encountered in recent ecosystems (e.g., Chandler, Williams, Spero, & Xiaodong, 1996; Wilson-Finelli, Chandler, & Spero, 1998; Toyofuku & Kitazato, 2005). Unfortunately, experiments with benthic foraminiferal faunas from open ocean environments are complicated (e.g., Hintz et al., 2004). It appears very difficult to artificially create environments in which deep-sea foraminiferal species feed, grow new chambers, and reproduce. Progress in this promising field of research requires important investments and is consequently rather slow.
- 4) Our knowledge of taphonomical processes is insufficient. Taphonomical processes are responsible for important compositional differences between living faunas, the subject of ecological studies, and fossil faunas as used by proxy methods to reconstruct past environmental parameters. As for many other proxies, a major effort is needed to study the transformations and losses taking place during the transition from a living to a sub-recent (core-top) fauna and finally to a fossil fauna.
- 5) A successful proxy method requires calibration with a very large dataset, encompassing as many different ecosystems and ecological settings as possible. The best way to develop such a dataset is by cooperation of a large group of scientists, adopting the same methodology and putting their data in a common database.

Wefer et al. (1999) give an overview of all commonly used paleoceanographical proxies, and their relatively limited attention to benthic foraminiferal assemblage studies reflects their decreased popularity. Following the important progress in foraminiferal ecology in recent years, several papers (e.g., Gooday, 1994; Jorissen, 1999a; Van der Zwaan, Duijnste, Den Dulk, Ernst, & Kouwenhoven, 1999; Murray, 2001; Smart, 2002) investigated the possibility to apply these new findings to paleoceanography. In a very thorough review, Gooday (2003) presents new ecological concepts, and gives an updated overview of the methods using benthic foraminiferal assemblage studies for paleoceanographical reconstructions. The use of benthic foraminiferal tests as carriers of geochemical proxies is reviewed in several publications (e.g., Lea, 2004; Lynch-Stieglitz, 2004; Ravizza & Zachos, 2004; Sigman & Haug, 2004; papers in Elderfield, 2004).

In the present chapter on paleoceanographical proxies based on deep-sea benthic foraminiferal assemblage characteristics we do not aim to present a complete treatment of all existing proxy methods, for which we strongly recommend Gooday (2003). Instead, we will give a rather personal view of the three proxy relationships that in our opinion are most promising: those between benthic foraminiferal faunas and benthic ecosystem oxygenation, export productivity and deep-sea water mass characteristics. For each, we will concentrate on the advantages and shortcomings of the existing methods, and try to indicate possible pathways for future improvements that may lead to a better application of this potentially useful group of micro-organisms. As a consequence of this approach, we will highlight the many problems encountered when developing proxies based on benthic foraminiferal assemblage data, but want to stress that most of these problems are not unique to proxies based on

faunal data. We will limit our treatment to the reconstruction of environmental parameters in open marine ecosystems, and will not consider more coastal ecosystems, such as estuaries, inner continental shelf areas, or reef ecosystems.

1.2. Historical Overview of the Use of Benthic Foraminiferal Assemblages

The practical application of benthic foraminifera started in the beginning of the 20th century, when Joseph Cushman and his co-workers developed their use as biostratigraphic markers, providing age control (Cushman, 1928), and leading to their wide-spread application in oil exploration. Natland (1933) first used benthic foraminifera in a strictly paleo-environmental way to determine the depth of deposition of sediments of the Ventura Basin. Sediments originally interpreted as outer shelf deposits later turned out to be turbidity sequences (Natland & Kuenen, 1951), confirming Natland's original paleobathymetric estimates. For the next 40 years, benthic foraminifera were used extensively to determine paleobathymetry. At first, efforts concentrated on the recognition of isobathyal species, supposed to have the same depth distribution in all oceans (e.g., Bandy, 1953a, 1953b; Bandy & Arnal, 1957; Bandy & Echols, 1964; Bandy & Chierici, 1966; Pujos-Lamy, 1973). In more coastal areas, the ratio between hyaline, porcellaneous and agglutinated taxa was used as an indicator of water depth and salinity (e.g., Bandy & Arnal, 1957; Sliter & Baker, 1972; Murray, 1973; Greiner, 1974). The ratio of planktonic and benthic foraminifera was also proposed as a paleobathymetrical proxy (Grimsdale & Van Morkhoven, 1955).

In this early period multivariate statistical methods were not yet widely available, and only relations between individual species or species groups and single environmental parameters could be analysed, leading to major oversimplification of the complex natural situation in which a host of environmental factors interact and control foraminiferal ecology. Pflum and Frerichs (1976) noted that some "delta-depressed" species in the Gulf of Mexico had a lower upper depth limit in front of the Mississippi delta, than in other areas, whereas others, "delta-elevated" taxa, showed the opposite tendency. They suggested that these differences in bathymetrical range were caused by specific redox conditions, resulting from the input of organic matter by river outflow (Pflum & Frerichs, 1976), and were probably first to recognise the influence of organic input on the bathymetrical range of benthic foraminiferal species.

The early 1970s saw the advent of the water mass concept: different deep-sea water masses are characterised by a specific combination of temperature, salinity and pH, and were supposed to be inhabited by highly characteristic faunas (e.g., Streeter, 1973; Schrader, 1974; Lohmann, 1978a). On the basis of this hypothesis it should have become possible to track past variations of the extension of the water masses bathing the ocean floor, thus deep-sea circulation patterns.

From the 1950s on, research on low oxygen basins off California (e.g., Smith, 1964; Phleger & Soutar, 1973; Douglas & Heitman, 1979) and on Mediterranean sapropels (e.g., Parker, 1958), led to the recognition that recent as well as ancient low oxygen environments were inhabited by specific faunas, generally with a low

diversity and strongly dominated by a few species adapted to these apparently hostile environments (e.g., Cita & Podenzani, 1980; Mullineaux & Lohmann, 1981; Parisi & Cita, 1982; Van der Zwaan, 1982). Bernhard (1986) showed that taxa inhabiting low oxygen environments are characterised by a specific morphology, often with a maximum surface-to-volume ratio. As a consequence, bottom water oxygenation became gradually accepted as a major environmental factor in many ecosystems.

The work of Lutze's group on the northwest Africa upwelling area (Lutze, 1980; Lutze & Coulbourn, 1984; Lutze, Pflaumann, & Weinholz, 1986) probably caused the breakthrough that led to the recent opinion that the organic flux to the ocean floor is the most important parameter controlling benthic life in deep, open ocean ecosystems. Since the 1980s, many efforts have been made to develop reliable proxies of bottom water oxygenation and various aspects of the organic flux to the ocean floor.

1.3. Recent Advances in Benthic Foraminiferal Ecology

It is outside the scope of this paper to give a complete overview of our present knowledge of deep-sea foraminiferal ecology (recent reviews in Murray, 1991; Sen Gupta, 1999; Van der Zwaan et al., 1999; Gooday, 2003), but we will highlight some of the most important advances since the 1980s, which have significantly changed our view of the parameters controlling foraminiferal faunas, thus the potential use of fossil faunas as paleoceanographical proxies.

- 1) Since the key papers of Basov and Khusid (1983) and Corliss (1985), and the numerous ecological studies confirming their observations (e.g., Mackensen & Douglas, 1989; Corliss & Emerson, 1990; Corliss, 1991; Barmawidjaja, Jorissen, Puskaric, & Van der Zwaan, 1992; Rathburn & Corliss, 1994; Kitazato, 1994; Rathburn, Corliss, Tappa, & Lohmann, 1996; Ohga & Kitazato, 1997; De Stigter, Jorissen, & Van der Zwaan, 1998; Jorissen, Wittling, Peypouquet, Rabouille, & Relexans, 1998; Kitazato et al., 2000; Schmiedl et al., 2000; Fontanier et al., 2002; Licari, Schumacher, Wenzhöfer, Zabel, & Mackensen, 2003), we know that deep-sea benthic foraminifera do not exclusively live at or on the sediment surface, but are present alive in the upper 1–10 cm of the sediment, in microhabitats that become increasingly oxygen-depleted from the sediment surface downwards. Elaborating ideas proposed by Shirayama (1984), Corliss and Emerson (1990), and Loubere, Gary, and Lagoe (1993), Jorissen, De Stigter, and Widmark (1995) proposed the so-called TROX model, which explains that the depth of the foraminiferal microhabitat in oligotrophic ecosystems is limited by the availability of food particles within the sediment, whereas in eutrophic systems a critical oxygen level decides down to what depth in the sediment most species can live (Figure 1). Some authors have suggested that oxygen concentration is not a major limiting factor for many taxa (e.g., Rathburn & Corliss, 1994; Moodley, Heip, & Middelburg, 1997, 1998a, 1998b), and that competition and predation may interfere (Buzas, Collins, Richardson, & Severin, 1989; Mackensen & Douglas, 1989; Van der Zwaan et al., 1999; Gooday, 2003), but the general validity of the TROX concept has been confirmed in many

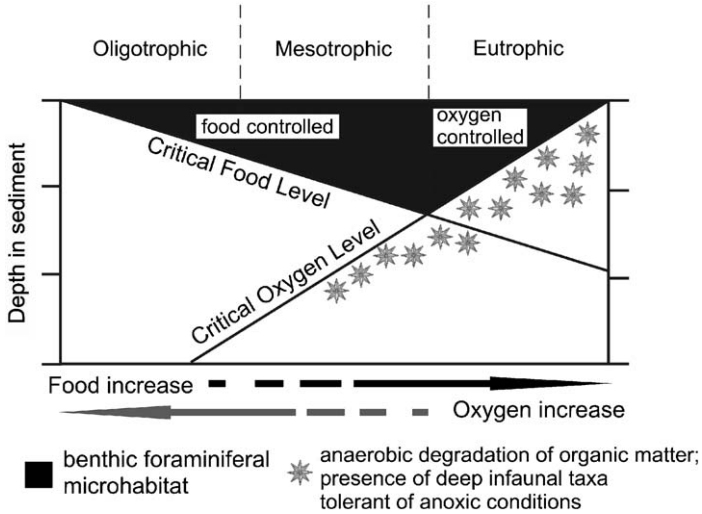


Figure 1 Trox-model (Jorissen et al., 1995, with permission from Springer), explaining the depth limits of the foraminiferal microhabitat by a combination of oxygen penetration and food availability in the sediment. In oligotrophic environments, the microhabitat depth is limited by the low amount of food in the sediment; in eutrophic ecosystems, the penetration depth of most taxa is limited by the shallow level at which no oxygen is present. In mesotrophic areas, microhabitat depth is maximal. Some deep infaunal taxa are not limited by the zero oxygen level, but may participate in anaerobic pathways of organic matter remineralisation at several cm depth in the sediment.

studies, and Carney (2005) suggested that the TROX model may also successfully explain many macrofaunal distribution patterns.

The foraminiferal niche is thus much more variable than thought previously, with different species inhabiting a wide range of biogeochemically different micro-environments, from the sediment–water interface to several cm depth in the sediment. Therefore, the isotopic and trace element composition ($\delta^{18}\text{O}$, $\delta^{13}\text{C}$, Mg/Ca, etc.) of the foraminiferal test can be interpreted adequately only if the microhabitat (and calcification depth) of each investigated species is precisely known. The study of site-specific differences in these parameters opens up new possibilities to reconstruct former redox conditions at and below the sediment–water interface, as well as their controlling parameters.

- 2) Since the early 1990s, Rose Bengal stained foraminifera have been reported to occur in anoxic environments below the sediment–water interface (e.g., Bernhard, 1989; Bernhard & Reimers, 1991; Loubere et al., 1993; Alve, 1994; Rathburn & Corliss, 1994; Rathburn et al., 1996; Jannink, Zachariasse, & Van der Zwaan, 1998; Jorissen et al., 1998; Fontanier et al., 2002). Rose Bengal is a protein stain (Walton, 1952) which has been widely used for the recognition of living foraminifera. Rose Bengal, however, will also stain protoplasm in a more or less advanced state of decay (e.g., Bernhard, 1988; Corliss & Emerson, 1990; Hannah & Rogerson, 1997; Jorissen, 1999a), and thus its reliability for recognition of

living foraminifera has been seriously questioned, especially in anoxic environments where protoplasm decay may be very slow (e.g., Corliss & Emerson, 1990). Studies using more specific vital stains have confirmed that some foraminiferal species can indeed live, and be active, in anoxic sediments (Bernhard & Reimers, 1991; Bernhard, 1993; Bernhard & Alve, 1996). The abundant evidence that populations may be active in low oxygen conditions suggests that oxygen concentration is a critical factor only below a certain threshold, which for most species is surprisingly low, i.e., below 1 ml/l or even less (Jorissen et al., 1995; Bernhard et al., 1997; Van der Zwaan et al., 1999; Levin et al., 2001). This suggests that the quantitative reconstruction of bottom water oxygenation may be feasible only for values below 1 ml/l, and will probably be much more complicated or impossible, at higher concentrations (Murray, 2001).

- 3) Since publication of the key papers of Lutze and co-workers (Lutze, 1980; Lutze & Coulbourn, 1984; Lutze et al., 1986) it has become clear that the flux of organic matter to the deep-sea floor is the main parameter structuring benthic foraminiferal faunas by controlling their density and species composition. Many foraminiferal taxa appear to have an optimum range with respect to organic input, in which their competitiveness is maximal, and under which conditions they become dominant faunal elements. If the organic input falls below, or exceeds the optimal flux levels, they are replaced by taxa that are more competitive under more oligotrophic or more eutrophic conditions. Changes in the bathymetrical range of foraminiferal species under different organic flux regimes were first documented by Pflum and Frerichs (1976) in front of the Mississippi delta, more recently by De Rijk, Jorissen, Rohling, and Troelstra (2000), who showed a progressive shoaling of the bathymetrical ranges of many species along a West–East transect through the Mediterranean, coincident with a transition from eutrophic to very oligotrophic ecosystems.

The fact that many foraminiferal species depend strongly upon the organic flux must open up new pathways for the reconstruction of paleoproductivity, but as we will explain in Section 2.3, the development of such methods is hampered by calibration problems (Altenbach et al., 1999).

- 4) Carney (1989) proposed that two independent organic matter remineralisation cycles exist at the sea floor, fueled by two types of organic input. Labile organic matter (marine organic matter, phytodetritus), is rapidly remineralised at the sediment–water interface using aerobic pathways, whereas more refractory organic matter (terrestrial organic matter, laterally advected material) is remineralised much more slowly in the dysaerobic ecosystems deeper in the sediment. Since this key publication, several authors have speculated on the importance of organic matter quality as a controlling ecological parameter. Most superficially living taxa probably participate in aerobic remineralisation, whereas deeper living taxa may contribute to dysaerobic mineralization of lower quality organic matter (Fontanier et al., 2002, 2005). In the eastern Atlantic Cap Ferret Canyon, for instance, where there is lateral input of large quantities of refractory organic matter, a rich deep infauna co-exists with much poorer surface faunas (Fontanier et al., 2005).

These observations suggest that it should be possible to use the benthic foraminiferal faunal composition to reconstruct not only the quantity, but also the quality of the organic input. On the basis of the succession of fossil faunas in a core off NW Africa, Caralp (1984, 1989) suggested that *Bulimina exilis* has the advantage over other species when there is an input of fresh, labile organic matter with a high phaeopigment/glucide ratio, whereas *Melonis barleeanus* was more abundant when the organic input consisted of more refractory, laterally advected organic material. Goldstein and Corliss (1994) observed that diatom frustules were ingested by *U. peregrina*, but not by *Globobulimina* spp., suggesting adaptation to different food types. On the basis of fatty acid analysis in foraminiferal protoplasm, Suhr, Pond, Gooday, and Smith (2003) concluded that some taxa are more selective than others. For instance, *Globocassidulina subglobosa* may preferentially ingest fresh diatoms, whereas *Thurammina albicans* feeds on degraded material. The shallow infaunal taxa *Uvigerina akitaensis* and *Bulimina aculeata* show a higher carbon assimilation rate than the deeper infaunal species *Textularia kattergatensis* and *Chilostomella ovoidea* in a field feeding experiment with ^{13}C -labeled algae (Nomaki, Heinz, Hemleben, & Kitazato, 2005). All these observations suggest that different species respond differently to the input of various types of food particles, and that food preferences exist.

- 5) From long-term observations based on sediment trap deployments, we know that in most areas the organic matter flux to the ocean floor strongly varies seasonally and/or interannually (e.g., Billett, Lampitt, Rice, & Mantoura, 1983; Berger & Wefer, 1990; Lohrenz et al., 1992). In some areas, episodic events are responsible for most of the organic input to the seafloor, and specialised benthic foraminiferal faunas may rapidly colonise phytodetritus deposits, reacting by accelerated growth and/or reproduction (e.g., Gooday, 1988, 1993, 1996; Gooday & Lamshead, 1989; Gooday & Rathburn, 1999; Gooday & Hughes, 2002). These phenomena have been documented most conclusively after episodic phytodetritus falls in otherwise oligotrophic deep oceanic basins, but also may control foraminiferal population dynamics in more eutrophic areas with a less variable organic input (e.g., Ohga & Kitazato, 1997; Kitazato et al., 2000; Rathburn et al., 2001; Fontanier, Jorissen, Anschutz, & Chaillou, 2003, 2006). This strong response of some foraminiferal taxa (e.g., *Epistominella exigua*) has led to attempts to use these taxa as indicators of pulsed organic input (Loubere, 1998; Smart, King, Gooday, Murray, & Thomas, 1994; Thomas, Booth, Maslin, & Shackleton, 1995; Thomas & Gooday, 1996).
- 6) Since the early publications of Buzas and co-workers (Buzas & Gibson, 1969; Buzas et al., 1989), it has become increasingly clear that spatial variability at a micro- or mesoscale (patchiness) may be important in the deep ocean (e.g., Gooday & Rathburn, 1999; Hughes & Gooday, 2004). This phenomenon may cause large differences between living faunas collected with multi-corers on one hand, and fossil samples in which patchiness has been eliminated by time averaging on the other hand. Taphonomical processes may be responsible for a partial or even total loss of many foraminiferal taxa and will further increase the differences between fossil and living faunas. Ecological observations of living

faunas are essential for a better understanding of proxy relationships, but we think that a successful calibration of a proxy relation is very difficult, if not impossible, to establish on the basis of living faunas only, because of the above mentioned processes which create large differences between living and fossil faunas. We argue that once the underlying mechanisms of proxy relations are fully understood, proxy calibration should be performed on the basis of sub-recent faunas in which the transformation of the living fauna to the fossil fauna due to early diagenetic processes has been concluded.

2. BENTHIC FORAMINIFERAL PROXIES: A STATE OF THE ART

2.1. Overview of Proxy Methods Based on Benthic Foraminiferal Assemblage Data

In this chapter, we will describe the present state of the art of the most commonly used proxy methods based on benthic foraminiferal assemblage data. Benthic foraminiferal assemblage data may be of several types: presence/absence data of various taxa, measures of faunal density, of biodiversity, or data on the morphology of dominant taxa. All of these have been proposed as paleoceanographical proxies. The reconstructed environmental parameters fall into two groups:

- a) Physico-chemical parameters, such as temperature, salinity, carbonate saturation, hydrodynamics, or oxygen concentration of the bottom water. Such parameters may be expected to act as limiting factors, determining whether a foraminiferal species can live somewhere, and if so, if it can feed actively, grow, calcify and reproduce. If all limiting factors fall within the optimum range for a specific taxon, it will attain maximum competitiveness, and may be expected to reach its maximum abundance. Such a maximum abundance may be very high for opportunistic species, but very low for highly specialised K-selected taxa (e.g., Levinton, 1970; Pianka, 1970; Dodd & Stanton, 1990).
- b) Resource parameters, such as the quality and quantity of the organic matter flux that directly influence the amount of food available to specific taxa. Resources are expected to act directly on the density of foraminiferal populations, but will also determine what species will dominate the faunas, because most taxa have maximal competitiveness within a specific flux range.

Traditionally, benthic foraminiferal assemblage data have been used to reconstruct water depth. In the deep-sea, water depth itself is not a controlling ecological parameter, but most other parameters change with water depth, thus causing the well-established bathymetrical successions seen in many ocean basins. Although elaborate bathymetrical distribution schemes have been proposed from the 1950s on, more recent data consistently show important differences in bathymetrical ranges between ocean basins, which appear to be at least partially caused by differences in organic flux regime, so that reliable reconstruction of paleo waterdepth by the use of benthic foraminiferal presence-absence patterns is probably not possible. Alternative methods, such as the ratio between planktonic

and benthic foraminifera (e.g., Van der Zwaan, Jorissen, & De Stigter, 1990; Van Hinsbergen, Kouwenhoven, & Van der Zwaan, 2005), or modern analogue techniques (Hayward, 2004) continue to be explored.

Foraminiferal assemblage characteristics have rarely been used for the reconstruction of bottom water temperature and salinity in open ocean environments, because the variability in these parameters in most oceanic basins is probably too limited to cause a significant faunal response. In shallow and coastal water environments, on the contrary, where strong temperature and salinity gradients exist, foraminifera have been used successfully to reconstruct these parameters. The combination of stable oxygen isotope and Mg/Ca values is considered the most promising method to reconstruct temperature and salinity in deep oceanic environments (e.g., Lear, Elderfield, & Wilson, 2000; Lear, Rosenthal, Coxall, & Wilson, 2004).

The combination of bottom water temperature and salinity determines the specific density of sea water, and the stratification of the oceanic water masses (e.g., Schmitz, 1992). Until the early 1980s, the benthic foraminiferal assemblage composition was thought to mainly reflect these water mass characteristics, and it was thought possible to reconstruct the geographical and depth extent of specific water masses on the basis of the distributional patterns of benthic foraminiferal marker species (e.g., Streeter, 1973; Schnitker, 1974, 1979, 1980; Gofas, 1978; Lohmann, 1978b; Lutze, 1979; Streeter & Shackleton, 1979; Corliss, 1979b, 1983a; Peterson, 1984; Caralp, 1987). With the realisation that the spatial and bathymetrical distribution of most recent foraminiferal taxa is predominantly based on organic flux rates, many scientists today doubt the validity of the so-called “water mass concept”. Although small changes in temperature and salinity in deep ocean ecosystems appear to have a minimal influence on benthic foraminiferal assemblages, other parameters related to water masses, such as carbonate saturation (alkalinity) may have a profound influence on bio-calcification, and in this way structure the live benthic foraminiferal assemblages (e.g., Mackensen, Grobe, Kuhn, & Fütterer, 1990; Mackensen, Schmiedl, Harloff, & Giese, 1995), and the preservation of their tests. Carbonate saturation becomes especially important close to the carbonate compensation depth (CCD) (e.g., Bremer & Lohmann, 1982).

The reconstruction of the extent of former water masses is still of paramount importance, and today a range of geochemical proxies, such as Cd/Ca ratios in calcite, Nd isotopes in fish teeth and manganese nodules, Pb isotopes in manganese nodules, or carbon isotopes in carbonates are used (e.g., Lynch-Stieglitz, 2004), and the foraminifera-based “water mass” concept can not be discarded without a serious discussion (Section 2.4).

The organic flux to the ocean floor, its quantity, quality and periodicity, is today considered to be the prevailing environmental parameter structuring deep-sea benthic foraminiferal faunas, so that our best hopes to develop paleoceanographic proxies on the basis of deep-sea benthic foraminiferal assemblages lay in the field of paleo-productivity. We will treat this subject in Section 2.3, where we will also focus on proxies reconstructing the periodicity of the organic input, and the presence/absence of episodic events.

Although many modern studies suggest that bottom water oxygen concentration is for most species less critical than thought previously, several foraminiferal proxies of bottom water oxygenation have been proposed, and yield promising

results. In Section 2.2 we will discuss the existing methods, and indicate possible pathways for future improvement.

The observation that a specific assemblage of benthic foraminifera occurs in areas with elevated current velocities (Schönfeld, 1997, 2002a, 2002b) has led to the tentative development of a proxy for bottom current velocity. This promising new field of proxy development could inform us about past variations of the intensity of deep-water circulation and is briefly treated in Section 2.5.

2.2. Proxies of Bottom Water Oxygenation

2.2.1. Introduction

Since the 1960s, abundant live foraminiferal faunas have been reported from low oxygen environments (e.g., Smith, 1964; Phleger & Soutar, 1973; Douglas & Heitman, 1979). Recent review papers (e.g., Bernhard, 1986; Sen Gupta & Machain-Castillo, 1993; Bernhard & Sen Gupta, 1999) agree that faunas from such environments have a characteristic taxonomic composition, and are generally strongly dominated by bolivinids, buliminids, globobuliminids and some other taxa, at least when oxygen concentrations reach concentrations below ~ 1 ml/l. If the relative proportion of these taxa would increase with decreasing bottom water oxygenation, it should be possible to use the composition of fossil faunas to reconstruct ancient bottom water oxygen concentrations. It has been argued, however, that two groups of fundamentally different taxa, with very different life strategies, may profit from the disappearance of less resistant taxa at low bottom water oxygen concentrations: (1) deep infaunal taxa, which at the onset of bottom water hypoxia migrate from the deeper sediment layers to the sediment surface (Jorissen, 1999a), and (2) epifaunal or shallow infaunal taxa which have developed adaptations to or tolerance for low oxygen conditions. Unlike the first group, the second group of taxa are usually rare in environments with well-oxygenated bottom waters, to become abundant under severely hypoxic conditions, at least in some locations.

Before looking at the available proxy methods for paleo bottom water oxygenation based on foraminiferal assemblages, we must discuss the terminology used to describe environments where bottom and/or pore waters are under-saturated in oxygen. Unfortunately, there is considerable confusion in the literature concerning the exact meaning of the various descriptors. Figure 2 presents an inventory of the terms most often encountered in the foraminiferal literature, and their meaning according to various authors. There is considerable confusion about the exact meaning of the terms “dysoxic” and “suboxic”, which have been used for environments with entirely different oxygen concentrations. The apparently precise term anoxic (a = without) is used for environments where oxygen concentrations are below the detection limit (dependent upon techniques used), whereas others use it for concentrations below 0.1 ml/l. Bernhard and Sen Gupta (1999) differentiate between anoxic environments with and without sulphate reduction, the latter being termed postoxic. In order to avoid further confusion, we will use the term hypoxic for all environments (without giving a precise range of oxygen concentration) where foraminiferal faunas may potentially be influenced by low oxygen

| | Tyson and Pearson, 1991 | Bernhard and Sen Gupta, 1999 | Kaiho, 1994 | |
|----------|-------------------------|------------------------------|-------------|----------|
| 8 ml/l | Oxic | Oxic | High oxic | 3 ml/l |
| | | | Low oxic | |
| 2 ml/l | Dysoxic | 1 ml/l | Suboxic | 1.5 ml/l |
| | | Dysoxic | Dysoxic | 0.3 ml/l |
| 0.2 ml/l | Suboxic | | | |
| 0 ml/l | Anoxic | Microoxic | Anoxic | 0.1 ml/l |
| 0 ml/l | | Anoxic/Postoxic | | |

Figure 2 Classification of environments with different oxygen concentrations, according to Tyson and Pearson (1991), Kaiho (1994), and Bernhard and Sen Gupta (1999).

concentrations, and we will use anoxic for all environments without detectable oxygen.

2.2.2. Potential problems for paleo-oxygenation proxies

Several problems must be considered when benthic foraminiferal assemblages are used to reconstruct bottom water oxygen concentration, some of which (points 1, 2, 4, and 5 in the following list) are specific for bottom water oxygenation proxies, whereas others are of a more general nature, and concern most proxies based on microfossil remains:

1. In most benthic ecosystems oxygen concentration is subject to a strong gradient. Starting in the more or less well-oxygenated bottom waters, oxygen concentrations decrease first at the sediment-water interface, followed by a rapid downward decrease in the superficial sediment layer, to become zero at a few mm or cm depth. In continental slope and margin environments the depth of the oxic sediment layer rarely exceeds 5 cm. Extreme oxygen penetration, down to 1 m or more, has been observed in the Angola Basin, Weddell Sea and in the West Equatorial Pacific (Rutgers van der Loeff, 1990). In contrast, oxygen penetration is limited to the topmost mm of the sediment in many of the world's oxygen minimum zones (OMZs) (e.g., Miao & Thunell, 1993; Levin, 2003).

In environments where oxygen concentrations change considerably over short vertical distances within the sediment and/or over short time periods, it must be made very clear what the bottom water oxygenation proxy exactly intends to reconstruct: bottom water oxygenation (at some tens of centimeter above the sediment-water interface), oxygen concentration at the sediment-water interface,

or the depth of oxygen penetration into the sediment. These three elements each tell part of the story, and only the knowledge of all three (which is probably unrealistic to expect from proxies) gives a complete picture of benthic ecosystem oxygenation.

2. Benthic foraminiferal faunas commonly inhabit a several cm deep superficial sediment layer, coinciding with the complete range from well oxygenated to strongly hypoxic conditions. Therefore, most faunas collected in environments with well-oxygenated bottom waters contain a mixture of taxa that inhabit fully oxic to strongly hypoxic, or even anoxic microhabitats. Although individual species usually have a preference for a specific depth interval (e.g., Corliss, 1985; Jorissen et al., 1998; Fontanier et al., 2002), even taxa considered “epifaunal” do not exclusively live at the sediment-water interface, but may migrate through the topmost mm or cm of the sediment, and will, as all other taxa, experience a wide range of oxygen concentrations during their life time (e.g., Barmawidjaja et al., 1992; Ohga & Kitazato, 1997).
3. Geological samples, the basis for paleoceanographic reconstructions, are always time-averaged. In the ideal case (without addition or loss of tests due to lateral transport), a sample will contain a mixture of all faunas that inhabited the site during several decennia. Even in laminated sediments with annual laminae, the fossil fauna found in a single lamina will represent the average conditions during a complete season, i.e., ~6 months. If environmental conditions were stable during the period under consideration, the composition of the fauna may indeed record bottom water oxygenation precisely. However, bottom water oxygen concentration (and oxygen penetration depth into the sediment) will very often experience important short-term variability, for instance when strong water column stratification, or pulsed phytodetritus deposits, cause seasonal hypoxia/anoxia. Such events may be annual, but can also be highly episodic, occurring once every 10 years, or even less often. In such cases it will be extremely difficult (if not impossible) to extract the precise oxygenation history from the time-averaged faunas. For instance, in cases of anoxic conditions (without fauna) interrupted by short periods with oxic bottom waters, characterised by rich pioneer faunas, a foraminifer-based bottom water oxygenation proxy will probably overestimate the average long-term bottom water oxygen concentration. In other settings the duration and severity of the periodical hypoxia/anoxia may easily be overestimated, for instance when oligotrophic, well-oxygenated ecosystems are affected by short-term anoxia leading to the explosive development of a few low-oxygen tolerant taxa.
4. The critical values at which the oxygen concentration starts to have a negative impact on the organisms are probably very low, below 1 ml/l or even below 0.5 ml/l (Levin & Gage, 1998). Many elongated, commonly biserial or triserial taxa, have been described as abundant at very low oxygen concentrations. In several papers (e.g., Bernhard, 1986; Corliss & Chen, 1988; Corliss & Fois, 1990; Corliss, 1991) it has been suggested that their morphology corresponds to an infaunal microhabitat. Not only elongated uniserial, biserial and triserial taxa, however, but a wide range of planoconvex to biconvex,

planispiral and trochospiral taxa have been found alive in low oxygen environments (Table 1). For example, *Cibicidoides wuellerstorfi*, usually described as a strictly epifaunal taxon and considered typical of well oxygenated bottom waters, is present in significant numbers in several stations in the Sulu Sea where bottom water oxygen concentrations are well below 2 ml/l (Rathburn & Corliss, 1994). Many of the planoconvex to biconvex taxa listed in Table 1 (e.g., *Cassidulina carinata*, *Cibicides ungerianus*, *Cibicidoides wuellerstorfi*, *Gavelinopsis*

Table 1 Observations of Planoconvex Planispiral and Trochospiral Taxa Generally Considered as Typical of Well-Oxygenated Bottom Waters in Low Oxygen Environments; the Second Column Gives the Lowest Bottom Water Oxygen Concentration at Which Living (Rose Bengal stained) Specimens have been Described; and the Third Column Gives the Reference: 1. Alve (1990); 2. Bernhard (1992); 3. Douglas and Heitman (1979); 4. Jannink et al. (1998); 5. Jorissen et al. (1992); 6. Maas (2000); 7. Mackensen and Douglas (1989); 8. Sen Gupta et al. (1997); 9. Smith (1964).

| | | | | | |
|-----------------------------------|----------|-----|----------------------------------|----------|---|
| <i>Ammonia batavus</i> | 0.3 ml/l | 1 | <i>Epistominella smithi</i> | 0.1 ml/l | 3 |
| <i>Anomalina</i> sp. | 0.2 ml/l | 4 | <i>Epistominella vitrea</i> | 0.8 ml/l | 5 |
| <i>Cancris auriculus</i> | 0.3 ml/l | 6 | <i>Eponides antillarum</i> | 0.9 ml/l | 9 |
| <i>Cancris inaequalis</i> | 0.2 ml/l | 7 | <i>Eponides leviculus</i> | 0.2 ml/l | 3 |
| <i>Cancris oblongus</i> | 0.2 ml/l | 4 | <i>Eponides regularis</i> | 0.1 ml/l | 6 |
| <i>Cancris panamensis</i> | 0.6 ml/l | 9 | <i>Gavelinopsis lobatulus</i> | 2.1 ml/l | 4 |
| <i>Cancris sagra</i> | 0.4 ml/l | 9 | <i>Gavelinopsis translucens</i> | 0.0 ml/l | 8 |
| <i>Cassidulina carinata</i> | 0.3 ml/l | 4 | <i>Gyroidina io</i> | 0.2 ml/l | 3 |
| <i>Cassidulina crassa</i> | 0.3 ml/l | 6 | <i>Gyroidina lamarckiana</i> | 0.1 ml/l | 6 |
| <i>Cassidulina cushmani</i> | 0.3 ml/l | 9 | <i>Gyroidina multilocula</i> | 0.4 ml/l | 9 |
| <i>Cassidulina delicata</i> | 0.1 ml/l | 3,7 | <i>Gyroidina parva/pulchra</i> | 0.9 ml/l | 4 |
| <i>Cassidulina depressa</i> | 0.5 ml/l | 4 | <i>Gyroidina rotundimargo</i> | 0.3 ml/l | 9 |
| <i>Cassidulina laevigata</i> | 0.2 ml/l | 7 | <i>Gyroidina umbonata</i> | 0.4 ml/l | 4 |
| <i>Cassidulina limbata</i> | 1.0 ml/l | 3 | <i>Gyroidinoides neosoldanii</i> | 0.5 ml/l | 2 |
| <i>Cassidulina minuta</i> | 0.6 ml/l | 9 | <i>Gyroidinoides soldanii</i> | 0.1 ml/l | 6 |
| <i>Cassidulina oblonga</i> | 0.6 ml/l | 9 | <i>Hanzawaia boueana</i> | 0.8 ml/l | 5 |
| <i>Cassidulina sgarellae</i> | 0.3 ml/l | 6 | <i>Hanzawaia concentrica</i> | 0.3 ml/l | 6 |
| <i>Cassidulina subcarinata</i> | 0.2 ml/l | 3 | <i>Hoeglundina elegans</i> | 0.2 ml/l | 3 |
| <i>Cassidulina subglobosa</i> | 0.2 ml/l | 4 | <i>Hyalinea balthica</i> | 0.3 ml/l | 4 |
| <i>Cassidulina teretis</i> | 0.5 ml/l | 3 | <i>Islandiella subglobosa</i> | 0.2 ml/l | 3 |
| <i>Cassidulina tumida</i> | 0.4 ml/l | 9 | <i>Islandiella</i> sp. | 0.3 ml/l | 6 |
| <i>Cibicides bradyi</i> | 0.3 ml/l | 4 | <i>Lenticulina articulata</i> | 0.3 ml/l | 6 |
| <i>Cibicides fletcheri</i> | 0.4 ml/l | 7 | <i>Oridorsalis umbonatus</i> | 0.4 ml/l | 4 |
| <i>Cibicides refulgens</i> | 0.3 ml/l | 2 | <i>Osangularia culter</i> | 0.3 ml/l | 4 |
| <i>Cibicides ungerianus</i> | 2.1 ml/l | 4 | <i>Osangularia rugosa</i> | 0.0 ml/l | 8 |
| <i>Cibicidoides wuellerstorfi</i> | 1.6 ml/l | 2 | <i>Planulina ariminensis</i> | 0.2 ml/l | 7 |
| <i>Elphidium albumbilicatum</i> | 0.0 ml/l | 1 | <i>Planulina exorna</i> | 0.6 ml/l | 9 |
| <i>Elphidium excavatum</i> | 0.1 ml/l | 1 | <i>Planulina limbata/ornata</i> | 0.3 ml/l | 9 |
| <i>Elphidium incertum</i> | 0.1 ml/l | 1 | <i>Pullenia</i> sp. | 0.4 ml/l | 4 |
| <i>Elphidium tumidum</i> | 2.4 ml/l | 9 | <i>Valvulineria araucana</i> | 0.2 ml/l | 3 |
| <i>Epistominella decorata</i> | 0.1 ml/l | 6 | <i>Valvulineria javana</i> | 0.2 ml/l | 4 |
| <i>Epistominella exigua</i> | 0.3 ml/l | 6 | <i>Valvulineria oblonga</i> | 0.3 ml/l | 9 |

- translucens*, *Hoeglundina elegans*), have been found as shallow infauna or epifauna in well-oxygenated environments. In some papers (e.g., Kaiho, 1994; Jannink et al., 2001) it was therefore implicitly suggested that these species need elevated oxygen concentrations, but observations of these taxa in strongly hypoxic environments suggest that they are much less sensitive to low oxygen concentrations than generally assumed. Their preference for superficial sediment niches may well be explained by their dependence on a supply of high quality food particles. It should also be realised that no foraminiferal species occur exclusively in low-oxygen environments (Sen Gupta & Machain-Castillo, 1993), and that most taxa indicative for strongly hypoxic conditions can also be found at much higher bottom water oxygenation values. Murray (2001) thus suggested that foraminifera can only be used as proxies for oxygen levels below concentrations of 1 or perhaps 2 ml/l, and above these values, there is no relation between oxygen levels and the composition of the foraminiferal faunas (Murray, 2001).
5. In recent benthic ecosystems there is a complex interplay between export productivity (organic flux to the sea floor) and oxygenation of bottom and pore waters. The competitive ability of most species appears to be determined by one or both of these parameters. Jorissen et al. (1995) suggested that deep infaunal taxa, often considered indicative of hypoxic conditions, are only present when the organic flux is sufficiently high to have organic detritus within the sediment that can be used metabolically. In areas with lower flux rates most organic matter is consumed at the sediment-water interface, and deeper sediment layers are too poor in organic matter to be inhabited by deep infaunal taxa (Figure 1). This means that a sudden appearance, or a relative frequency increase, of these deep infaunal taxa in the fossil record may very well be the result of an increased organic flux, and not of lowered bottom water oxygen concentrations. Oxygen concentrations have a direct impact on the faunal composition only in areas where bottom water concentrations become so low that they cause a diminished competitive ability and/or reproductive potential of the more superficially living taxa. In such cases, less resistant taxa will disappear, and deep infaunal taxa that are perfectly adapted to live in low oxygen environments will take over niches at the sediment surface and dominate the foraminiferal assemblages (Jorissen, 1999b). Such conditions will occur only when the oxygen concentrations fall below a threshold level critical for the surface dwelling taxa (≤ 1 ml/l; Murray, 2001). Finally, even faunas that are strongly dominated by deep infaunal taxa may form in environments with well-oxygenated bottom waters. For instance, at a 2,800 m deep station in Cap Ferret Canyon (north-eastern Atlantic) bottom waters are well oxygenated (5 ml/l), but deep infaunal taxa are dominant, probably because of intensive degradation of large quantities of low quality organic matter by anaerobic bacterial stocks (Fontanier et al., 2005).
 6. The living fauna always differs considerably from the fossil faunas accumulating at the same site (e.g., Murray, 1991; Jorissen & Wittling, 1999; Walker & Goldstein, 1999). This difference is caused by early diagenetic processes, causing a preferential loss of the more fragile taxa, but also by biological parameters, such

as large interspecific differences in test production, or selective predation on some foraminiferal species (e.g., [Culver & Lipps, 2003](#)). Additionally, transport by bottom currents may be responsible for the addition of allochthonous foraminifera and/or the removal of small autochthonous elements. Since paleoceanographic reconstructions are based on fossil material that has been seriously modified by taphonomic processes, it is probably not a good practice to use living faunas for the calibration of paleoceanographical proxies. Individuals living at any specific moment of sample collection do not necessarily reflect a fauna as averaged over 1 year or a full seasonal cycle, for instance. We think that proxy relationships between faunal characteristics and oceanographical parameters should rather be calibrated by a comparison of the recent values of the target parameter with “subfossil” faunas that represent recent environmental conditions but have fully concluded the suit of diagenetic processes and will undergo no more post-mortem compositional changes. Unfortunately, in most ocean basins, it takes several thousands of years before a fauna has passed through the biologically active top part of the sediment, where most of the diagenetic processes and losses of foraminiferal tests take place (e.g., [Walker & Goldstein, 1999](#)). It is very improbable that the Holocene fossil faunas collected below this topmost sediment layer where early diagenesis is taking place, can directly be compared with the recent (anthropogenically-influenced) environmental conditions, a procedure that cannot be avoided when a proxy relationship has to be calibrated. These problems can be at least partly solved by a careful, high-resolution study of a succession of Holocene faunas in areas with high sedimentation rates.

7. Most oceanic environments are much less stable than previously thought: even abyssal environments experience episodic phytodetritus inputs, provoking a strong response of some highly opportunistic taxa that may rapidly attain high densities (e.g., [Gooday, 1988, 1993](#); [Gooday & Lambshead, 1989](#)). Other unpredictable events such as whale falls or pulsed food input from hydrothermal vents may also have a profound impact on benthic foraminiferal faunas, although hydrothermal vent regions and cold seep areas do not contain endemic benthic assemblages (e.g., [Rathburn et al., 2003](#); [Barberi & Panieri, 2004](#)). At present, it cannot be excluded that in many parts of the ocean an important part of the fossil faunas is formed during ephemeral events. If true, the fossil fauna should not be considered representative of average conditions, but for the very specific conditions associated with the short periods of high productivity.

In spite of these problems, there appear to be significant differences in tolerance for low oxygen levels between species, and therefore oxygen concentration must have an impact on the assemblage composition, at least at low to very low oxygenation levels. The possibility to estimate former oxygen levels with a high precision in the 0–1 ml/l range is important, because it will open up the possibility to reconstruct the history (including extent and severity) of OMZs. These continental margin environments potentially play an important role as organic carbon sinks in the global carbon cycle, and the reconstruction of the variability of intensity of OMZs through time may give insight into the dynamics of intermediate and deeper water masses.

2.2.3. Existing proxies of bottom water oxygenation

The proposed methods for the reconstruction of ancient bottom water oxygenation concentrations on the basis of benthic foraminiferal assemblage characteristics fall into four categories:

1. Most often, a number of taxa is considered indicative of hypoxic conditions, and the relative frequency of these taxa (expressed as a percentage of the total benthic foraminiferal fauna) is used as a semi-quantitative index of bottom water oxygenation. The best example of this method is the widely applied benthic foraminiferal oxygen index (BFOI) of Kaiho (1991, 1994, 1999). In this index, a subdivision of taxa into categories of dysoxic, suboxic or oxic indicators is made rather arbitrarily, mainly based on test morphology, and differs between the 1991 paper that deals with Paleogene faunas, and the 1994 and 1999 papers that try to calibrate the method on the basis of recent faunas. It is surprising to see that for many species adult individuals ($\geq 350 \mu\text{m}$) are considered as “oxic” indicators, whereas smaller individuals ($< 350 \mu\text{m}$) of the same species are considered as “suboxic” indicators (Kaiho, 1994, 1999). The hypothesis underlying the method is that in well-oxygenated bottom waters, dysoxic indicators live in poorly oxygenated deep infaunal microhabitats. In the case of hypoxic conditions at the sediment-water interface, less resistant taxa disappear and the “dysoxic indicators” become dominant. However, only a few of these “dysoxic” indicators (21 taxa listed by Kaiho, 1994) have actually been observed alive in intermediate or deep infaunal microhabitats, and the suggested microhabitat separation between oxic and suboxic indicators (Kaiho, 1994, Figure 1) is not supported by data. In its latest version (Kaiho, 1999), Kaiho’s oxygen index distinguishes between five classes of bottom water oxygenation (Figure 1), and at levels above 3.2 ml/l is no longer correlated with bottom water oxygenation. Other authors have slightly modified the attribution of taxa to the categories of dysoxic, suboxic or oxic indicators (e.g., Baas, Schönfeld, & Zahn, 1998), in order to better fit the method (and results!) to their ideas.
2. The method proposed by Jannink et al. (2001) follows an inverse approach: taxa that are found consistently living in the topmost sediment are considered oxiphylic, and their cumulative percentage is considered a proxy for bottom water oxygenation. The rationale behind this procedure is that oxygen penetration into the sediment increases with increasing bottom water oxygenation, leading to an increased volume of the niche potentially occupied by oxiphylic taxa. The problem with this method is the determination of which species are oxiphylic. A species found living close to the sediment-water interface may prefer this microhabitat because it does not tolerate the lower oxygen concentrations deeper in the sediment, but also because it prefers high quality food particles that are most concentrated at the sediment-water interface, as indicated by an increasing amount of data (e.g., Kitazato, Nomaki, Heinz, & Nakatsuka, 2003; Fontanier et al., 2003, 2006; Ernst, Bours, Duijnste, & Van der Zwaan, 2005; Nomaki et al., 2005).
3. Several authors (e.g., Loubere, 1994, 1996; Morigi, Jorissen, Gervais, Guichard, & Borsetti, 2001) have attempted to link sub-recent faunal assemblages to

bottom water oxygenation values by multivariate statistical methods, with some encouraging results for recent faunas. It is very difficult to know, however, whether application of these relationships to fossil material would yield reliable results, because the investigated datasets contain a rather limited array of combinations of oxygen concentration and organic flux (and other environmental parameters). Non-analog conditions thus may not be correctly reconstructed. An additional problem is the fact that in order to be robust, the multivariate statistical methods need many more data (samples) than variables (taxa) (e.g., Tabachnick & Fidell, 1983), and at present the size of the analysed datasets is small in comparison with the number of species.

4. Schmiedl et al. (2003) proposed a method based on a combination of the relative proportion of low-oxygen tolerant marker species and a diversity index, counting each for 50% of the final score of their oxygen index, because all hypoxic environments show a lower biodiversity than well-oxygenated settings. By adding a factor independent of taxonomic composition the proxy method may become more robust, and applicable in an array of areas with different faunal compositions.

Unfortunately, all four procedures suffer from one or more of the pitfalls outlined before. The main problem is the fact that all species simultaneously respond to bottom and pore water oxygenation and to the organic flux level. Many taxa that have been proposed to be low oxygen indicators should probably be considered high productivity markers instead. Although these taxa are indeed abundantly present in many low oxygen environments, they also dominate faunas in high productivity areas with fairly high bottom water oxygen concentrations, such as the upwelling area off NW Africa (e.g., Jorissen et al., 1995; Morigi et al., 2001). The reverse also may occur: benthic environments with low oxygen concentrations are not necessarily dominated by “low oxygen indicators” in areas where export production is low (e.g., Rathburn & Corliss, 1994).

2.2.4. Future developments of bottom water oxygenation proxies

Some of the best examples of past changes in bottom water oxygenation have been preserved in Mediterranean sapropels (e.g., Jorissen, 1999b; Schmiedl et al., 2003). A detailed study of the faunal succession at the transition from homogeneous sediments characteristic of oxygenated bottom waters to the laminated sediments associated with anoxic conditions can yield essential information about the tolerance levels of the various species. Rapid faunal changes took place at the onset of the anoxic periods that lasted for several thousands of years (Figure 3). The faunas that lived during the centuries immediately preceding the onset of anoxia (resulting in azoic sediments) have a low diversity and are strongly dominated by taxa (e.g., *Globobulimina* spp., *Chilostomella* spp.), which in recent ecosystems have been described in deep infaunal microhabitats. Surface dwellers adapted to strongly hypoxic conditions, such as those in the Santa Barbara Basin (e.g., *Epistominella smithi*, *Nonionella stella* and *Cassidulina delicata*; Mackensen & Douglas, 1989; Bernhard et al., 1997) were absent in the late Quaternary Mediterranean.

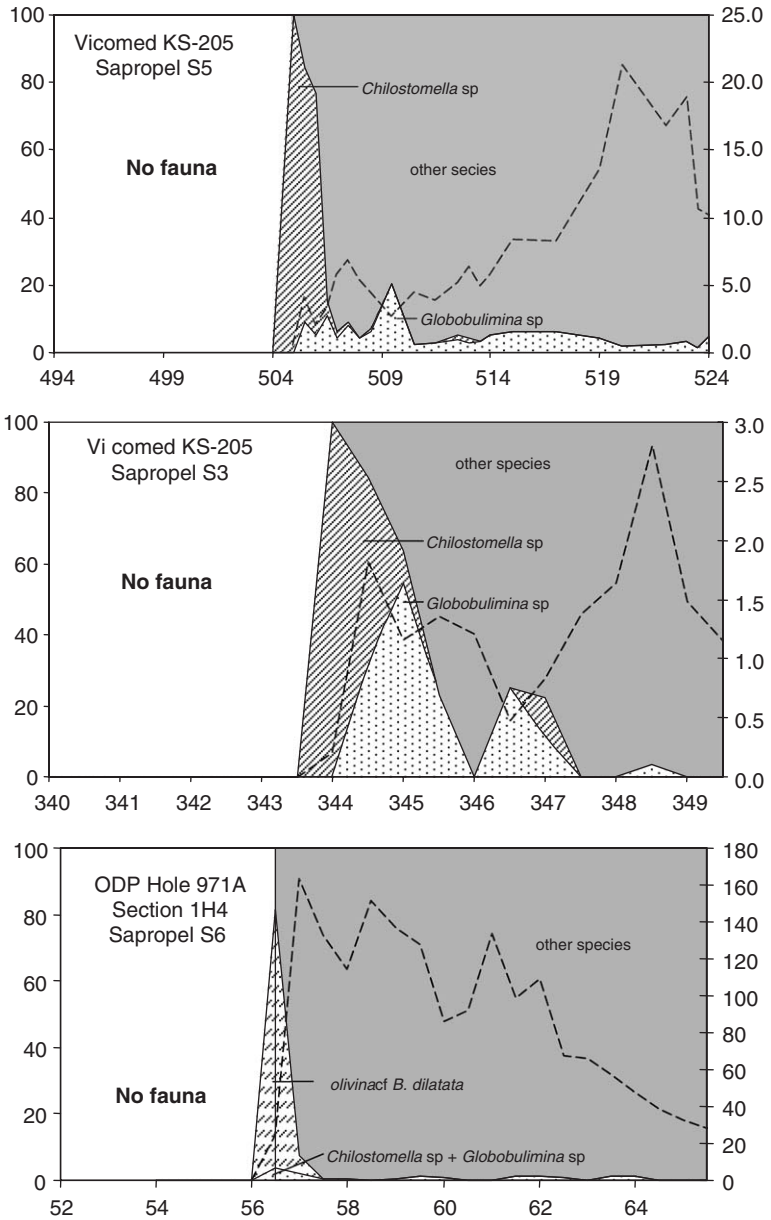


Figure 3 Relative frequency of deep infaunal taxa (percentage, scale on the left), in the centuries before deposition of sapropels S5, S3 (both from core Vicomed KS205, 38°11,86'N, 18°08,04'E, 2,345 m) and S6 (ODP Hole 971A, section 1H4, 33°42,19'N, 24°42,81'E, 2,143 m) and total faunal density (interrupted line, in number of foraminifera per gram dry weight, scale on the right), in function of depth in the core (in centimeter, scale below). Note the abrupt frequency increase to a total dominance in the last centimeter below azoic sediments, which coincides with the disappearance of almost all other taxa. In all three cases this frequency increase is accompanied by a strong drop in faunal density. Sapropele S5 but not S6 shows a preliminary frequency increase (at 510 cm) ~5 cm below this ultimate frequency increase.

Figure 3 shows the relative frequencies of deep infaunal taxa immediately below sapropels S5 and S6. In the rather oligotrophic environments of the central Mediterranean, *Globobulimina* spp., *Chilostomella* spp. and other deep infaunal taxa are rare or even absent at this water depth (De Rijk et al., 2000). These taxa increase dramatically in relative abundance in the 1–2 cm below the azoic sediments (here reflecting a time period of 1–2 centuries), to attain dominance in the last fauna-containing sample (Figure 3). In these examples, the increase in deep infaunal taxa to percentages of 80–100% is accompanied by a strong decrease in the number of foraminiferal tests per gram dry sediment, suggesting that the increase in relative abundance is mainly caused by the disappearance of other taxa, which are less resistant to low oxygen concentrations.

In some but not all sapropels the takeover of the benthic faunas by deep infaunal taxa is preceded by a preliminary increase in relative abundance of these taxa (as seen at 510 cm for sapropel S5, 347 cm for sapropel S3; but not for sapropel S6). This first increase of deep infaunal taxa is not accompanied by a disappearance of less resistant taxa. Two explanations are possible:

- 1) an increased organic flux to the ocean floor may have opened up niches deeper in the sediment for deep infaunal taxa.
- 2) a short-term strong decrease in bottom water oxygenation may have caused mixing of deep infaunal taxa with the more diverse faunas deposited before and/or after this short hypoxic event.

The first frequency increase of deep infaunal taxa at both S3 and S5 is accompanied by a temporary decrease of overall faunal density, so that the second possibility seems most probable.

The curves in Figure 4 present different scenarios during a strong decrease of bottom water oxygenation. In Figure 4a the decrease of bottom water oxygenation is not accompanied by an increase in organic flux, and could occur because of stratification of the water column, or a long-term lack of renewal of deep waters. The percentage of deep infaunal taxa will initially remain stable, until a critical point (indicated by arrow; here arbitrarily placed at 1 ml/l), at which taxa less resistant to hypoxic conditions start to disappear. With a further decrease of bottom water oxygenation, deep infaunal taxa rapidly take over hypoxic niches at the sediment-water interface, and their abundance increases exponentially, reaching 100% at ~ 0.01 ml/l. The initial percentage of deep infaunal taxa (the horizontal lines to the right of the threshold point in Figure 4a) is determined by the background organic flux level. Curve (1) depicts an oligotrophic setting without deep infaunal taxa where only the sediment-water interface is inhabited. Curve (2) characterises more eutrophic settings where the organic flux can sustain a fauna living at several cm depth in the sediment. Curve (1) corresponds to the situation for sapropel S6, ODP hole 971A (Figure 3); curve (2) corresponds to the succession below sapropel S5, core Vicomed KS-205 (Figure 3).

In Figure 4b the decrease in bottom water oxygenation is at least partially accompanied by an increase of the organic flux, as in places where hypoxia is caused by eutrophication. Deep infaunal taxa are then expected to show a first increase in abundance due to an increased food availability within the sediment, but with a

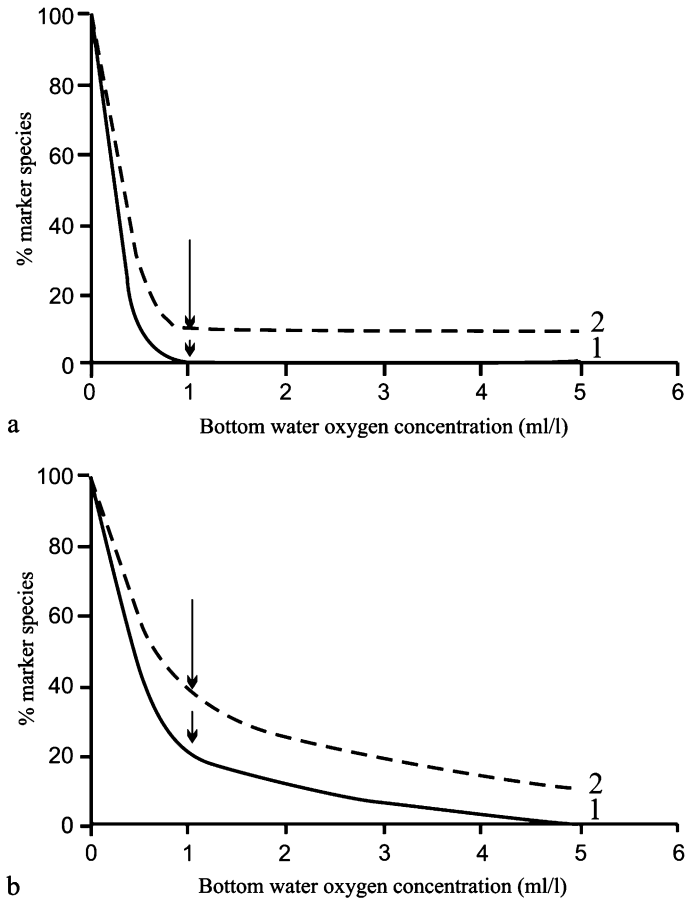


Figure 4 Hypothetical curves showing the percentage of deep infaunal species as a function of bottom water oxygen concentration and organic flux to the ocean floor. (a) The decrease of bottom water oxygenation is not accompanied by an increase in organic flux. (b) The decrease of bottom water oxygenation is accompanied by an increase of organic flux to the ocean floor. (a/b), Curve 1: oligotrophic setting without deep infaunal taxa in the background fauna. Curve 2: More eutrophic setting with ~10% deep infaunal taxa in the background fauna.

further decrease of bottom water oxygenation, less resistant taxa will start to disappear (vertical arrows in Figure 4), and the increase of deep infaunal taxa will strongly accelerate. From this threshold point on the percentage of deep infaunal taxa is no longer determined by the organic flux, but mainly by bottom water oxygenation.

This theoretical example illustrates that the interplay between organic flux and bottom water oxygenation, which controls the percentage of deep infaunal taxa, may be complex. As soon as oxygenation levels fall below a certain threshold value, further increase of the percentage of deep infaunal taxa seems to be largely due to the disappearance of taxa less resistant to low oxygen conditions, and therefore

bottom water oxygenation, and it thus should be possible to use the percentage of deep infaunal taxa to estimate bottom water oxygenation. For this approach to work, it must be determined precisely at what oxygenation level the various less resistant taxa start to disappear from the ecosystem, and the percentage of deep infaunal taxa starts its final increase. In Figure 4, this level has arbitrarily been placed at 1 ml/l, but a different threshold value would not significantly change the shape of the curves. At bottom water oxygenation levels above this critical boundary, on the contrary, the percentage of deep infaunal taxa is determined by a combination of bottom water concentration and organic flux level, and the reconstruction of bottom water oxygenation is more difficult. However, when longer records are available (such as the sapropel records, Figure 3), it may be possible to constrain the organic flux level by other methods (Section 2.3).

Summarising, there is potential to develop a quantitative proxy method for bottom water oxygenation in spite of the complexity of the interplay between bottom water oxygenation and organic matter flux, on the basis of the relative proportions of a group of indicator species that replace taxa less resistant to low oxygen concentrations below a critical threshold value. The calibration of such a proxy has to be based on the rigorous application of the following conditions:

- 1) Indicator species for low oxygen conditions should be selected on the basis of observations in recent low oxygen ecosystems. All deep infaunal taxa (e.g., *Chilostomella* spp., *Globobulimina* spp.) qualify as marker species. Other taxa observed to be abundant in low oxygen settings have a planoconvex morphology (e.g., *Epistominella smithi*, *Nonionella stella*, and *Cassidulina delicata*) that suggests an adaptation to a microhabitat close to the sediment–water interface (e.g., Mackensen & Douglas, 1989; Bernhard et al., 1997).
- 2) In order to take taphonomical processes into account, the proxy method should be calibrated by a comparison between upper Holocene fossil faunas (if existing) and the recent value of the proxy parameter. The calibration should be performed in many different settings (specifically different productivity regimes) with oxygenation values between 0 and 2 ml/l. Above 2 ml/l, benthic foraminiferal assemblage composition is probably not influenced by bottom water oxygenation (Murray, 2001).
- 3) It should be tested whether the addition of a diversity index (Schmiedl et al., 2003) can make the proxy method more robust, and more globally applicable. However, diversity indices may decrease in response to an increase in organic flux as well as to a decrease in oxygen concentration.

If the relation between the percentage of deep infaunal taxa, bottom water oxygen concentration and downward organic flux can be successfully calibrated for a wide range of recent and/or sub-recent environments, a precise proxy of bottom water oxygenation may be obtained for the range of 0–1 ml/l. Potential success of this approach is suggested by Martinez et al. (1999), who documented that changes in the percentage of deep infaunal taxa mirrored changes in values of Mo/Al (Molybdenum/Aluminium), a geochemical oxygenation index. If we succeed in the development of a reliable proxy for bottom water oxygenation, the next challenge will be the reconstruction of the redox conditions within the superficial sediments.

2.3. Paleoproductivity Proxies

2.3.1. Introduction

Knowledge of past changes in primary production and the subsequent transport of organic matter to the ocean floor is essential for the understanding of the response of the biological carbon pump to climate change. Paleoproductivity proxies, which address these complex phenomena, may have several objectives: in some cases they attempt to reconstruct surface water primary production, whereas in other cases a less ambitious, but equally valuable approach is followed, aiming to reconstruct the flux of organic carbon to the ocean floor. A major complication is the fact that most paleoproductivity proxies based on responses to the organic carbon flux to the ocean floor (e.g., sedimentary organic carbon flux, benthic ecosystem response, benthic $\delta^{13}\text{C}$ values) will not only be influenced by the quantity of organic matter arriving at the sea floor, but also by other aspects of the organic flux, such as the quality of the organic matter, and the timing (constant versus pulsed) of the organic input. Another problem is that the mechanisms of organic material transport from the surface waters to the ocean floor are still badly known. If we are able to gain insight into these secondary aspects of paleoproductivity, we will also improve our understanding of the functioning of surface water, pelagic phytoplankton communities and the biological carbon pump in past oceans.

The use of benthic foraminifera as markers of export productivity to the sea floor has become conceivable since the 1970s, when it became progressively clear that not water depth, temperature and salinity (“water masses”), but organic input and bottom water oxygenation are the main environmental parameters controlling faunal patterns in open ocean benthic ecosystems. In the majority of open ocean benthic ecosystems, the flux of organic matter from the productive surface waters to the ocean floor constitutes the main food source for benthic organisms. Although hydrocarbon seeps and hydrothermal vents have received much attention in the past decennia because of their rich and often spectacular faunas, in the present oceans their cumulative surface area is probably small in comparison with sea floor areas, which depend on the input of organic detritus produced in the surface waters. Unfortunately, there is still considerable uncertainty about the processes of transport of particulate organic carbon to the ocean floor. Since the late 1970s, sediment trap data have been used to develop equations describing the downward organic flux through the water column (e.g., Eppley & Peterson, 1979; Suess, 1980; Betzer et al., 1984; Martin, Knauer, Karl, & Broenkow, 1987; Pace, Knauer, Karl, & Martin, 1987; Berger, Smetacek, & Wefer, 1989; Berger & Wefer, 1990). In all these equations, the downward organic flux is described simply as a function of primary production in the surface waters and water depth. More recent data, however, suggest that many other parameters may interfere:

- Several authors (e.g., Eppley & Peterson, 1979; Berger et al., 1989; Berger & Wefer, 1990) have suggested that the organic flux to the ocean floor constitutes a higher percentage of total productivity in unstable settings, such as ecosystems with a high seasonal variability in primary production. Recently, however, François, Honjo, Krishfield, and Manganini (2002) described a decreasing particle flux with higher seasonality.

- The amount of particulate organic carbon transported from the surface to deeper layers seems to depend on the structure and functioning of the pelagic food webs (e.g., Wassmann, 1993). For instance, Boyd and Newton (1995) describe a twofold increase of the primary organic carbon flux as a function of community structure. In general, vertical transport of large particles, such as marine snow (Turley, 2002), diatom aggregates (e.g., Kemp, Baldauf, & Pearce, 1995, 2000) or zooplankton fecal pellets, is much faster and more efficient than transport of small particles.
- Ballasting of organic particles with siliceous or carbonate tests (François et al., 2002), terrigenous dust (Ittekkot, 1993) or tunicate feeding structures (Robinson, Reisenbichler, & Sherlock, 2005) may significantly increase the efficiency of organic matter transport to deeper parts of the water column. Armstrong, Lee, Hedges, Honjo, and Wakeham (2002) suggest that ballast minerals are largely responsible for the deep-water (>1,800 m) POC fluxes.
- Particle organic carbon fluxes through the water column appear to be strongly modulated by biological, physical and chemical transformation processes. Unfortunately, the various mechanisms of POC degradation in the water column are still poorly known (Jackson & Burd, 2002), and the potentially important role of dissolved organic carbon (DOC) is not taken into account in many organic flux studies.
- In many parts of the ocean, and especially in ocean margin settings, lateral advection by intermediate or deep water currents or by slope failure and turbidity currents is a major factor responsible for the transport of particulate organic carbon to the ocean floor (e.g., Antia, Von Bodungen, & Peinert, 1999). Laterally advected organic matter is often aged, with the more labile, easily consumable components stripped off. As a consequence, laterally advected organic matter will be only partially remineralised in the oxic ecosystems at the sediment-water interface. The more refractory parts, however, may trigger intensive early diagenetic processes in the anaerobic ecosystems deeper in the sediment (Carney, 1989; Fontanier et al., 2005).

In spite of these many complicating factors, an important part of the particle rain appears to fall vertically (e.g., Berelson et al., 1997; Nelson et al., 2002), and the POC flux varies as a function of primary production in the surface waters at least down to ~1,000 depth (Fischer, Ratmeyer, & Wefer, 2000). The above-mentioned flux equations could therefore be useful as a first approximation of the quantity of particulate organic matter arriving at the ocean floor.

One of the major problems inhibiting a more precise knowledge of the quantity (and quality) of organic matter arriving at the ocean floor is the difficulty to obtain reliable measurements. Empirical flux formulae are useful, but can only give a rough, long-term estimate of the organic flux to the ocean floor. More precise estimates can be obtained by measuring the amount of organic matter remineralisation in the benthic ecosystem, for instance by *in situ* measurements of oxygen consumption by benthic landers (e.g., Reimers, 1987; Gundersen & Jørgensen, 1990; Epping & Helder, 1998) or in benthic chambers (e.g., Tengberg et al., 1995). Onboard ship measurements in multi-cores are easier to perform, but generally

considered less reliable. In order to have a more complete picture of the extent of organic matter degradation, including mineralisation under anaerobic conditions, vertical profiles of important redox species (e.g., nitrate, manganese, reactivated iron-oxides and sulphate) are measured in the superficial sediment and pore waters (Froelich et al., 1979). Models integrating the downcore concentration profiles of all major redox species provide accurate estimates of the total amount of organic matter remineralised in the benthic ecosystem (e.g., Jahnke, Emerson, & Murray, 1982; Rabouille & Gaillard, 1991; Soetaert, Herman, & Middelburg, 1996; Kelly-Gerreyn, Hydes, & Waniek, 2005).

Although these approaches may provide reliable data about the remineralisation of labile organic matter, the data reflect rather short periods of time (days to weeks), and do not necessarily reflect the long-term, average conditions represented by a fossil benthic foraminiferal fauna, or by any other proxy carrier. Furthermore, these measurements mostly represent the labile, highly reactive part of the organic matter, but not the more refractory components that are degraded in the anaerobic ecosystems deeper in the sediment, over time spans of months to years, or are fossilised, to become sedimentary organic matter.

Problems concerning the quantification of the downward organic flux and the relative importance of laterally advected material do not only complicate reconstructions based on benthic foraminiferal faunas, but concern most paleoproductivity proxies. If we want to apply benthic foraminiferal assemblages successfully as quantitative paleoproductivity proxies we need to understand in detail how recent assemblages are structured by the input of various amounts of organic matter of different quality. In addition, we have to understand how low oxygen concentrations may modify the foraminiferal response to different organic flux regimes.

Several foraminiferal proxies for paleo-export production have been proposed during the last decennia, based on three different approaches:

- 1) Most proxies are based on supposed relations between faunal composition and organic carbon flux rates. In several areas, preferred flux regimes have been tentatively determined for dominant benthic foraminiferal taxa (e.g., Altenbach et al., 1999, 2003; De Rijk et al., 2000; Schönfeld & Altenbach, 2005). For example, *Uvigerina mediterranea* and *U. peregrina* are typical for flux rates above $2.5 \text{ g/m}^2/\text{yr}$, whereas *Cibicides wuellerstorfi*, *Gyroidina altiformis*, and *G. orbicularis* are almost exclusively found below this flux range (Altenbach et al., 1999; de Rijk et al., 2000). However, important inconsistencies appear to exist between studied areas (Schönfeld & Altenbach, 2005), and some important taxa, such as *Epistominella exigua* (Altenbach et al., 1999) or *Melonis* spp. (Altenbach et al., 1999; de Rijk et al., 2000) occur in areas with very different flux regimes, and appear to be not very dependent on a specific organic flux level. These inconsistencies may be due to inadequate estimates of export production, but also to the impact of other ecological factors that may interfere with the foraminiferal dependence on export production. Until today, observed flux regimes for individual taxa have not been used systematically to reconstruct paleo-productivity.
- 2) Multivariate statistical methods have been applied in order to investigate the response of the foraminiferal assemblage composition to different flux regimes

(e.g., Loubere, 1991, 1994, 1996, 1998; Fariduddin & Loubere, 1997; Kuhnt et al., 1999; Loubere & Fariduddin, 1999; Wollenberg & Kuhnt, 2000; Morigi et al., 2001). Loubere (1998) and Loubere and Fariduddin (1999) presented multiple regression functions that relate the foraminiferal assemblage composition quantitatively to primary production in the surface waters. Others (e.g. Kuhnt et al., 1999; Wollenburg, Kuhnt, & Mackensen, 2001) used the statistical relations observed in recent ecosystems to reconstruct paleoproductivity on the basis of fossil faunas.

- 3) Herguera and Berger (1991) and Herguera (1992) proposed a paleoproductivity proxy based on the accumulation rate of the benthic foraminiferal fossil fauna larger than 150 μm . Their data from the western equatorial Pacific show that the benthic foraminiferal accumulation rate (BFAR) varied linearly with export production for stations with primary production ranging from 40 to 135 $\text{g}/\text{m}^2/\text{yr}$, so that for every mg of organic carbon reaching the ocean floor, one foraminiferal shell larger than 150 μm is deposited. The striking simplicity of this idea is appealing but also discouraging, because somehow we feel that nature cannot be that simple. It is difficult to imagine that faunas subject to the much higher flux regimes characteristic of more eutrophic areas will have test production rates per unit of organic carbon identical to those described by Herguera (1992), and a non-linear response to increasing export production has indeed been suggested by Schmiedl and Mackensen (1997). Naidu and Malmgren (1995) showed that in low oxygen environments on the Oman margin OMZ, BFAR does not reflect surface-water productivity. In spite of these problems, BFARs have tentatively been applied to reconstruct paleo-productivity semi-quantitatively (e.g., Den Dulk, Reichart, van Heyst, Zachariasse, & Van der Zwaan, 2000; Herguera, 2000; Wollenburg & Kuhnt, 2000). No objective tests are available to check the reliability of the results, a problem with all paleo-productivity proxies, so that we are still in doubt about the reliability of this method.

2.3.2. Paleoproductivity proxies based on flux-dependence of individual species

The realisation that foraminiferal species depend on specific organic flux rates dates from the 1970s, when Lutze and co-workers (Lutze, 1980; Lutze & Coulbourn, 1984) conclusively showed that some foraminiferal taxa (e.g., *Uvigerina* spp.) have high relative densities in areas with high organic input. Until today, only very few studies have tried to accurately quantify the flux-dependence of the various benthic foraminiferal taxa.

In a study of the morphology of benthic foraminifera in core-top samples from the Norwegian Sea, Corliss and Chen (1988) observed a rapid shift from faunas dominated by morphotypes considered typical of epifaunal lifestyles (trochospiral, milioline) to faunas dominated by morphotypes considered typical of infaunal microhabitats (rounded planispiral, flattened ovoid, tapered cylindrical, spherical, tapered flattened) at a water depth of $\sim 1,500$ m. They estimated that this change in morphotype dominance takes place at an organic flux level of 3–6 $\text{gC}/\text{m}^2/\text{yr}$. Buzas, Culver, and Jorissen (1993) calculated that microhabitat assignments on the

basis of morphology have only a 75% accuracy, but the data of Corliss and Chen (1988) clearly document the existence of a major faunal change related to a specific organic flux level.

In a new analysis of the large datasets generated by the Lutze-team, Altenbach et al. (1999) investigated the dependence on specific organic flux rates for various foraminiferal taxa. Rather disappointingly, the range of most foraminiferal taxa appears to be very large: correlation coefficients between the percentage in the total fauna of almost all taxa and the organic flux rates at the stations where they are found are very weak. Altenbach et al. (1999) concluded that down to 2,000 m the succession of faunal species seems to be related mainly to bathymetry, whereas only below 2,000 m a relation with export production becomes visible, as confirmed for faunas from the Gulf of Guinea (Altenbach, Lutze, Schiebel, & Schönfeld, 2003). Altenbach et al. (1999) concluded that the presence or absence of a taxon does not seem to be a valuable measure for the reconstruction of flux rates. In spite of these rather disappointing results, they suggested that an organic flux rate of $\sim 2\text{--}3$ gram organic carbon per meter square per year ($\text{gC}/\text{m}^2/\text{yr}$) is an important threshold limit for many species, as confirmed by Jian et al. (1999), De Rijk et al. (2000) and Weinelt et al. (2001). De Rijk et al. (2000) observed that such a faunal threshold occurs at an estimated input of labile organic matter of $\sim 2.5 \text{ g}/\text{m}^2/\text{yr}$ in the Mediterranean, which occurs at $\sim 1,500$ m water depth in the eutrophic Western Mediterranean, but shoals to ~ 400 m depth in the much more oligotrophic eastern Mediterranean. Morigi et al. (2001) used multivariate statistical analysis of recent foraminiferal thanatocoenoses in the upwelling area off Cap Blanc, a part ($19\text{--}27^\circ\text{N}$) of the much larger area (Arctic Ocean, Norwegian-Greenland Sea, NW Africa, and Guinea Basin) investigated by Altenbach et al. (1999), to subdivide frequent taxa into six groups, each with a preference for a rather narrow range of organic flux rates.

In view of these studies, we think that Altenbach et al. (1999), who studied a huge area, may have been too pessimistic when they concluded that the presence or absence of foraminiferal species cannot be used to reconstruct organic flux levels. Taxa with a preference for a rather narrow range of flux levels will necessarily display a weak correlation coefficient with organic flux when their percentage distribution is compared with the total range of observed flux rates over a huge area.

The direct application of these observed flux-dependencies is hampered by three problems:

- 1) In all previously mentioned studies, the estimated organic flux values to the ocean floor were based on a combination of (usually satellite-image derived) primary productivity (PP) values and empirical flux equations. As a consequence, the resulting J_z (organic flux arriving at a water depth of z meters) values are very approximate, and it is difficult to compare flux-dependencies between different regions. The obvious solution would be to calibrate these calculated flux values by a comparison with actual flux data observed in sediment traps in the same areas. Unfortunately, sediment trap data are still scarce, in many areas non-existent, and we do not have measured flux data for larger sets of stations. However, sediment trap data for only one or two stations in each investigated area

would already allow us to verify the correctness of the order of magnitude of values provided by the flux equations, and, when necessary, to apply corrections for nearby stations. Successful proxy calibration depends on the acquisition of reliable flux data for many open ocean and continental margin areas.

- 2) Presently most scientists agree that the organic flux is the main parameter structuring open ocean benthic faunas, but evidently it is not the only one. Other factors may modify the foraminiferal response to varying flux levels, especially the response as expressed in presence/absence patterns. Therefore, proxy calibration efforts should consider not only the relations between faunal characteristics and J_z , but should take other environmental factors (such as bottom water oxygenation, current velocity or sediment grain-size) into account.
- 3) Foraminifera have a wide range of ecological strategies. Species vary from K-selected, highly specialised taxa, occupying very specific and narrow ecological niches, to much more opportunistic, r-selected taxa, occupying a wide range of ecological niches. The group of r-selected taxa shows a strong response to highly episodic phytodetritus deposits, but will probably be weakly dependent upon annual flux rates. Good marker species for annual organic flux rates are expected to be K-selected species.

These problems are serious, but we do not think that they are insoluble. Tentative quantifications of preferred flux regimes for individual species are still scarce, but in our opinion, the first results are encouraging. A large research effort is needed to improve the quantification of the flux of organic matter to the benthic ecosystem, and to determine upper and lower tolerance limits as well as the optimum flux range for many foraminiferal taxa. We are convinced that such an inventory could be the basis of a reliable paleoproductivity proxy.

2.3.3. Paleoproductivity proxies based on flux-dependency revealed by multivariate statistical methods

Since the early 1990s Loubere and co-workers (Loubere, 1991, 1994, 1996, 1998; Fariduddin & Loubere, 1997; Loubere & Fariduddin, 1999) built up a large set of core-top data on benthic foraminiferal assemblage distribution in the Atlantic, Indian and Pacific Ocean, and developed a multiple regression function linking the composition of the foraminiferal faunas to the primary production in the overlying surface waters. For their samples from a fairly narrow bathymetrical range (2,800–3,500 m), the correlation between bottom water oxygenation and primary production was very weak, so that apparent reactions to different organic flux rates cannot be caused by co-varying oxygen concentrations. In the 1994 and 1996 papers, Loubere presents a Pacific Ocean dataset of 74 samples. The multiple regression analysis shows a very strong correlation coefficient to surface water primary production values. Fariduddin and Loubere (1997) present a similar analysis for an 84 station Atlantic Ocean dataset, where productivity also seems to be the most important ecological factor. In 1998, Loubere presented a paper in which the Pacific multiple regression function was tested with Indian Ocean core-top material (water depth 2,400–3,500 m). Although in general the Pacific regression function

reproduced the Indian Ocean primary production values rather well, the Indian Ocean benthic foraminiferal faunas differed by much higher percentages of *Epistominella*, interpreted as a response to the more seasonal aspect of the organic matter flux in the Indian Ocean. In 1999, Loubere and Fariduddin presented a multiple regression analysis of a 207 station sample set from the Pacific, Indian and Atlantic Oceans, from water depths between 2,300 and 3,700 m. Again, the correlation coefficients between primary production, seasonality and other parameters are weak.

Kuhnt et al. (1999) applied correspondence analysis to data on benthic foraminiferal faunas in 43 box cores from the South China Sea. The first factor of their correspondence analysis has a very strong correlation to the estimated organic carbon flux to the ocean floor. This relation was then used to reconstruct the organic carbon flux to the ocean floor for the last glacial–interglacial cycle. In a study of 37 Rose-bengal stained samples from the Laptev Sea (Arctic Ocean), Wollenburg and Kuhnt (2000) found a strong positive correlation between their first factor and the organic flux to the ocean floor, which was used by Wollenburg et al. (2001) to reconstruct a paleoproductivity record on the basis of faunal successions found in two piston cores from the Arctic Ocean.

These multivariate statistical approaches are promising, but they suffer from the general weaknesses of such methods. First, multiple regression analyses ideally need about 20 times more samples than variables in order to be reliable (Tabachnick & Fidell, 1983). Even the 207 station dataset of Loubere and Fariduddin (1999) is far too small in view of the large number of variables (species abundances). Next, all described regression functions are strictly empirical equations, thus not based on understanding of the ecological mechanisms controlling the faunal distribution. With other words, the statistical relationships are black boxes, with no underlying logic to explain the results. Recurrent combinations between environmental parameters observed in today's oceans (e.g., low temperature and high primary production, or high organic flux and low oxygen concentrations) may not have existed or may have been different in the past, causing different responses of faunal composition to the target parameter. The application of statistical relations observed in recent ecosystems becomes especially hazardous when extrapolation is used to reconstruct conditions not present in the dataset used for calibration. Such non-analog conditions include extreme primary production regimes (higher or lower than today), or combinations of environmental parameters (organic flux, oxygenation, quality of organic matter, etc.) not encountered in the recent oceans. For instance, in the geological past deep ocean temperatures were considerably higher than today, and it has been speculated that the resulting increased metabolic activity would lead to increased organic matter mineralisation in the water column (and a lower flux to the ocean floor). Benthic faunas would probably have required higher food fluxes to sustain a similar biomass (Thomas, 2007).

Finally, as for most proxies, we can only check the validity of the results by comparing them to estimates obtained by independent (e.g., geochemical) proxy methods. In spite of our instinctive mistrust when confronted with methods not based on a mechanistical understanding of observed ecological patterns, we agree that multivariate statistical methods are a promising pathway to a useful

paleoproductivity proxy. It is therefore important to continue the efforts to increase the size of available datasets, and to extend this type of study to a wider range of continental slope and open ocean environments.

2.3.4. Paleoproductivity proxies based on the benthic foraminiferal accumulation rate (BFAR)

In 1991, Herguera and Berger proposed the BFAR (number of benthic foraminifera per unit of area per unit of time) as a paleoproductivity proxy. The underlying hypothesis of the method is that for every mg of organic carbon reaching the ocean floor, a fixed number of fossil foraminiferal tests is deposited. In their 1991 benchmark paper, Herguera and Berger indicated that for such a concept to work, four conditions must be met: (1) a linear relation must exist between the organic matter flux and the number of fossilised foraminifera; (2) the flux of organic matter arriving at the ocean floor must in a linear way depend on surface water primary production; (3) the sedimentation rate must be invariable or known in sufficient detail; and (4) there must be no significant carbonate dissolution. The authors suggest that probably none of these assumptions is fully met.

Herguera and Berger (1991) investigated two series of cores in the western equatorial Pacific, from oligotrophic to mesotrophic areas with primary production rates of 65–80, and 105–135 g/m²/yr, respectively. After calculation of the flux of organic matter to the ocean floor by applying a flux equation based on sediment trap data, and a comparison with the accumulation rates of foraminifera in core-tops, they arrived at the conclusion that for every mg of organic carbon reaching the ocean floor, ~1 foraminiferal test larger than 150 µm is fossilised. Herguera (1992) exploits various flux formulae, and concludes that the BFAR is not reliable below a water depth of ~4,000 m due to carbonate dissolution. Naidu and Malmgren (1995), who tested the BFAR in the Oman Margin OMZ, concluded that the proxy does not work in a low oxygen setting.

Although BFAR has been applied widely as a semi-quantitative proxy of paleoproductivity (e.g., Thomas et al., 1995; Nees, Altenbach, Kassens, & Thiede, 1997; Schmiedl & Mackensen, 1997; Den Dulk et al., 2000; Herguera, 2000; Wollenburg & Kuhnt, 2000), most authors have not used the BFAR concept to obtain quantitative estimates of primary production, or even of the organic flux to the ocean floor, because of the problems outlined by Herguera and Berger (1991), and the absence of a reliable calibration, either in the western equatorial Pacific or in regions with different patterns of productivity.

Such a calibration is needed because it is very improbable that in different flux regimes a similar number of foraminifera will be deposited per gram organic carbon arriving at the ocean floor (Schmiedl & Mackensen, 1997). One would expect that in eutrophic areas, with a high export production, faunas would be dominated by rather opportunistic taxa which may produce much more offspring per mg organic carbon than the more K-selected taxa encountered in more oligotrophic areas. The scheme presented in Figure 5 presents the successive steps for the calibration of a paleoproductivity proxy based on the BFAR.

The first problem (A in Figure 5) is the quantitative relation between primary production in the surface waters (PP), the quantity of organic matter that leaves the

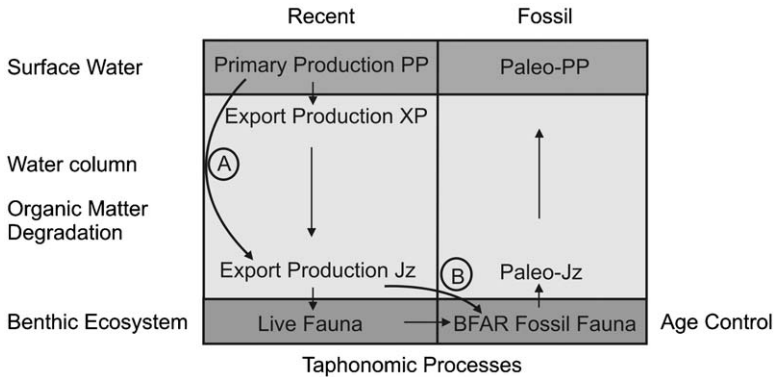


Figure 5 Scheme indicating the various steps and potential problems involved in the calibration of the BFAR as a paleoproductivity proxy. See text for further explanation.

productive surface layer (Export Production, XP), and the flux of organic matter to the sea floor at a water depth of z meters (J_z). As outlined in Section 2.3.1, equations quantitatively describing these relations have been developed on the basis of a comparison of primary production values measured in surface waters with sediment trap data. In paleoceanography, once the paleo- J_z (flux to the sea floor) has been reconstructed, these flux equations can theoretically be used to calculate paleo-PP (past primary production). However, all attempts to translate a reconstructed paleo- J_z into a paleo-PP value should accept a wide error margin, due to the uncertainties of the flux equations and the possibility of a significant contribution of laterally advected material. Only a very small portion (0.01–1%) of the total primary production is transported to the ocean floor (e.g., Murray et al., 1996), so that even small changes in the transport mechanism may have a large impact on the amount of organic matter arriving at the sea floor. It seems therefore judicious to limit the use of the BFAR concept to the reconstruction of the organic flux to the ocean floor (paleo- J_z) for the time being.

For calibration purposes, the characteristics of the benthic fauna have to be compared quantitatively with recent J_z values, usually calculated by introducing satellite-derived PP-values into the afore-mentioned flux equations, adding another source of uncertainty. Difficulties to obtain precise J_z values, needed to quantify its impact on the foraminiferal faunas, have seriously hampered the development of the BFAR concept.

A next problem is the calibration of the BFAR by comparison with recent J_z values (B in Figure 5). When more or less accurate estimates of J_z are available, they can easily be compared with the density of a living foraminiferal fauna (e.g., Fontanier et al., 2002), but it is much more complicated to compare J_z with the accumulation rate of fossil foraminifera. In order to do so one must have a precise knowledge of the sediment accumulation rate, requiring a very detailed age control for the topmost sediment layer that in ideal cases and for the last ~100 years may be obtained by downcore profiles of excess ^{210}Pb and/or ^{237}Cs profiles (e.g., Nittrouer, DeMaster, McKee, Cutshall, & Larsen, 1984). However, bioturbation activities

and/or sediment resuspension after deposition may significantly alter the ^{210}Pb -profiles, which can lead to incorrect calculation of the sediment accumulation rates. In aquatic environments, assumptions must also be made about the exact transfer process of the radionuclides from atmosphere to sea floor (e.g., Appleby & Oldfield, 1992). Next, the loss of foraminiferal tests due to taphonomical processes should be extremely well constrained. Unfortunately, our present knowledge of taphonomical processes is far too fragmentary to have any realistic hope to calibrate the BFAR method by comparing J_z with recent or core-top foraminiferal faunas, for which taphonomical processes have not been finished. An additional problem is the fact that recent faunas may show important seasonal variability, and are therefore not always representative for longer periods.

We thus are left with the option to calibrate the BFAR on the basis of (sub-) recent material, for which two essential conditions have been met: (1) taphonomical processes have been terminated, and (2) the fauna is representative of recent environmental conditions. Unfortunately, it is very difficult, if not impossible, to be certain that both conditions are fulfilled. In recent or very modern faunas, faunal transformations due to taphonomical processes have not been finished (as discussed above). For fossil faunas we do not know whether the environmental parameters by which they were structured were similar to the recent conditions observed at the site. The best solution seems to calibrate the proxy relationship by using faunas deposited during the early Holocene or other recent interglacial periods under environmental conditions (as far as we know them) relatively comparable to modern ones. A considerable uncertainty will be the result, thus paleo- J_z estimates resulting from BFAR will always have a large error margin.

A final problem is the quality of the organic flux to the ocean floor. Fresh marine organic matter (phytodetritus) is directly consumed by benthic foraminifera, and its availability in large quantities may lead to the production of a relatively large number of produced tests per mg of organic carbon (e.g., , 1988, 1993). Aged, or continental organic matter, on the contrary, will contain more refractory components, and only a small part can be directly consumed by the benthic fauna. As a consequence, fewer foraminifera will be produced per mg organic carbon when refractory organic carbon accounts for an important part of the input. There are as yet no methods available that are routinely used to describe the nutritious value of organic matter, and it may be particularly complicated to take this parameter into account.

Below, we use data from three piston cores (Table 2) from the continental margin off Cap Blanc (NW Africa) to illustrate some of the calibration problems. Core Sed17aK was sampled in an oligotrophic area, at 2,975 m depth, Core KS04 at 1,000 m depth in an area of strong coastal upwelling, whereas core Sed20bK comes from a site at 1,445 m depth, presently outside the direct influence of upwelling. For all three cores age control is based on stable isotope analyses of planktonic foraminifera (*G. bulloides* for Sed17aK and Sed20bK; *G. inflata* for KS04) and correlation to an orbitally tuned age model. We will use core Sed20bK, which has the highest time resolution, as an example (Figure 6).

In all three cores the topmost sediment layer has been lost and only the lower part of the Holocene is present, as is often the case for piston cores. In core Sed20bK (as well as in the other cores) the BFAR (of the $>150\ \mu\text{m}$ size fraction of

Table 2 Cores KSo4, Sed20bK, and Sed 17aK: Geographical Position, Water Depth, Age at the Bottom of the Core (Indicated as MIS: Marine Isotopic Stage), Number of Age Control Points, Primary Production (According to [Schemainda et al., 1975](#)), Total Estimated Organic Carbon Flux to the Sea Floor, and its Labile Component (Calculated According to Equation (1), After [Herguera \(1992\)](#)).

| Core | Latitude | Longitude | Water depth | Age at bottom core | Number of age control points | Primary production (g/m ² /yr) | J_z total C _{org} flux (g/m ² /yr) | J_z labile component (g/m ² /yr) |
|---------|------------|------------|-------------|--------------------|------------------------------|---|--|---|
| KSo4 | 20°34.70'N | 18°08.80'W | 1,000 m | MIS7 | 8 | 200 | 7.9 | 5.65 |
| Sed20bK | 25°01.70'N | 16°39.02'W | 1,445 m | MIS5d | 12 | 100 | 2.7 | 1.3 |
| Sed17aK | 25°16.80'N | 17°06.45'W | 2,975 m | MIS6 | 15 | 65 | 1.1 | 0.35 |

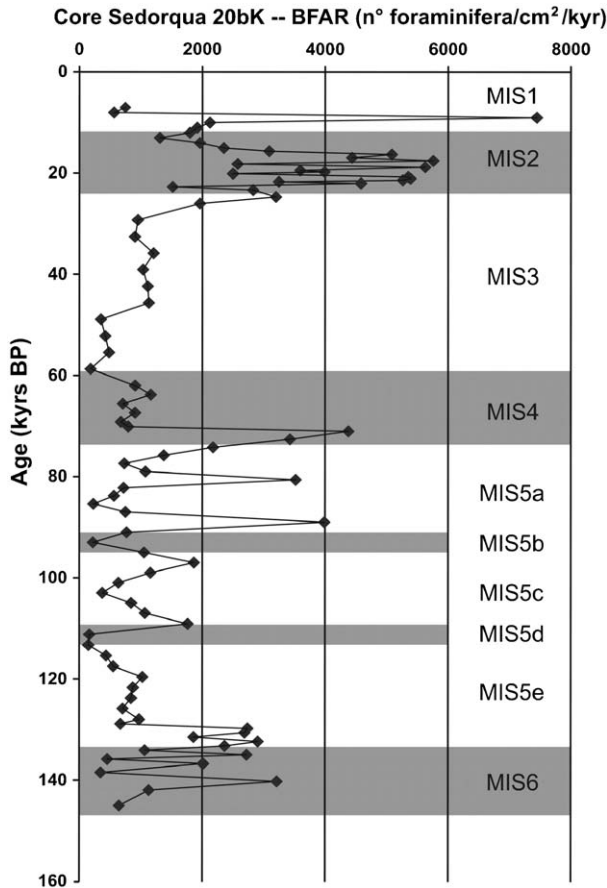


Figure 6 Variation of BFAR in NW Africa core Sed20bK (25°01.70'N, 16°39.02'W, 1,445 m), over the last two glacial-interglacial cycles. After Guichard et al. (1999).

benthic foraminifers, following Herguera & Berger, 1991) shows a strong variability, with peak values ~ 10 times higher than the baseline (Figure 6). Such peaks are present in glacial and interglacial periods, and may be due to rapid changes in sediment accumulation rate which are not recognised due to the imperfect time resolution, or may be real, due to the occurrence of long-lived benthic foraminiferal bloom periods in response to upwelling events that have resulted in periods with large-scale phytodetritus deposition. Because the topmost sediment layer has not been recovered, we do not know whether the present organic flux to the ocean floor corresponds to (1) a peak of BFAR, (2) Holocene background BFAR values, or (3) some intermediate BFAR value. The only acceptable solution is to consider the two extreme situations, and use those to define an error envelope. Guichard, Jorissen, and Peypouquet (1999) decided to compare recent J_z values with BFAR values of Marine Isotope Stage (MIS) 5a, an interglacial period with climatic conditions more or less comparable to the Holocene. With the exception of the peak

value observed at $\sim 9,000$ BP, BFAR values observed in MIS 5a cover the range of values observed in the lower Holocene part of the core.

Today, primary production at site SED20bK is ~ 100 gC/m²/yr (Schemainda, Nehring, & Schultz, 1975). For the calculation of the flux to the sea floor, we use the flux equation proposed by Herguera (1992):

$$J_z = \left(2\sqrt{PP} \times \frac{PP}{z} \right) + \left(\frac{5}{\sqrt{PP}} \times \frac{PP}{\sqrt{z}} \right) \quad (1)$$

in which J_z is the total organic flux to the sea floor at water depth of z meter, and PP primary production in the surface waters. We prefer this flux equation over other ones (review in Herguera, 1992), because it distinguishes between a labile component (first term) that rapidly diminishes with water depth, and a more refractory component (second term) that is much more constant with water depth.

For site KS20bK, the estimated total J_z is 2.7 gC/m²/yr; the labile component is estimated at 1.4 gC/m²/yr, the more refractory component 1.3 gC/m²/yr. Three different hypotheses can now be tested. Estimated recent flux rates:

1. correspond to the background (minimal) BFAR values observed in MIS 5a,
2. correspond to the average BFAR values during MIS 5a
3. correspond to the peak values during MIS 5a, which seems improbable in view of the moderate recent primary production value.

Next, for these three different hypotheses (recent PP corresponds to minimum, average or peak BFAR) the BFAR values can be compared with the estimated values of the total organic flux to the sea floor, and with its labile component. The result is the number of foraminifera deposited per mg of organic carbon (total or labile) arriving at the sea floor (Table 3); for core Sed20bK this number varies from 2.9 to 28 foraminifera per mg for total organic carbon, and from 5.6 to 54 foraminifer per mg if only the labile component is considered.

This first estimate produces values that differ by an order of magnitude, but it seems very improbable that the recent primary production value (very moderate for the Cap Blanc region) corresponds to peak BFAR values (Figure 6), which probably represent periods of intensified upwelling. It seems more probable that recent PP values correspond to a BFAR value between the minimum and average value, which would mean that for every mg of organic carbon arriving at the ocean floor, 2.9–5.6 fossil foraminifera >150 μ m are preserved (or 5.6–10.7 per mg labile organic carbon).

For cores KS04 (water depth 1,000 m) and Sed17aK (water depth 2,975 m) (Guichard et al., 1997), a similar procedure was followed (Table 3). The flux rates were estimated by introducing primary production measurements of Schemainda et al. (1975) into flux equation (1) (Herguera, 1992). Next, the number of fossil foraminifera per mg organic carbon was calculated for each of the three hypotheses, once using the total organic flux, and once using only the labile component (Table 3).

From these BFAR calibrations we observe:

- 1) The tentative value of 1 foraminifer larger than >150 μ m deposited per 1 mg organic matter reaching the sea floor (Herguera & Berger, 1991) has the right order of magnitude, but appears too low for our continental margin cores.

Table 3 Cores Sed20bK, KSo4, and Sed17aK: Comparison of Observed Benthic Foraminiferal Accumulation Rates (BFAR, According to Three Hypotheses) with the Estimated Flux of Organic Carbon to the Sea Floor (Total Flux and Labile Component).

| Core Sed20bK–1,445 m | BFAR (forams/cm²/kyr) | Total C_{org} flux (2.7 gC/m²/year) | C_{org}, labile component (1.4 gC/m²/yr) |
|-----------------------------|---|---|---|
| Background BFAR | 780 | 2.9 forams/mg C _{org} | 5.6 forams/mg labile C _{org} |
| Average BFAR | 1,500 | 5.6 forams/mg C _{org} | 10.7 forams/mg labile C _{org} |
| Peak values BFAR | 4,000–7,500 | 15–28 forams/mg C _{org} | 28–54 forams/mg labile C _{org} |
| Core KS04–1,000 m | BFAR (forams/m²/yr) | Total C_{org} flux (7.9 gC/m²/yr) | C_{org}, labile component (5.65 gC/m²/yr) |
| Background BFAR | 17,000 | 2.2 forams/mg C _{org} | 3.0 forams/mg labile C _{org} |
| Average BFAR | 24,800 | 3.1 forams/mg C _{org} | 4.4 forams/mg labile C _{org} |
| Peak values BFAR | 50,000 | 6.3 forams/mg C _{org} | 8.8 forams/mg labile C _{org} |
| Core Sed17aK–2,975 m | BFAR (forams/m²/yr) | Total C_{org} flux (1.1 gC/m²/yr) | C_{org}, labile component (0.35 gC/m²/yr) |
| Background BFAR MIS5a | 450 | 0.4 forams/mg C _{org} | 1.3 forams/mg labile C _{org} |
| Peak value BFAR MIS1 | 4,000 | 3.6 forams/mg C _{org} | 11 forams/mg labile C _{org} |

Note: Three scenarios are tested, and the numbers of foraminifera > 150 μm preserved in the sediment are given for each case.

If we do not consider the rather improbable numbers based on Hypothesis 3 (recent situation corresponds to peak BFAR), we find that 0.4–5.6 fossilised foraminifera $>150\ \mu\text{m}$ are deposited per mg total organic carbon arriving at the sea floor off NW Africa.

- 2) The values for our deepest core (Sed17aK; Table 3) correspond rather well to the values proposed by Herguera and Berger (1991), whereas our two continental margin cores, representing much more eutrophic ecosystems, show much higher values (2.2–5.6 foraminifera deposited per mg total organic carbon arriving at the ocean floor). This suggests that the relationship between J_z and BFAR is not linear. More eutrophic areas appear indeed to be inhabited by more opportunistic taxa that produce more tests per unit of organic carbon.
- 3) By only considering the labile component of the downward organic flux, the differences between our deep oligotrophic site, and our shallower, more eutrophic sites diminish slightly (Table 3). This suggests that it may be judicious to consider only the labile part of the organic flux to the ocean floor.

We think that this example documents that BFAR is a promising proxy for paleo- J_z . We need to develop a large data-basis of BFAR records in well-dated cores from regions with different patterns of PP (oligotrophic to eutrophic, highly seasonal to less seasonal), in order to determine how BFAR responds to various organic flux regimes.

2.3.5. Reconstructing the quality of the downward organic flux

Benthic foraminifera, as well as most other benthic organisms, will mainly respond to the flux to the ocean floor of labile organic particles that are easily metabolised. Terrigenous or old marine organic particles (refractory organic matter) have only a limited nutritious value, and will commonly bypass the oxic niches at the sediment-water interface. Only in the anaerobic environments deeper in the sediment part of this material is recycled (e.g., Fenchel & Finlay, 1995), whereas the remainder will be preserved as sedimentary organic matter. The absolute quantities and relative proportions of these two types of organic matter (labile versus refractory) differ as a function of the amount of terrigenous input (usually determined by the distance from land) and the local biological and sedimentological conditions. At sites with an influx of predominantly labile organic matter, most will be consumed by the benthic fauna at the sediment-water interface, whereas at sites with more refractory components, a larger part of the organic matter will become preserved in the sediments. It thus seems unlikely that sedimentary organic flux rates (the amount of fossilised C_{ORG} per surface area per year) can provide a reliable paleoproductivity proxy (e.g., Wefer et al., 1999), although the sedimentary organic carbon content has often been used to reproduce past biological production (e.g., Müller & Suess, 1979; Sarnthein, Winn, Duplessy, & Fontugne, 1987, 1988).

In our opinion, the differential response of benthic foraminiferal faunas to influx of these two types of organic matter could be used to gain insight into the character of past organic particle supply, but more information is required about the dependence of various species on food quality. Surface-dwelling taxa have been said to rely on labile organic matter, whereas intermediate and deep infaunal taxa tolerate

more refractory food particles made available by partial recycling under anaerobic conditions (e.g., Jorissen, 1999a; Fontanier et al., 2005).

In order to gain insight into the organic matter quality, Guichard et al. (1999) combined data on BFAR and sedimentary organic matter flux (Figure 7). The underlying idea is that the BFAR varies as a function of the labile organic carbon input, but is rather insensitive to the input of refractory organic carbon. The quality of the organic matter flux probably changed considerably over time: during periods with intensified upwelling, large amounts of labile organic matter reached the sea floor where they undoubtedly triggered biological production, and high BFARs. During prolonged periods without sustained upwelling (as today), on the contrary, the quality of the organic input was probably much lower, but a larger proportion was preserved in the sediment, leading to a relatively low BFAR with respect to the sedimentary C_{org} flux.

Sedimentary C_{org} flux is positively correlated to BFAR, but there is a wide scatter around regression line (1), which represents the average ratio between the BFAR and sedimentary C_{org} flux (of 15 foraminiferal tests per 1 mg sedimentary organic carbon), and thus an average quality of organic matter. Samples with minimal BFAR values probably represent periods when the quality of the organic influx was poor (i.e., minimal labile fraction). Points high above the regression line represent samples with a very high BFAR relative to the sedimentary organic carbon flux, suggesting an organic matter flux dominated by labile components, in this region possibly representing periods with strongly intensified upwelling. The variability of BFAR is not only a response to a varying organic input, but may be partially caused by short-term variability of the sediment accumulation rate, not recognised due to the limits on the resolution of the age model. In spite of these complications, we think that this approach deserves to be further explored.

2.3.6. Reconstructing the periodicity of the downward organic flux

Studies on the Porcupine abyssal plane (Lampitt, 1985; Gooday, 1988; Lamshead & Gooday, 1990) have conclusively shown the importance of the massive phytoplankton

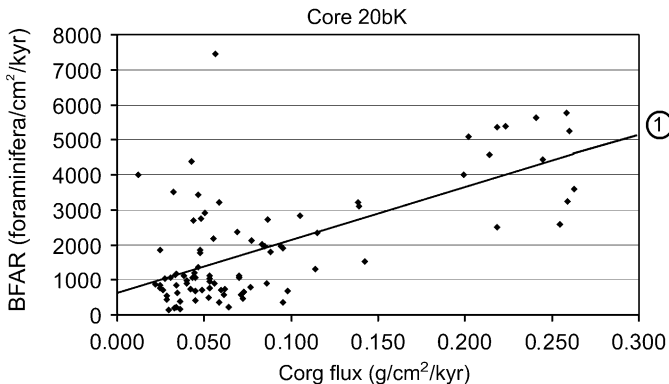


Figure 7 Core Sed20bK; comparison between BFAR and sediment C_{org} flux (after Guichard et al., 1999); 88 samples; regression line (1): $\text{BFAR} = 15,160 \times C_{\text{org}} + 616.5$ ($r^2 = 0.42$).

deposits in the deep ocean, to which the benthic fauna responds by a period of intense biological activity, for many taxa leading to accelerated growth and/or reproduction (Pfannkuche & Lochte, 1993). Benthic foraminifera are among the organisms with the strongest response to such events (e.g., Gooday, 1988, 1993, 1996; Gooday, Levin, Linke, & Heeger, 1992; Gooday & Hughes, 2002). Several small taxa (e.g., *Alabaminella weddellensis*, *Epistominella exigua*, *Epistominella pusilla*, *Fursenkoina* spp., *Globocassidulina subglobosa*) as well as soft-shelled forms rapidly colonise the phytodetritus, producing abundant offspring, leading to the prediction that these taxa will probably become dominant elements in the fossil assemblages at sites with episodic phytodetritus deposits. Smart et al. (1994) applied this concept to Miocene and Quaternary deep-sea cores; Thomas et al. (1995) to two mid-latitude North Atlantic sediment cores (last 45 kyr). They interpreted the abundance of phytodetritus species and increased BFAR during the last deglaciation, as the result of increased surface primary production, and subsequent transport of phytodetritus to the ocean floor in response to a northward migration of the polar front (Thomas et al., 1995). Thomas and Gooday (1996) argued that phytodetritus deposition increased strongly at the establishment of the Antarctic ice sheet (earliest Oligocene), leading to the occurrence of lower diversity assemblages with dominant phytodetritus species at high latitudes. Several other papers follow a similar approach to reconstruct past phytodetritus flux events (e.g., Nees, Armand, De Deckker, Labracherie, & Passlow, 1997, 1999; Nees & Struck, 1999; Ohkushi, Thomas, & Kawahata, 2000).

A potential problem is the fact that most of the indicator species of these pulsed phytodetritus deposits are small (63–150 μm), have therefore not been studied very often, and may be sensitive to dissolution. The *E. exigua*–*A. weddellensis* assemblage was positively correlated with the seasonality of primary production, whereas individual species of this assemblage did not show such a relationship (Sun, Corliss, Brown, & Showers, 2006). In fact, all opportunistic taxa will be advantaged by unstable conditions such as episodic transport of phytodetritus to the ocean floor, but some opportunistic species can respond strongly to phytodetritus input in an oligotrophic deep ocean setting, whereas other taxa may be typical for similar conditions in a more eutrophic continental slope context. For instance, at 550 m water depth in the Bay of Biscay large (adult specimens >150 μm) *Uvigerina peregrina* and *Uvigerina mediterranea* show (together with *E. exigua*) a marked increase in abundance in response to phytodetritus deposits following phytoplankton blooms (Fontanier et al. (2003). These observations are in contrast with the situation in the open-ocean environments of the north-western Pacific (Ohkushi et al., 2000), where *Uvigerina* spp. reacted very differently from *E. exigua*. These results show that we can not validly extrapolate from observations at one location to large areas, and that more observations of faunal variability over time are needed to further understand the periodicity of the past organic matter flux.

2.4. The Water Mass Concept

As described in Section 2.1, the ‘water mass concept’ implied that the differences in physico-chemical parameters between various water masses were responsible for the fact that they could be characterised by a specific benthic foraminiferal fauna.

Below we will discuss arguments that led to the rejection of the fixed bathymetry concept, then consider the pros and cons of the water mass concept, and discuss present ideas on the relations between foraminiferal assemblage structure and the physico-chemical characteristics of bottom waters.

2.4.1. Deep-sea foraminifera as indicators of bathymetry

Attempts to compare bathymetrical species distributions in various oceans led to the conclusion that for many species important differences in bathymetrical distribution existed. For instance, Pujos-Lamy (1973) compared the bathymetrical species succession for shelf to abyssal environments on the French continental margin (Bay of Biscay, NE Atlantic; Caralp, Lamy, & Pujos, 1970) with successions in other ocean basins such as the western Pacific (Polski, 1959), the north-western Pacific (Stschedrina, 1957), the north Pacific (Saidova, 1961), the Californian margin (Bandy, 1961), and the Gulf of Mexico (Phleger, 1960). At water depths of less than 2,000 m at least some cosmopolitan taxa showed comparable depth distributions in the Pacific and Atlantic Oceans, but below that depth the successive faunal associations occurred at significantly different depths. Pujos-Lamy (1973) (in agreement with Boltovskoy, 1965) concluded that these differences in bathymetrical distribution could be explained by the existence of different oceanic provinces, thereby preparing the ground for the water mass theory.

Many subsequent papers (e.g., Streeter, 1973; Schnitker, 1974, 1980; Lohmann, 1978a; Douglas & Woodruff, 1981; Bremer & Lohmann, 1982; Corliss, 1983b; Mackensen et al., 1990, 1995; Schmiedl, Mackensen, & Müller, 1997) confirmed that the upper and lower depth limits of many foraminiferal species show significant differences between various ocean basins, although bathymetrical species successions may be very similar. Many papers on the distribution of living and fossil deep-sea foraminiferal faunas (e.g., Corliss, 1985, 1991; Woodruff, 1985; Thomas, 1986; Mackensen & Douglas, 1989; Woodruff & Savin, 1989; Corliss & Emerson, 1990; Barmawidjaja et al., 1992; Rathburn & Corliss, 1994; Kitazato, 1994; Rathburn et al., 1996; Ohga & Kitazato, 1997; de Stigter et al., 1998; Jorissen et al., 1998; Jannink et al., 1998; Kitazato et al., 2000; Schmiedl et al., 2000; Fontanier et al., 2002; Licari et al., 2003; Hess, Jorissen, Venet, & Abu-Zied, 2005) show that cosmopolitan species, such as *Nuttallides umboniferus*, *Epistominella exigua* or *Cibicides wuellerstorfi*, thrive in all deep-sea basins, without showing a clear preference for a specific bathymetrical range or bio-province.

2.4.2. Deep-sea foraminifera as water mass indicators?

In the 1970s and early 1980s deep-sea foraminiferal assemblages were related to specific water masses, and various authors (e.g., Streeter, 1973; Schnitker, 1974, 1980; Lagoe, 1977; Lohmann, 1978a, 1978b; Gofas, 1978; Corliss, 1978, 1979; Streeter & Shackleton, 1979; Osterman & Kellogg, 1979; Belanger & Streeter, 1980; Miller & Lohmann, 1982; Streeter, Belanger, Kellogg, & Duplessy, 1982; Bremer & Lohmann, 1982; Weston, 1982; Corliss, 1983a, 1983b; Peterson, 1984; Woodruff, 1985; Murray, Weston, Haddon, & Powell, 1986) investigated whether their past spatial distribution could be used to reconstruct deep-sea circulation patterns (i.e., the geographical distribution of water masses). In most of these

papers, the presence of foraminiferal species in some deep-sea environments and their absence in others was explained by their dependence on water mass characteristics rather than by their bathymetrical preferences. Several “index species”, or a “specific combination of benthic foraminiferal species” were related to the specific physical and chemical properties of water masses, in some cases rather subjectively, in others by using statistical methods (Appendix 1). Most of these data pertained to foraminiferal thanatocoenoses sampled in core-tops (of piston, gravity, trigger-weight and grab cores), considered representative of the living faunas. However, the use of thanatocoenoses to determine a possible correlation between foraminiferal distribution and the geographical extension of water masses is highly problematic (e.g., Mackensen et al., 1990; Douglas & Woodruff, 1981). In most cases the uppermost material does not contain recent assemblages, because the sediment-water interface is only rarely sampled by these methods. Taphonomical processes, such as carbonate dissolution below the lysocline or in organic-rich sediments, differential disintegration of fragile tests, deposition of reworked material, or winnowing of autochthonous species may all cause large differences between living and fossil faunas (e.g., Corliss & Honjo, 1981; Mackensen et al., 1990, 1995). In some of the studies documenting the water mass concept (Lohmann, 1978a; Schnitker, 1979; Streeter & Shackleton, 1979), agglutinated and porcellaneous foraminifera were excluded from the faunal counts, leading to less reliable correlations between water mass distribution and foraminiferal assemblages. Moreover, as underlined by several authors (e.g., Thomas et al., 1995; Gooday, 2003), the use of different size fractions in most of these studies (>125, >150, or >250 μm) created important methodology-related faunal discrepancies in datasets from the various study areas.

One of the major problems of the water mass concept is the fact that in most of the classical papers water mass characteristics are presented very simplistically. For instance, the physico-chemical definition of North Atlantic deep Water (NADW) is very complex (e.g., Frew, Dennis, Heywood, Meredith, & Boswell, 2000; Van Aken, 2000). NADW can be subdivided in a number of regional water masses (e.g., LSW, DSOW, ISOW, NEADW, NWADW) with a large variability in physico-chemical parameters (temperature, salinity, nutrient concentrations, oxygen concentration, etc.). In the “Southern Ocean”, some physico-chemical water mass properties such as oxygenation, alkalinity, nutrient concentration and corrosiveness progressively change in watermasses as they move along their pathways along the ocean floor (e.g., Peterson, 1984; Mackensen et al., 1995). Appendix 1 thus does not show clear-cut physico-chemical differences between the various water masses, and one cannot pinpoint the exact water mass properties that limit the occurrence of a specific group of taxa. Furthermore, many environmental parameters defining a specific water mass are cross-correlated (temperature, salinity, pressure, oxygen concentration, nutrient concentration, alkalinity/acidity, current velocity, sedimentological patterns, etc.), posing another obstacle to efforts to determine which physico-chemical parameter(s) control the distribution of the foraminiferal assemblages. We are therefore of the opinion that the water mass concept has lost much of its credibility over the last 20 years.

The alleged relation between faunal composition and water masses has been used to reconstruct the Quaternary history of bottom water circulation

(e.g., Streeter, 1973; Schnitker, 1974, 1976, 1979, 1980; Gofas, 1978; Lohmann, 1978b; Streeter & Shackleton, 1979; Corliss, 1979, 1983a; Caralp, Grousset, Moyes, Peypouquet, & Pujol, 1982; Peterson & Lohmann, 1982; Peterson, 1984; Caralp, 1984; Caralp, 1987; Murray et al., 1986; Murray, 1988). In our opinion, the interpretations of the data presented in these studies are no longer valid.

2.4.3. Deep-sea foraminifera as indicators of complex environmental conditions

A number of ecological studies, mainly based on living (Rose Bengal stained) foraminiferal faunas (e.g., Mackensen et al., 1990, 1995; Schmiedl et al., 1997; Murray, 2001), has shown that the composition of deep-sea benthic foraminiferal faunas is related to a complex of environmental parameters. The faunal composition appears to be mainly determined by the food supply, the characteristics of the sediment and of the overlying surface waters, whereas water mass properties appear to play at best only a minor role. For example, Mackensen et al. (1990) show that the distribution of live foraminiferal faunas in the eastern Weddell Sea is related to parameters such as sediment granulometry, current velocity, organic matter deposition and the corrosiveness of bottom and interstitial waters. The interplay of four main environmental parameters controls the distribution of benthic foraminiferal taxa in a large South Atlantic Ocean database (Mackensen et al., 1995):

1. organic carbon content of the sediment and the exported organic matter flux to the sea-floor,
2. hydrodynamical properties of the benthic environment, and the related sediment grain size,
3. bottom water oxygenation, and
4. bottom water carbonate saturation.

The abundance of *Fontbotia* (= *Cibicidoides*) *wuellerstorfi* is related to the occurrence of young and well-ventilated (NADW-like) bottom waters, where the organic carbon flux does not exceed $1 \text{ gC/m}^2/\text{yr}$. *Cassidulina laevigata*, *Uvigerina peregrina*, and some buliminids and bolivinids are correlated to higher organic carbon flux rates ($>3 \text{ gC/m}^2/\text{yr}$), whereas assemblages dominated by *Epistominella exigua* occur in the low salinity core of NADW, where primary production shows a large seasonal contrast. *Angulogerina angulosa* is well correlated with coarse-grained sediment in environments characterised by strong bottom currents. *Nuttallides umboniferus* is found above the CCD, but below the lysocline (Mackensen et al., 1995). Similarly, Schmiedl et al. (1997) related the spatial distribution of seven living and dead assemblages to the dissolved oxygen content of the bottom water, the organic carbon flux, the organic carbon content of the surface sediment, the nature of the substrate and the related porosity.

2.5. Benthic foraminiferal faunas as indicators of current velocity

Intensified bottom water currents (contour, tidal or slope currents) have been shown to influence the microhabitats and composition of the benthic foraminiferal fauna (Lutze & Altenbach, 1988; Lutze & Thiel, 1989; Linke & Lutze, 1993;

Schönfeld, 1997, 2002a, 2002b). Under high current velocities (20–50 cm/s), some species may live preferentially on elevated substrates, or within sedimentary objects such as pteropod shells. Elevated substrates include large biogenic objects such as coarse shell debris, sponges, crinoids, hydroids, tube worms (Sen Gupta, Smith, & Lobegeier, 2006), or living arborescent benthic foraminifera (e.g., *Rhabdammina* spp.), as well as large terrigenous particles (pebbles). At some mm or cm above the sediment–water interface, specialised epibenthic foraminifera may catch suspended organic matter particles (suspension-feeders). Mackensen et al. (1995) recognised a faunal association dominated by *Angulogerina angulosa*, correlated with very coarse-grained sediments (sand–gravel) and putatively strong bottom currents. On the southern Portuguese continental margin, Schönfeld (1997) described common *Cibicides lobatulus*, *Planulina ariminensis*, *Discanomalina* spp., and *Epistominella exigua* living on elevated substrates, and suggested that these taxa thus maximise the acquisition of suspended organic compounds. A similar assemblage of sessile epifaunal and/or epibenthic suspension feeders was found on the Spanish continental margin, influenced by high velocity bottom currents related to the Mediterranean Outflow Water (Schönfeld, 2002a). Using data from Gulf of Cadiz and the southern Portuguese margin, Schönfeld (2002b) showed a linear correlation between the percentage of taxa thought typical for elevated microhabitats and bottom water current velocity, and used this relation to reconstruct the Holocene history of bottom current velocity.

The use of the composition of benthic foraminiferal assemblages as a proxy for past current regimes is complicated by the fact that active currents may constitute one of the main taphonomical factors, causing resuspension and/or transport at the sediment–water interface, leading to winnowing of or addition of components. Schönfeld (1997) recommended the use of the >250 μm fraction benthic foraminifera in a proxy for bottom current regimes because tests in this large size fraction may be least affected by transport.

The development of such a current velocity proxy requires more ecological studies based on living and subfossil faunas, in order to understand taphonomical modifications, and the record of the faunal response to current regimes in the sediments (Schönfeld, 1997, 2002b). The calibration of such proxies is hampered by the lack of precise and highly detailed measurements of bottom current velocities in regions where information on benthic foraminiferal assemblages is available. For example, no precise current measurements are available for the Bay of Biscay, where foraminiferal ecology has been well studied (e.g., Fontanier et al., 2002, 2003, 2005, 2006; Hess et al., 2005). In the absence of current measurements, various non-faunal proxies for current activity could be used, such as grain size analysis, X-ray photographs and ^{210}Pb and/or ^{234}Th profiles of the upper sediment layers (Fontanier et al., 2005). Direct observations of sedimentary structures in core material, photographic/video surveys of the seafloor and parasound records, may also give information on sedimentary processes and the associated current flow regimes (Schönfeld, 1997). Finally, a potential problem of this proxy method is the fact that we do not have observations that the benthic foraminifera living on elevated substrates are indeed suspension feeders. At least some species considered to be indicators of high current velocities (Schönfeld, 1997, 2002a, 2002b) also

occur in shallow infaunal microhabitats (e.g., *Cibicoides* spp. in Corliss, 1985; Rathburn & Corliss, 1994; Fontanier et al., 2002; *Planulina ariminensis* in De Stigter et al., 1998). Species such as *Cibicides lobatulus*, *Planulina ariminensis*, and especially *Epistominella exigua* are common in areas where we have no indications of significant bottom currents. Most of these species appear to be typical of rather oligotrophic environments, where a microhabitat close to the sediment–water interface and perhaps a suspension–feeding life strategy, may be advantageous.

We thus suggest that the method proposed by Schönfeld should be used with caution, and may not be useful in all open ocean environments. In a well–constrained local context, on the contrary, this method could be used to provide valuable information about past current regimes, but more research is needed.

3. CONCLUSIONS

The main environmental parameters structuring benthic foraminiferal faunas appear to be the organic flux to the ocean floor (its quantity, quality, and periodicity), and bottom water oxygenation (especially at very low ranges). To a lesser degree, sediment grain size and current velocity may act as limiting factors (e.g., Miller & Lohmann, 1982; Lutze & Coulbourn, 1984; Mackensen et al., 1990, 1995; Schmiedl et al., 1997). Conservative water mass properties such as salinity and temperature play only a minor role. Only in the deepest part of ocean basins, where strongly oligotrophic conditions prevail, the corrosiveness of the bottom waters (highest in waters such as AABW), may control the distribution of a few cosmopolitan taxa, such as *Nuttallides umboniferus* or *Oridorsalis tener* (e.g., Bremer & Lohmann, 1982; Mackensen et al., 1990, 1995; Schmiedl et al., 1997).

One of the first conclusions of this overview of the most important paleoceanographic proxy methods based on benthic foraminiferal assemblage characteristics could be that the scientific community working on benthic foraminiferal assemblages has been too modest. Many recent ecological studies show conclusively that under most circumstances the composition of deep–sea benthic foraminiferal assemblages is controlled by a rather limited number of environmental factors. Our understanding of the often complex interactions between these factors, and of the way in which they structure the faunal composition, has made much progress over the last 20 years. Once we understand how environmental parameters influence the faunal composition, it should be possible to use these relations in the reverse way, i.e., to use fossil faunas for the reconstruction of the controlling environmental parameters in the past, although more research is needed in order to establish for which part of earth history conditions resembled those in the present oceans sufficiently for this approach to work.

Methods to reconstruct past values of the essential oceanographic parameters of organic flux to the sea floor and bottom water oxygenation have largely remained qualitative, or at best semi–quantitative. In spite of the indisputable presence of a number of problems (many of which are shared with other, e.g., geochemical, proxy methods), some of the proposed proxies are based on firm ecological observations. Other physico–chemical parameters, such as current velocity, water

corrosiveness to CaCO_3 , temperature and/or salinity appear less important, except perhaps in some specific environmental contexts where benthic foraminiferal proxies may provide adequate reconstructions of their past variability.

The available proxies based on benthic foraminiferal assemblage composition show that they have major potential, but further research is needed to add or improve the quantitative aspects (Table 4). In many cases (e.g., bottom water oxygenation, and C_{org} flux to the ocean floor) this can be done by significantly increasing the size of existing databases. In others (e.g., periodicity of the organic flux), time series observations are necessary. A major obstacle is our insufficient

Table 4 Overview of Proxy Methods Based on Benthic Foraminiferal Assemblage Characteristics, Their Problems and Possible Remedies.

| Parameter | Method | Problems and remedies |
|--|--|--|
| Bottom water oxygenation | Marker species approach | Needs a more objective choice of marker species and a better calibration, probably only feasible for low O_2 concentrations |
| Bottom water oxygenation | Marker species+faunal diversity index | Needs further testing |
| Organic matter flux to the sea floor (J_z) | Flux dependency of species or species groups | Needs better estimates of J_z , needs calibration in many more areas |
| Organic matter flux to the sea floor (J_z) | BFAR | Needs calibration, using a large number of cores from different PP regimes |
| Primary production (PP) | BFAR, flux dependency | Needs better knowledge of relations between PP and J_z |
| Quality of organic input | Marker species approach | Needs much more data on the geochemical composition of organic matter |
| Quality of organic input | BFAR/ C_{org} flux | Needs more research |
| Periodicity of the organic flux | | Needs more time series studies in various productivity contexts |
| Physico-chemical characteristics of water masses | | May work in oligotrophic abyssal environments; more research needed |
| Current velocity | | More current velocity measurements needed; probably only feasible in areas with high current velocities |

knowledge of the differences between recent and fossil faunas due to taphonomical alterations. This phenomenon, of importance for all paleoceanographic proxies, can to some extent be solved relatively easily in the case of foraminiferal assemblages by detailed studies of their vertical succession in sediments deposited in the last 5,000 years, when environmental conditions were probably rather invariable in many areas. Unfortunately, such taphonomical studies are extremely time-consuming. Finally, we want to stress that scientists working with benthic foraminiferal assemblages should try to quantify the very complex relations observed in nature. If a single proxy reconstruction has a large degree of uncertainty, a multi-proxy approach, with as many independent proxies as possible, may successfully reduce the uncertainty related to each single method. Proxies based on foraminiferal assemblage composition are fundamentally different from all geochemical proxies, and thus may provide independent reconstructions of essential oceanographic parameters. We conclude that benthic foraminiferal proxies deserve to be much more widely applied than they are today.

ACKNOWLEDGEMENTS

This paper could not exist without the many prolific scientific discussions we had with numerous colleagues; we would especially like to mention the contributions of Joan Bernhard, Gérald Duchemin, Emmanuelle Geslin, Andy Gooday, Sophie Guichard, Johann Hohenegger, Hélène Howa, Andreas Mackensen, Gerhard Schmiedl, Stefanie Schumacher, and Bert van der Zwaan. We (FJ, CF) acknowledge funding of the Foramprox program by the French national programs PNEDC and PROOF; ET acknowledges funding by NSF and USSSP.

4. APPENDIX 1

Summary of early studies dealing with putative relationship between foraminiferal assemblages and water mass properties and/or bathymetry. Note the differences between papers concerning the water mass characteristics and the faunas inhabiting specific water masses. References are numbered in the first column and listed below. Water mass nomenclature is also detailed below. “nbd” means “no bathymetric data” available in the related note. Taxonomic names are not homogenised between studies. Reference list: (1) Caralp et al. (1970) and Pujos-Lamy (1973); (2) Streeter (1973); (3) Schnitker (1974); (4) Lagoe (1977); (5) Gofas (1978); (6) Gofas (1978); (7) Corliss (1978a); (8) Corliss (1978b, 1979); (9) Osterman and Kellogg (1979); (10) Schnitker (1980); (11) Belanger and Streeter (1980) and Streeter et al. (1982); (12) Burke (1981); (13) Corliss (1981, 1983b); (14) Miller and Lohmann (1982); (15) Peterson (1984); (16) Weston (1982), and Murray et al. (1986). Water mass nomenclature: CPDW/CDW/Circumpolar Deep Water; NADW/North Atlantic Deep Water; AABW/Antarctic Bottom Water; ABW/Arctic Bottom Water; NSOW/Norwegian Sea Overflow Water; AAIW/Antarctic Intermediate Water; IDW/Indian Deep Water; IBW/Indian Bottom Water; PBW/Pacific Bottom Water; PDW/Pacific Deep Water; MW/Mediterranean Water; NEADW/North East Atlantic Deep Water.

| Reference | Study area | Water mass/depth | Temperature (°C) | Salinity (psu) | Oxygenation(ml/l) | Dominant species |
|-----------|---|---|------------------|---------------------|-------------------|---|
| 1 | Northeast Atlantic Ocean, Bay of Biscay | 100–250 m | – | – | – | <i>Cibicides lobatulus</i> , <i>Cibicides refulgens</i> , <i>Gaudryana</i> spp., <i>Textularia saggitula</i> |
| | Northeast Atlantic Ocean, Bay of Biscay | 200–1,700 m | – | – | – | <i>Trifarina angulosa</i> , <i>Trifarina bradyi</i> , <i>Bulimina buchiana</i> , <i>Uvigerina peregrina</i> (s.l.), <i>Epistominella umbonifera</i> |
| | Northeast Atlantic Ocean, Bay of Biscay | 1,700–3,000 m | – | – | – | <i>Bulimina inflata</i> , <i>Bulimina alazanensis</i> , <i>Uvigerina peregrina dirupta</i> , <i>Hyperammina</i> div. sp., <i>Planulina wuellerstorfi</i> , <i>Nonion pompilioides</i> |
| | Northeast Atlantic Ocean, Bay of Biscay | 2,500–4,500 m | – | – | – | <i>Epistominella exigua</i> , <i>Bulimina alanazensis</i> , <i>Planulina wuellerstorfi</i> , <i>Eponides</i> spp., <i>Oridorsalis umbonatus</i> |
| 2 | North Atlantic | NADW, <2,500 m | >3.0 | >35.00 | – | <i>Nummuloculina irregularis</i> , <i>Cibicides kullenbergi</i> |
| | North Atlantic | NADW, 2,500–4,000 m | 2.0 to 3.0 | From 34.90 to 35.00 | – | <i>Planulina wuellerstorfi</i> , <i>Epistominella exigua</i> |
| 3 | North Atlantic | AABW, >4,000 m | <2.0 | <34.90 | – | <i>Epistominella umbonifera</i> |
| | Western North Atlantic Ocean | Lower NADW (nbd) | From 0 to 4.0 | – | – | <i>Hoeglundina</i> sp., <i>Uvigerina</i> sp., <i>Gyroidina</i> sp. |
| | Western North Atlantic Ocean | ABW (NSOW) (nbd) | 1.9 | – | – | <i>Epistominella exigua</i> |
| | Western North Atlantic Ocean | AABW (nbd) | 1.5 | – | – | <i>Epistominella umbonifera</i> |
| 4 | Central Arctic Ocean | <2,500 m (permanently under Arctic pack ice) | –0.5 | ~35.00 | 6.5 | <i>Epistominella arctica</i> , <i>Buliminella elegans</i> var. <i>hensoni</i> , <i>Cassidulina teretis</i> , <i>Valvulineria arctica</i> |
| | Central Arctic Ocean | 2,000–2,500 m (permanently under Arctic pack ice) | –0.5 | ~35.00 | 6.5 | <i>Eponides tener</i> , <i>Eponides tumidulus</i> var. <i>horvathi</i> , <i>Quinqueloculina akneriana</i> , <i>Planulina wuellerstorfi</i> , <i>Ceratobulimina arctica</i> |
| | Central Arctic Ocean | 2,500–3,700 m (permanently under Arctic pack ice) | –0.5 | ~35.00 | 6.5 | <i>Stetsonia horvathi</i> , <i>Eponides tener</i> , <i>Triloculina frigida</i> |

(Continued)

Appendix 1 (Continued)

| Reference | Study area | Water mass/depth | Temperature (°C) | Salinity (psu) | Oxygenation(ml/l) | Dominant species |
|-----------|------------------------------|--|------------------|---------------------|-------------------|--|
| 5 | Western South Atlantic Ocean | NADW, 3,000–3,500 m | From 2.0 to 2.7 | ~34.90 | ~5.7 | <i>Mitolidea</i> , <i>Cibicides wuellerstorfi</i> , <i>Gyroidina altiformis</i> , <i>Pyrgo murrhina</i> |
| | Western South Atlantic Ocean | Transition to AABW, 3,500–4,000 m | From 1.0 to 2.0 | From 34.75 to 34.90 | From 5.0 to 5.7 | <i>Epistominella umbonifera</i> |
| | Central Pacific | PBW (upper layers), ~4,500 | ~1.0 | ~34.71 | – | <i>Cibicides wuellerstorfi</i> , <i>Favocassidulina favus</i> , <i>Pullenia bulloides</i> , <i>Melonis sphaeroides</i> , <i>Melonis bradyi</i> , <i>Pyrgo murrhina</i> |
| | Central Pacific | PBW (lower layers), 4,500–5,000 m | From 1.0 to 1.2 | ~34.71 | – | <i>Epistominella umbonifera</i> , <i>Globocassidulina subglobosa</i> |
| 6 | Western South Atlantic Ocean | Upper NADW/AAIW (= upper branch of CPDW), <2,200 m | ~3.0 | <34.90 | <5.6 | <i>Uvigerina peregrina</i> , <i>Planulina wuellerstorfi</i> , <i>Globocassidulina subglobosa</i> |
| | Western South Atlantic Ocean | NADW, 2,200–4,000 m (+ lower branch of CPDW, ~4,200 m) | From 1.0 to 3.0 | From 34.75 to 34.90 | From 5.0 to 5.6 | <i>Planulina wuellerstorfi</i> , <i>Hoeglundina elegans</i> , <i>Pyrgo</i> spp., <i>Quinqueloculina</i> spp., <i>Nummoloculina irregularis</i> |
| | Western South Atlantic Ocean | AABW, >4,000 m | <1.0 | <34.75 | ~5.0 | <i>Nuttallides umbonifera</i> , <i>Oridorsalis tener</i> |
| 7 | Southeast Indian Ocean | IBW (variable depth) | From 0.8 to 1.2 | From 34.72 to 34.74 | From 4.4 to 4.8 | <i>Uvigerina</i> spp., <i>Epistominella exigua</i> |
| | Southeast Indian Ocean | Mixing between IBW and AABW (variable depths) | From 0.4 to 0.8 | From 34.70 to 34.72 | From 4.8 to 5.0 | <i>Epistominella umbonifera</i> , <i>Planulina wuellerstorfi</i> , <i>Globocassidulina subglobosa</i> |
| | Southeast Indian Ocean | AABW (variable depths) | From –0.2 to 0.8 | From 34.68 to 34.70 | From 5.0 to 5.6 | <i>Globocassidulina subglobosa</i> , <i>Planulina wuellerstorfi</i> , <i>Astronomion echolsi</i> , |
| 8 | Southeast Indian Ocean | AABW1 (variable depths) | From 0.6 to 0.8 | – | – | <i>Oridorsalis tener</i> , <i>Gyroidinoides soldanii</i> , <i>Pullenia bulloides</i> |
| | Southeast Indian Ocean | AABW2 (variable depths) | From –0.2 to 0.4 | – | – | <i>Epistominella umbonifera</i> , <i>Oridorsalis tener</i> , <i>Pullenia wuellerstorfi</i> |
| 9 | Ross Sea | Eastern shelf (almost permanently under pack ice), 400–2,000 m | – | – | – | <i>Cyclammina</i> spp., <i>Hormosina ovicula gracilis</i> , <i>Milliammina arenacea</i> , <i>Reophax nodulosus</i> |

| | | | | | | |
|----|--|--|------------------|---------------------|-----------------|--|
| | Ross Sea | Western shelf (seasonally under pack ice), 500–800 m | – | – | – | <i>Globocassidulina subglobosa</i> , <i>Erhenbergina glabra</i> |
| 10 | Ross Sea | Continental slope | – | – | – | <i>Trifarina earlandi</i> , <i>Cibicides lobatulus</i> |
| | Western North Atlantic Ocean | NADW (nbd) | From 2.0 to 4 | – | – | <i>Globocassidulina subglobosa</i> , <i>Elphidium incertum</i> , <i>Uvigerina peregrina</i> |
| | Western North Atlantic Ocean | (NSOW) (nbd) | 1.9 | – | – | <i>Epistominella exigua</i> |
| | Western North Atlantic Ocean | AABW (nbd) | 1.4 | – | – | <i>Osangularia umbonifera</i> |
| 11 | Norwegian-Greenland Sea | Source of Atlantic Bottom Water, 600–1,200 m | >–1.0 | 34.90 | >6.7 | <i>Melonis barleeanus</i> , <i>Pullenia bulloides</i> , <i>Islandiella norcrossi</i> |
| | Norwegian-Greenland Sea | Source of Atlantic Bottom Water, 950–1,500 m | >–1.0 | 34.90 | >6.7 | <i>Cassidulina teretis</i> , <i>Trifarina angulosa</i> |
| | Norwegian-Greenland Sea | Source of Atlantic Bottom Water, 1,250–3,200 m | –1.0 | 34.90 | From 6.7 to 7.1 | <i>Cibicides wuellerstorfi</i> , <i>Oridorsalis tener</i> |
| | Norwegian-Greenland Sea | Source of Atlantic Bottom Water, >2,900 m | –1.0 | 34.90 | From 6.7 to 7.1 | <i>Oridorsalis tener</i> , <i>Epistominella exigua</i> , <i>Cibicides wuellerstorfi</i> |
| 12 | Western Equatorial Pacific Ontong java Plateau | Deep oxygen minimum | From 1.9 to 3.9 | From 34.58 to 34.65 | From 2.8 to 3.9 | <i>Siphonogobosoma interrupta</i> , <i>Cibicides pseudoungerianus</i> , <i>Astrononion stelligerum</i> , <i>Hoeglundina elegans</i> , <i>Gyrogonia lamarkiana</i> |
| | Western Equatorial Pacific Ontong java Plateau | PDW, 2,500–3,000 m (above lysocline) | From 1.4 to 1.9 | 34.74 | From 4.0 to 4.4 | <i>Epistominella exigua</i> , <i>Pullenia bulloides</i> , <i>Melonis affinis</i> , <i>Melonis pompilioides</i> , <i>Nuttallides umbonifera</i> |
| | Western Equatorial Pacific Ontong java Plateau | PBW, 3,000–4,300 m (below lysocline) | From 1.5 to 1.75 | 34.70 | From 3.4 to 4.6 | <i>Nuttallides umbonifera</i> , <i>Epistominella exigua</i> |
| 13 | Southwest Indian Ocean | CDW (nbd) | – | – | – | <i>Uvigerina</i> sp. |
| | Southwest Indian Ocean | NADW, 1,600–3,800 m | From 0.8 to 2.6 | From 34.66 to 34.72 | – | <i>Planulina wuellerstorfi</i> , <i>Globocassidulina subglobosa</i> , <i>Astrononion echolsi</i> , <i>Pullenia bulloides</i> |

(Continued)

Appendix 1 (Continued)

| Reference | Study area | Water mass/depth | Temperature (°C) | Salinity (psu) | Oxygenation(ml/l) | Dominant species |
|-----------|---------------------------------|--------------------------------------|------------------|---------------------|-------------------|--|
| 14 | Southwest Indian Ocean | AABW, 3,600–4,800 m | From –0.3 to 0.8 | From 34.66 to 34.72 | – | <i>Epistominella umbonifera</i> |
| | Northeast Atlantic Ocean | <700 m | >4.5 | >35.10 | <6.0 | <i>Globobulimina</i> spp., <i>Bulimina</i> spp. |
| | Northeast Atlantic Ocean | 700–2,500 m | From 3.0 to 4.5 | ~35.10 | From 5.5 to 6.0 | <i>Uvigerine peregrina</i> |
| 15 | Northeast Atlantic Ocean | 2500–4,000 m | From 1.5 to 3 | From 35.1 to 35.4 | ~6.0 | <i>Hoeglundina elegans</i> |
| | Eastern Equatorial Indian Ocean | IDW, 2,000–3,800 m | From 1.2 to 2.6 | From 34.72 to 34.76 | From 3.1 to 4.2 | <i>Globocassidulina subglobosa</i> , <i>Pyrgo</i> spp., <i>Uvigerine peregrina</i> , <i>Eggerella bradyi</i> , <i>Cibicidoides wuellerstorfi</i> |
| | Eastern Equatorial Indian Ocean | IBW, 3,800–4,600 m (below lysocline) | ~1.2 | ~34.72 | From 4.1 to 4.4 | <i>Epistominella exigua</i> , <i>Cibicidoides wuellerstorfi</i> , <i>Pullenia bulloides</i> , <i>Oridorsalis umbonatus</i> |
| 16 | Eastern Equatorial Indian Ocean | IBW, 4,200–5,000 m (below lysocline) | From 1.2 to 1.4 | ~34.72 | From 3.9 to 4.4 | <i>Nuttallides umbonifera</i> , <i>Cibicidoides wuellerstorfi</i> , <i>Pullenia bulloides</i> , <i>Oridorsalis umbonatus</i> |
| | Northeast Atlantic Ocean | MW | >4.0 | >35.00 | ~4.0 | <i>Cassidulina obtusa</i> , <i>Globocassidulina subglobosa</i> |
| | Northeast Atlantic Ocean | NEADW (or upper NADW) | From 3.0 to 4.0 | From 34.92 to 34.97 | 6.0 | <i>Epistominella exigua</i> |
| | Northeast Atlantic Ocean | NADW | From 2.5 to 4.0 | From 34.95 to 35.00 | From 5.2 to 5.6 | <i>Planulina wuellerstorfi</i> , <i>Globocassidulina subglobosa</i> , <i>Cibicidoides kullenbergi</i> , <i>Oridorsalis umbonatus</i> |
| | Northeast Atlantic Ocean | AABW | <2.0 | 34.90 | >6.0 | <i>Osangularia umbonifera</i> |

REFERENCES

- Altenbach, A. V., Lutze, G. F., Schiebel, R., & Schönfeld, J. (2003). Impact of interrelated and interdependent ecological controls on benthic foraminifera: An example from the Gulf of Guinea. *Palaeogeography, Palaeoclimatology, Palaeoecology*, *197*, 213–238.
- Altenbach, A. V., Pflaumann, U., Schiebel, R., Thies, A., Timm, S., & Trauth, M. (1999). Scaling percentages and distributional patterns of benthic foraminifera with flux rates of organic carbon. *Journal of Foraminiferal Research*, *29*, 173–185.
- Alve, E. (1990). Variations in estuarine foraminiferal biofacies with diminishing oxygen conditions in Drammensfjord, SE Norway. In: C. Hemleben, M. A. Kaminski, W. Kuhnt & D. B. Scott (Eds), *Paleoecology, biostratigraphy, paleoceanography and taxonomy of agglutinated foraminifera* (pp. 661–694). The Netherlands: Kluwer Academic Publishers.
- Alve, E. (1994). Opportunistic features of the foraminifer *Stainforthia fusiformis* (Williamson): Evidence from Frierfjord, Norway. *Journal of Micropaleontology*, *13*, 24.
- Antia, A. N., Von Bodungen, B., & Peinert, R. (1999). Particle flux across the mid-European continental margin. *Deep-Sea Research*, *I*, *46*, 1999–2024.
- Appleby, P. G., & Oldfield, F. (1992). Application of lead-210 to sedimentation studies. In: M. Ivanovich & R. S. Harmon (Eds), *U-series disequilibrium: Applications to earth, marine and environmental studies* (pp. 731–778). Oxford: Clarendon Press.
- Armstrong, R. A., Lee, C., Hedges, J. I., Honjo, S., & Wakeham, S. G. (2002). A new, mechanistic model for organic carbon fluxes in the ocean based on the quantitative association of POC with ballast minerals. *Deep-Sea Research II*, *49*, 219–236.
- Baas, J. H., Schönfeld, J., & Zahn, R. (1998). Mid-depth oxygen drawdown during Heinrich events: evidence from benthic foraminiferal community structure, trace-fossil tiering, and benthic $\delta^{13}\text{C}$ at the Portuguese Margin. *Marine Geology*, *152*, 25–55.
- Bandy, O. L. (1953a). Ecology and paleoecology of some California Foraminifera, Part I. The frequency distribution of Recent Foraminifera off California. *Journal of Paleontology*, *27*, 161–192.
- Bandy, O. L. (1953b). Ecology and paleoecology of some California Foraminifera, Part II. Foraminiferal evidence of subsidence rates in the Ventura Basin. *Journal of Paleontology*, *27*, 200–203.
- Bandy, O. L. (1961). Distribution of foraminifera, radiolaria and diatoms in sediments of the Gulf of California. *Micropaleontology*, *7*, 1–26.
- Bandy, O. L., & Arnal, R. (1957). Distribution of Recent foraminifera off the west coast of Central America. *American Association of Petroleum Geologists Bulletin*, *41*, 2037–2053.
- Bandy, O. L., & Chierici, M. (1966). Depth-temperature evaluation of selected California and Mediterranean bathyal foraminifera. *Marine Geology*, *4*, 254–271.
- Bandy, O. L., & Echols, J. (1964). Antarctic foraminiferal zonation. *American Geophysical Union, Antarctic Research Series*, *1*, 73–91.
- Barberi, R., & Panieri, G. (2004). How are benthic foraminiferal faunas influenced by cold seeps? Evidence from the Miocene of Italy. *Palaeogeography, Palaeoclimatology, Palaeoecology*, *204*, 257–275.
- Barmawidjaja, D. M., Jorissen, F. J., Puskaric, S., & Van der Zwaan, G. J. (1992). Microhabitat selection by benthic foraminifera in the northern Adriatic Sea. *Journal of Foraminiferal Research*, *22*, 297–317.
- Basov, I. A., & Khusid, T. A. (1983). Biomass of benthic foraminifera in the sediments of the Sea of Okhotsk. *Oceanology*, *33*, 489–495.
- Belanger, P. E., & Streeter, S. S. (1980). Distribution and ecology of benthic foraminifera in the Norwegian Greenland Sea. *Marine Micropaleontology*, *5*, 401–428.
- Berelson, W., Anderson, R., Dymond, J., DeMaster, D., Hammond, D., Collier, R., Honjo, S., Leinen, M., McManus, J., Pope, R., Smith, C., & Stephens, M. (1997). Biogenic budgets of particle rain, benthic remineralization and sediment accumulation in the equatorial Pacific. *Deep-Sea Research II*, *44*, 2251–2282.
- Berger, W., Smetacek, V., & Wefer, G. (1989). Ocean productivity and paleoproductivity—An overview. In: W. Berger, V. Smetacek & G. Wefer (Eds), *Productivity of the Ocean: Present and Past, Dahlem Workshop Reports* (Vol. 44, pp. 1–34). Chichester: Wiley.

- Berger, W., & Wefer, G. (1990). Export production: Seasonality and intermittency, and paleoceanographic implications. *Palaeogeography, Palaeoclimatology, Paleoecology*, *89*, 245–254.
- Bernhard, J. M. (1986). Characteristic assemblages and morphologies of benthic foraminifera from anoxic, organic-rich deposits: Jurassic through Holocene. *Journal of Foraminiferal Research*, *16*, 207–215.
- Bernhard, J. M. (1988). Postmortem vital staining in benthic foraminifera: Duration and importance in population and distributional studies. *Journal of Foraminiferal Research*, *18*, 143–146.
- Bernhard, J. M. (1989). The distribution of benthic foraminifera with respect to oxygen concentration and organic carbon levels in shallow-water Antarctic sediments. *Limnology and Oceanography*, *34*, 1131–1141.
- Bernhard, J. M. (1992). Benthic foraminiferal distribution and biomass related to pore water oxygen: Central California Continental slope and rise. *Deep Sea Research*, *39*, 586–605.
- Bernhard, J. M. (1993). Experimental and field evidence of Antarctic foraminiferal tolerance to anoxia and hydrogen sulphide. *Marine Micropaleontology*, *20*, 203–213.
- Bernhard, J. M., & Alve, E. (1996). Survival, ATP pool, and ultrastructural characterization of benthic foraminifera from Drammensfjord (Norway): Response to anoxia. *Marine Micropaleontology*, *28*, 5–17.
- Bernhard, J. M., & Reimers, C. E. (1991). Benthic foraminiferal population fluctuations related to anoxia: Santa Barbara Basin. *Biogeochemistry*, *15*, 127–149.
- Bernhard, J. M., & Sen Gupta, B. K. (1999). Foraminifera of oxygen-depleted environments. In: B. K. Sen Gupta (Ed.), *Modern Foraminifera* (pp. 201–216). New York, NY: Kluwer Academic Press.
- Bernhard, J. M., Sen Gupta, B. K., & Borne, P. F. (1997). Benthic foraminiferal proxy to estimate dysoxic bottom-water oxygen concentrations; Santa Barbara basin, U.S. Pacific continental margin. *Journal of Foraminiferal Research*, *27*, 301–310.
- Betzer, P. R., Showers, W. J., Laws, E. A., Winn, C. D., Di Tullio, G. R., & Kroopnick, P. P. (1984). Primary productivity and particle fluxes on a transect of the equator at 153°W in the Pacific Ocean. *Deep-Sea Research*, *31*, 1–11.
- Billett, D. S. M., Lampitt, R. S., Rice, A. L., & Mantoura, R. F. C. (1983). Seasonal sedimentation of phytoplankton to the deep-sea benthos. *Nature*, *302*, 520–522.
- Boltovskoy, E. (1965). *Los Foraminiferos recientes*, Editorial (pp. 1–510). Argentina: Universitaria de Buenos Aires..
- Boyd, P. W., & Newton, P. (1995). Evidence of the potential influence of planktonic community structure on the interannual variability of particulate organic carbon flux. *Deep-Sea Research*, *1*, 42, 619–637.
- Bremer, M. L., & Lohmann, G. P. (1982). Evidence of primary control of the distribution of certain Atlantic Ocean benthic foraminifera by degree of carbonate saturation. *Deep-Sea Research*, *29*, 987–998.
- Burke, S. C. (1981). Recent benthic foraminifera of the Ontong Java Plateau. *Journal of Foraminiferal Research*, *11*, 1–19.
- Buzas, M. A., Collins, L. S., Richardson, S. L., & Severin, K. P. (1989). Experiments on predation, substrate preference, and colonization of benthic foraminifera at the shelfbreak off the Ft. Pierce Inlet, Florida. *Journal of Foraminiferal Research*, *19*, 146–152.
- Buzas, M. A., Culver, S. J., & Jorissen, F. J. (1993). A statistical evaluation of the microhabitats of living (stained) infaunal benthic foraminifera. *Marine Micropaleontology*, *20*, 311–320.
- Buzas, M. A., & Gibson, T. G. (1969). Species diversity: Benthic foraminifera in the western North Atlantic. *Science*, *163*, 72–75.
- Caralp, M. H. (1984). Quaternary calcareous benthic foraminifera. *Initial Reports of the Deep Sea Drilling Project*, *80*, 725–755.
- Caralp, M. H. (1987). Deep-sea circulation in the northeastern Atlantic over the past 30,000 years: the benthic foraminiferal record. *Oceanologica Acta*, *10*, 27–40.
- Caralp, M. H. (1989). Abundance of *Bulimina exilis* and *Melonis barleeanum*: Relationship to the quality of marine organic matter. *Geo-Marine Letters*, *9*, 37–43.
- Caralp, M., Grousset, F., Moyes, J., Peypouquet, J.-P., & Pujol, C. (1982). L'environnement confiné du Golfe de Gascogne avant le maximum glaciaire. Actes Colloque International CNRS,

- Bordeaux, Sept. 1981, Bulletin de l'Institut Géologique du Bassin d'Aquitaine, Bordeaux (Vol. 31, pp. 411–422).
- Caralp, M., Lamy, A., & Pujos, M. (1970). Contribution à la connaissance de la distribution bathymétrique des foraminifères dans le Golfe de Gascogne. *Revista Española De Micropaleontología*, 2, 55–84.
- Carney, R. S. (1989). Examining relationship between organic carbon flux and deep-sea deposit feeding. In: G. Lopez, G. Taghon & J. Levinton (Eds), *Ecology of marine deposit feeders* (Vol. 31, pp. 24–58). *Lecture Notes on Coastal and Estuarine Studies*, New York: Springer.
- Carney, R. S. (2005). Zonation of deep biota on continental margins. *Oceanography and Marine Biology. Annual Review*, 43, 211–278.
- Chandler, G. T., Williams, D. F., Spero, H. J., & Xiaodong, G. (1996). Sediment microhabitat effects on carbon stable isotopic signatures of microcosm-cultured benthic foraminifera. *Limnology and Oceanography*, 41, 680–688.
- Cita, M. B., & Podenzani, M. (1980). Destructive effects of oxygen starvation and ash falls on benthic life: A pilot study. *Quaternary Research*, 13, 230–241.
- Corliss, B. H. (1978). Recent deep-sea benthonic foraminiferal distributions in the southeast Indian ocean: Inferred bottom water routes and ecological implications. *Marine Geology*, 31, 115–138.
- Corliss, B. H. (1979). Quaternary Antarctic bottom-water history: Deep-sea benthonic foraminiferal evidence from the Southeast Indian Ocean — Inferred bottom-water routes and ecological implications. *Marine Geology*, 31, 115–138.
- Corliss, B. H. (1983a). Quaternary circulation of the Antarctic Circumpolar Current. *Deep-Sea Research*, 30, 47–61.
- Corliss, B. H. (1983b). Distribution of Holocene deep-sea benthonic foraminifera in the southwest Indian Ocean. *Deep-Sea Research*, 30, 95–117.
- Corliss, B. H. (1985). Microhabitats of benthic foraminifera within deep-sea sediments. *Nature*, 314, 435–438.
- Corliss, B. H. (1991). Morphology and microhabitat preferences of benthic foraminifera from the northwest Atlantic Ocean. *Marine Micropaleontology*, 17, 195–236.
- Corliss, B. H., & Chen, C. (1988). Morphotype patterns of Norwegian Sea deep-sea benthic foraminifera and ecological implications. *Geology*, 16, 716–719.
- Corliss, B. H., & Emerson, S. (1990). Distribution of Rose Bengal stained deep-sea benthic foraminifera from the Nova Scotia continental margin and Gulf of Maine. *Deep-Sea Research*, 37, 381–400.
- Corliss, B. H., & Fois, E. (1990). Morphotype analysis of deep-sea benthic foraminifera from the northwest Gulf of Mexico. *Palaios*, 5, 589–605.
- Corliss, B. H., & Honjo, S. (1981). Dissolution of deep-sea benthonic foraminifera. *Micropaleontology*, 27, 356–378.
- Crosta, X., Pichon, J. J., & Burckle, L. H. (1998). Application of the modern analog technique to marine Antarctic diatoms reconstruction of maximum sea-ice extent at the Last Glacial Maximum. *Paleoceanography*, 13, 284–297.
- Culver, S. J., & Lipps, J. (2003). Predation on and by Foraminifera. In: P. H. Kelley, M. Kowalewski & T. A. Hansen (Eds), *Predator-prey interactions in the fossil record* (pp. 7–32). New York, NY: Kluwer Academic Press/Plenum Publishers.
- Cushman, J. A. (1928). Foraminifera: Their classification and economic use. *Cushman Laboratory for Foraminiferal Research, Special Publications* (Vol. 1, p. 401).
- De Rijk, S., Jorissen, F. J., Rohling, E. J., & Troelstra, S. R. (2000). Organic flux control on bathymetric zonation of Mediterranean benthic foraminifera. *Marine Micropaleontology*, 40, 151–166.
- De Stigter, H. C., Jorissen, F. J., & Van der Zwaan, G. J. (1998). Bathymetric distribution and microhabitat partitioning of live (Rose Bengal stained) benthic foraminifera along a shelf to deep-sea transect in the southern Adriatic Sea. *Journal of Foraminiferal Research*, 28, 40–65.
- Den Dulk, M., Reichert, G. J., van Heyst, S., Zachariasse, W. J., & Van der Zwaan, G. J. (2000). Benthic foraminifera as proxies of organic matter flux and bottom water oxygenation? A case history from the northern Arabian Sea. *Palaeogeography, Palaeoclimatology, Palaeoecology*, 161, 337–359.

- Dodd, J. R., & Stanton, R. J. (1990). *Paleoecology: Concepts and applications* (p. 502). New York, NY: Wiley.
- Douglas, R. G., & Heitman, H. L. (1979). Slope and basin benthic foraminifera of the California Borderland. *Society of Economic Paleontologists and Mineralogists*, 27, 231–246.
- Douglas, R. G., & Woodruff, F. (1981). Deep-sea benthic foraminifera. The oceanic Lithosphere. In: C. Emiliani (Ed.), *The sea* (7, pp. 1233–1327). New York, NY: Wiley-Interscience.
- Elderfield, H. (Ed.) (2004). *Treatise on geochemistry: The oceans and marine geochemistry* (Vol. 6). Amsterdam: Elsevier.
- Epping, E., & Helder, W. (1998). Oxygen budgets calculated from in situ oxygen microprofiles for northern Adriatic sediments. *Continental Shelf Research*, 17, 1737–1764.
- Eppley, R., & Peterson, B. (1979). Particulate organic matter flux and planktonic new production in the deep ocean. *Nature*, 282, 677–680.
- Ernst, S. R., & Van der Zwaan, G. J. (2004). Effects of experimentally induced raised levels of organic flux and oxygen depletion on a continental slope benthic foraminiferal community. *Deep-Sea Research I*, 51, 1709–1739.
- Ernst, S. R., Bours, I., Duijnste, Y., & Van der Zwaan, G. J. (2005). Experimental effects of an organic matter pulse and oxygen depletion on a benthic foraminiferal shelf community. *Journal of Foraminiferal Research*, 35, 177–197.
- Fariduddin, M., & Loubere, P. (1997). The surface ocean productivity response of deeper water benthic foraminifera in the Atlantic Ocean. *Marine Micropaleontology*, 32, 289–310.
- Fenchel, T., & Finlay, B. J. (1995). *Ecology and evolution in anoxic worlds* (p. 276). Oxford: Oxford studies in Ecology and Evolution, New York: Oxford University Press.
- Fischer, G., Ratmeyer, V., & Wefer, G. (2000). Organic carbon fluxes in the Atlantic and the Outhern Ocean: Relationship to primary production compiled from satellite radiometer data. *Deep-Sea Research II*, 47, 1961–1997.
- Fontanier, C., Jorissen, F. J., Anschutz, P., & Chaillou, G. (2006). Seasonal variability of foraminiferal faunas at 1000 m depth in the Bay of Biscay. *Journal of Foraminiferal Research*, 36, 61–76.
- Fontanier, C., Jorissen, F. J., Chaillou, G., Anschutz, P., Grémare, A., & Griveaud, C. (2005). Live foraminiferal faunas from a 2800 m deep lower canyon station from the Bay of Biscay: Faunal response to focusing of refractory organic matter. *Deep-Sea Research I*, 52, 1189–1227.
- Fontanier, C., Jorissen, F. J., Chaillou, G., David, C., Anschutz, P., & Lafon, V. (2003). Seasonal and interannual variability of benthic foraminiferal faunas at 550 m depth in the Bay of Biscay. *Deep-Sea Research I*, 50, 457–494.
- Fontanier, C., Jorissen, F. J., Licari, L., Alexandre, A., Anschutz, P., & Carbonel, P. (2002). Live benthic foraminiferal faunas from the Bay of Biscay: Faunal density, composition, and microhabitats. *Deep-Sea Research I*, 49, 751–785.
- François, R., Honjo, S., Krishfield, R., & Manganini, S. (2002). Factors controlling the flux of organic carbon to the bathypelagic zone of the ocean. *Global Biogeochemical Cycles*, 16, 1087–1097.
- Froelich, P. N., Klinkhammer, G. P., Bender, M. L., Luedke, N. A., Heath, G. R., Cullen, D., dauphin, P., Hammond, D., Hartman, B., & Maynard, V. (1979). early oxidation of organic matter in pelagic sediments of the eastern Equatorial Atlantic: Suboxic diagenesis. *Geochimica et Cosmochimica Acta*, 43, 1075–1090.
- Frew, R. D., Dennis, P. F., Heywood, K. J., Meredith, P. H., & Boswell, S. M. (2000). The oxygen isotope composition of water masses in the northern North Atlantic. *Deep-Sea Research I*, 47, 2265–2286.
- Geslin, E., Heinz, P., Hemleben, Ch., & Jorissen, F. J. (2004). Migratory response of deep-sea benthic foraminifera to variable oxygen conditions: Laboratory investigations. *Marine Micropaleontology*, 53, 227–243.
- Gofas, S. (1978). *Une approche du paléoenvironnement océanique: les foraminifères benthiques calcaires traceurs de la circulation abyssale*. Ph.D. thesis, University of Western Brittany, France.
- Goldstein, S. T., & Corliss, B. H. (1994). Deposit feeding in selected deep-sea and shallow-water benthic foraminifera. *Deep-Sea Research I*, 41, 229–241.
- Gooday, A. J. (1988). A response by benthic Foraminifera to the deposition of phytodetritus in the deep-sea. *Nature*, 332, 70–73.

- Gooday, A. J. (1993). Deep-sea benthic foraminifera species which exploit phytodetritus: Characteristic features and controls on distribution. *Marine Micropaleontology*, 22, 187–205.
- Gooday, A. J. (1994). The biology of deep-sea foraminifera: A review of some advances and their applications in paleoceanography. *Palaïos*, 9, 14–31.
- Gooday, A. J. (1996). Epifaunal and shallow infaunal foraminiferal communities at three abyssal NE Atlantic sites subject to differing phytodetritus input regimes. *Deep-Sea Research I*, 43, 1395–1421.
- Gooday, A. J. (2003). Benthic foraminifera (Protista) as tools in deep-water paleoceanography: Environmental influences on faunal characteristics. *Advances in Marine Biology*, 46, 1–90.
- Gooday, A. J., & Hughes, J. A. (2002). Foraminifera associated with phytodetritus deposits at a bathyal site in the northern Rockall Trough (NE Atlantic): Seasonal contrasts and a comparison of stained and dead assemblages. *Marine Micropaleontology*, 46, 83–110.
- Gooday, A. J., & Lambhead, P. J. D. (1989). Influence of seasonally deposited phytodetritus on benthic foraminiferal populations in the bathyal northeast Atlantic: The species response. *Marine Ecology Progress Series*, 5, 53–67.
- Gooday, A. J., Levin, L. A., Linke, P., & Heeger, T. (1992). The role of benthic foraminifera in deep-sea food webs and carbon cycling. In: G. T. Rowe & V. Pariente (Eds), *Deep-sea food chains and the global carbon cycle* (pp. 63–91). New York, NY: Kluwer Academic Publishers.
- Gooday, A. J., & Rathburn, A. E. (1999). Temporal variability in living deep-sea foraminifera: A review. *Earth Sciences Reviews*, 46, 187–212.
- Greiner, G. O. G. (1974). Environmental factors controlling the distribution of benthonic foraminifera. *Breviora*, 420, 1–35.
- Grimsdale, T. F., & Van Morkhoven, F. P. C. M. (1955). The ratio between pelagic and benthonic foraminifera as a means of estimating depth of deposition of sedimentary rocks. *Proceedings of 4th World Petroleum Congress, Section I/D4* (pp. 473–491).
- Guichard, S., Jorissen, F. J., Bertrand, P., Gervais, A., Martinez, P., Peypouquet, J.-P., Pujol, C., & Vergnaud-Grazzini, C. (1997). Foraminifères benthiques et paléoproduktivité : réflexions sur une carotte de l'upwelling (NW Africain). *Comptes rendus de l'Académie des Sciences, Paris. Sciences de la Terre et des Planètes*, 325, 65–70.
- Guichard, S., Jorissen, F. J., & Peypouquet, J. P. (1999). Late Quaternary benthic foraminiferal records testifying lateral variability of the Cape Blanc upwelling signal. *Comptes Rendus de l'Académie des Sciences, Paris. Sciences de la Terre et des Planètes*, 329, 295–301.
- Gundersen, J. K., & Jørgensen, B. B. (1990). Microstructure of diffusive boundary layers and oxygen uptake of the sea floor. *Nature*, 345, 604–607.
- Hannah, F., & Rogerson, A. (1997). The temporal and spatial distribution of foraminiferans in marine benthic sediments of the Clyde Sea area, Scotland. *Estuarine, Coastal and Shelf Science*, 44, 377–383.
- Hayward, B. W. (2004). Foraminifera-based estimates of paleobathymetry using Modern Analogue Technique, and the subsidence history of the early Miocene Waitemata Basin, New Zealand. *Journal of Geology and Geophysics*, 47, 749–767.
- Heinz, P., Schmiedl, G., Kitazato, H., & Hemleben, C. (2001). Response of deep-sea benthic foraminifera from the Mediterranean Sea to simulated phytoplankton pulses under laboratory conditions. *Journal of Foraminiferal Research*, 31, 210–227.
- Herguera, J. C. (1992). Deep-sea benthic foraminifera and biogenic opal: Glacial to postglacial productivity changes in the western equatorial Pacific. *Marine Micropaleontology*, 18, 79–98.
- Herguera, J. C. (2000). Last glacial paleoproductivity patterns in the eastern equatorial Pacific: Benthic foraminifera records. *Marine Micropaleontology*, 40, 259–275.
- Herguera, J. C., & Berger, W. (1991). Paleoproductivity from benthic foraminifera abundance: Glacial to postglacial change in the west-equatorial Pacific. *Geology*, 19, 1173–1176.
- Hess, S., Jorissen, F. J., Venet, V., & Abu-Zied, R. (2005). Benthic foraminiferal recovery after recent turbidite deposition in Cap Breton Canyon, Bay of Biscay. *Journal of Foraminiferal Research*, 35, 114–129.
- Hintz, C. J., Chandler, G. T., Bernhard, J. M., McCorkle, D. C., Havach, S. M., Blanks, J. K., & Shaw, T. J. (2004). A physicochemically constrained seawater culturing system for production of benthic foraminifera. *Limnology and Oceanography: Methods*, 2, 160–170.

- Hughes, J. A., & Gooday, A. J. (2004). Associations between living benthic foraminifera and dead tests of *Syringammina gracilissima* (Xenophyophorea) in the Darwin Mounds region (NE Atlantic). *Deep-Sea Research*, *1*, 51, 1741–1758.
- Imbrie, J., & Kipp, N. (1971). A new micropaleontological method for quantitative paleoclimatology: Application to a late Pleistocene Caribbean core. In: K. K. Turekian (Ed.), *The late Cenozoic glacial ages* (pp. 71–181). New Haven, CT: Yale University Press.
- Ittekkot, V. (1993). The abiotically driven biological pump in the ocean and short-term fluctuations in atmospheric CO₂ contents. *Global and Planetary Change*, *8*, 17–25.
- Jackson, G. A., & Burd, A. B. (2002). A model for the distribution of particle flux in the mid-water column controlled by subsurface biotic interactions. *Deep-Sea Research II*, *49*, 193–217.
- Jahnke, R. A., Emerson, S. R., & Murray, J. W. (1982). A model of oxygen reduction, denitrification, and organic matter mineralization in marine sediments. *Limnology and Oceanography*, *27*, 610–623.
- Jannink, N. T., Van der Zwaan, G. J., Almogi-Labin, A., Duijnste, I., & Jorissen, F. J. (2001). A transfer function for the quantitative reconstruction of oxygen contents in marine paleo-environments. *Geologica Ultraeicincta*, *203*, 161–169.
- Jannink, N. T., Zachariasse, W. J., & Van der Zwaan, G. J. (1998). Living (Rose Bengal stained) benthic foraminifera from the Pakistan continental margin (northern Arabian Sea). *Deep-Sea Research I*, *45*, 1483–1513.
- Jian, Z., Wang, L., Kienast, M., Sarntheim, M., Kuhnt, W., Lin, H., & Wang, P. (1999). Benthic foraminiferal paleoceanography of the South China Sea over the last 40,000 years. *Marine Geology*, *156*, 159–186.
- Jorissen, F. J. (1999a). Benthic foraminiferal microhabitats below the sediment–water interface. In: B. K. Sen Gupta (Ed.), *Modern Foraminifera* (pp. 161–179). Dordrecht: Kluwer Academic Publishers.
- Jorissen, F. J. (1999b). Benthic foraminiferal successions across late Quaternary Mediterranean sapropels. In E. J. Rohling (Ed.), *Fifth decade of Mediterranean paleoclimate and sapropel studies*, *Marine Geology*, *153*, 91–101.
- Jorissen, F. J., Barmawidjaja, D. M., Puskaric, S., & Van der Zwaan, G. J. (1992). Vertical distribution of benthic Foraminifera in the Northern Adriatic Sea. The relation with high organic flux. *Marine Micropaleontology*, *19*, 131–146.
- Jorissen, F. J., De Stigter, H. C., & Widmark, J. G. V. (1995). A conceptual model explaining benthic foraminiferal microhabitats. *Marine Micropaleontology*, *22*, 3–15.
- Jorissen, F. J., & Wittling, I. (1999). Ecological evidence from taphonomical studies; living–dead comparisons of benthic foraminiferal faunas off Cape Blanc (NW Africa). *Palaogeography, Palaoclimatology, Palaecoecology*, *149*, 151–170.
- Jorissen, F. J., Wittling, I., Peypouquet, J. P., Rabouille, C., & Relexans, J. C. (1998). Live benthic foraminiferal faunas off Cap Blanc, NW Africa: Community structure and microhabitats. *Deep-Sea Research I*, *45*, 2157–2188.
- Kaiho, K. (1991). Global changes of Paleogene aerobic/anaerobic benthic foraminifera and deep-sea circulation. *Palaogeography, Palaoclimatology, Palaecoecology*, *83*, 65–85.
- Kaiho, K. (1994). Benthic foraminiferal dissolved-oxygen index and dissolved-oxygen levels in the modern ocean. *Geology*, *22*, 719–722.
- Kaiho, K. (1999). Evolution in the test size of deep-sea benthic foraminifera during the past 120 m.y.. *Marine Micropaleontology*, *37*, 53–65.
- Kelly-Gerrey, B. A., Hydes, D. J., & Waniek, J. J. (2005). Control of the diffusive boundary layer on benthic fluxes: A model study. *Marine Ecology Progress Series*, *292*, 61–74.
- Kemp, A. E. S., Baldauf, J. G., & Pearce, R. B. (1995). Origins and paleoceanographic significance of laminated diatom ooze from the eastern equatorial Pacific. *Proceedings Ocean Drilling Program*, *138*, 641–663.
- Kitazato, H. (1994). Diversity and characteristics of benthic foraminiferal microhabitats in four marine environments around Japan. *Marine Micropaleontology*, *24*, 29–41.
- Kitazato, H., Nomaki, H., Heinz, P., & Nakatsuka, T. (2003). The role of benthic foraminifera in deep-sea food webs at the sediment–water interface: Results from in situ feeding experiments in Sagami Bay. *Frontier Research on Earth Evolution*, *1*, 227–232.

- Kitazato, H., Shirayama, Y., Nakatsuka, T., Fujiwara, S., Shimanaga, M., Kato, Y., Okada, Y., Kanda, J., Yamaoka, A., Masukawa, T., & Suzuki, K. (2000). Seasonal phytodetritus deposition and responses of bathyal benthic foraminiferal populations in Sagami Bay, Japan: Preliminary results from "Project Sagami 1996–1999". *Marine Micropaleontology*, *40*, 135–149.
- Kuhnt, W., Hess, S., & Jian, Z. (1999). Quantitative composition of benthic foraminiferal assemblages as a proxy indicator for organic carbon flux rates in the South China Sea. *Marine Geology*, *156*, 123–157.
- Lagoe, M. L. (1977). Recent benthic foraminifera from the Central Arctic Ocean. *Journal of Foraminiferal Research*, *7*, 106–129.
- Lambshhead, P. J. D., & Gooday, A. J. (1990). The impact of seasonally deposited phytodetritus on epifaunal and shallow infaunal benthic foraminiferal populations in the bathyal northeast Atlantic: The assemblage response. *Deep-Sea Research I*, *37*, 1263–1283.
- Lampitt, R. S. (1985). Evidence for seasonal deposition of detritus to deep sea floor and its subsequent resuspension. *Deep-Sea Research*, *32*, 885–897.
- Lea, D. L. (2004). Elemental and Isotopic Proxies of Past Ocean Temperatures. In: H. Elderfield (Ed.), *Treatise on geochemistry* (Vol. 6, pp. 365–390). The Oceans and Marine Geochemistry, Chapter 14. Amsterdam: Elsevier.
- Lear, H., Elderfield, H., & Wilson, P. A. (2000). Cenozoic deep-sea temperatures and global ice volumes from Mg/Ca in benthic foraminiferal calcite. *Science*, *287*, 269–272.
- Lear, C. H., Rosenthal, Y., Coxall, H. K., & Wilson, P. A. (2004). Late Eocene to early Miocene ice sheet dynamics and the global carbon cycle. *Paleoceanography*, *19*, PA4015, doi:10.1029/2004PA001039, 2004.
- Levin, L. A. (2003). Oxygen minimum zone benthos: Adaptation and community response to hypoxia. *Oceanography and Marine Biology: An Annual Review*, *41*, 1–45.
- Levin, L. A., Etter, R. J., Rex, M. A., Gooday, A. J., Smith, C. R., Pineda, J., Stuart, C. T., Hessler, R. R., & Pawson, D. (2001). Environmental influences on regional deep-sea species diversity. *Annual Review of Ecology and Systematics*, *132*, 51–93.
- Levin, L. A., & Gage, J. D. (1998). Relationships between oxygen, organic matter and the diversity of bathyal macrofauna. *Deep-Sea Research*, *45*, 129–163.
- Levinton, J. S. (1970). The paleoecological significance of opportunistic species. *Lethaia*, *3*, 69–78.
- Licari, L. N., Schumacher, S., Wenzhöfer, F., Zabel, M., & Mackensen, A. (2003). Communities and microhabitats of living benthic foraminifera from the tropical East Atlantic: Impact of different productivity regimes. *Journal of Foraminiferal Research*, *33*, 10–31.
- Linke, P., & Lutze, G. F. (1993). Microhabitats preferences of benthic foraminifera – A static concept or a dynamic adaptation to optimise food acquisition? In: M. R. Langer (Ed.), *Foraminiferal Microhabitats*. *Marine Micropaleontology*, *20*, 215–234.
- Lohmann, G. P. (1978a). Abyssal benthonic foraminifera as hydrographic indicators in the western South Atlantic ocean. *Journal of Foraminiferal Research*, *8*, 6–34.
- Lohmann, G. P. (1978b). Response of the deep sea to ice ages. *Oceanus*, *21*, 58–64.
- Lohrenz, S. E., Knauer, G. A., Asper, V. L., Tuel, M., Michaels, A. F., & Knap, A. H. (1992). Seasonal variability in primary production and particle flux in the northwestern Sargasso Sea: U.S. JGOFS Bermuda Atlantic Time-series Study. *Deep-Sea Research*, *39*, 1373–1391.
- Loubere, P. (1991). Deep sea benthic foraminiferal assemblage response to a surface ocean productivity gradient: A test. *Paleoceanography*, *6*, 193–204.
- Loubere, P. (1994). Quantitative estimation of surface ocean productivity and bottom water oxygen concentration using benthic foraminifera. *Paleoceanography*, *9*, 723–737.
- Loubere, P. (1996). The surface ocean productivity and bottom water oxygen signals in deep water benthic foraminiferal assemblages. *Marine Micropaleontology*, *28*, 247–261.
- Loubere, P. (1998). The impact of seasonality on the benthos as reflected in the assemblages of deep-sea foraminifera. *Deep-Sea Research I*, *45*, 409–432.
- Loubere, P., & Fariduddin, M. (1999). Benthic foraminifera and the flux of organic carbon to the seabed. In: B. K. Sen Gupta (Ed.), *Modern foraminifera* (pp. 181–199). Dordrecht: Kluwer.
- Loubere, P., Gary, A., & Lagoe, M. (1993). Benthic foraminiferal microhabitats and the generation of a fossil assemblage: Theory and preliminary data. *Marine Micropaleontology*, *20*, 165–181.

- Lozano, J. A., & Hays, J. D. (1976). Relationship of radiolarian assemblages to sediment types and physical oceanography in the Atlantic and western Indian oceanic sectors of the Antarctic Ocean. In R. M. Cline, J. D. Hays (Eds), *Investigations of Late Quaternary Paleooceanography and Paleoclimatology*. Geological Society of America, *Memoirs*, 145, 303–336.
- Lutze G. F. (1979). Benthic foraminifers at Site 397: fluctuations and ranges in the Quaternary. In: U. von Rad, W. B. F. Ryan et al. (Eds), *Init. Repts. DSDP* (Vol. 47 (Pt. 1), pp. 419–431), Washington: U.S. Government Printing Office.
- Lutze, G. F. (1980). Depth distribution of benthic foraminifera on the continental margin off NW Africa. *Meteor Forschungs-Ergebnisse*, C32, 31–80.
- Lutze, G. F., & Altenbach, A. (1988). *Rupertina stabilis* (WALLICH), a highly adapted, suspension feeding foraminifer. *Meyniana*, 40, 55–69.
- Lutze, G. F., & Coulbourn, W. T. (1984). Recent benthic foraminifera from the continental margin of northwest Africa: Community structure and distribution. *Marine Micropaleontology*, 8, 361–401.
- Lutze, G. F., Pflaumann, U., & Weinholz, P. (1986). Jungquartäre Fluktuationen der benthischen Foraminiferenfaunen in Tiefsee-Sedimenten vor NW-Afrika — Eine Reaktion auf Produktivitätsänderungen im Oberflächennwasser. *Meteor Forschungs-Ergebnisse*, C40, 163–180.
- Lutze, G. F., & Thiel, H. (1989). Epibenthic foraminifera from elevated microhabitats: *Cibicoides wuellerstorfi* and *Planulina ariminensis*. *Journal of Foraminiferal Research*, 19, 153–158.
- Lynch-Stieglitz, J. (2004). Tracers of past ocean circulation. In: H. Elderfield (Ed.), *Treatise on geochemistry* (Vol. 6, pp. 433–451). The Oceans and Marine Geochemistry, Chapter 16. Amsterdam: Elsevier.
- Maas, M. (2000). Verteilung lebendgefärbter benthischer Foraminiferen in einer intensivierten Sauerstoffminimumzone, Indo-Pazifischer Kontinentalrand, nördliches Arabisches Meer. *Meyniana*, 52, 101–129.
- Mackensen, A., & Douglas, R. G. (1989). Down-core distribution of live and dead deep-water benthic foraminifera in box cores from the Weddell Sea and the California continental borderland. *Deep-Sea Research*, 36, 879–900.
- Mackensen, A., Grobe, H., Kuhn, G., & Fütterer, D. K. (1990). Benthic foraminiferal assemblages from the eastern Weddell Sea between 68 and 73°S: Distribution, ecology and fossilization potential. *Marine Micropaleontology*, 16, 241–283.
- Mackensen, A., Schmiedl, G., Harloff, J., & Giese, M. (1995). Deep-sea foraminifera in the South Atlantic Ocean: Ecology and assemblage generation. *Micropaleontology*, 41, 342–358.
- Martin, J. H., Knauer, G. A., Karl, D. M., & Broenkow, W. W. (1987). VERTEX: Carbon cycling in the north-east pacific. *Deep-Sea Research*, 34, 267–285.
- Martinez, P., Bertrand, P., Shimmield, G. B., Cochran, K., Jorissen, F. J., Foster, J., & Dignan, M. (1999). Upwelling intensity and ocean productivity changes off Cape Blanc (northwest Africa) during the last 70,000 years: Geochemical and micropaleontological evidence. *Marine Geology*, 158, 57–74.
- Miao, Q., & Thunell, R. C. (1993). Recent deep-sea benthic foraminiferal distributions in the South China and Sulu Seas. *Marine Micropaleontology*, 22, 1–32.
- Miller, K. G., & Lohmann, G. P. (1982). Environmental distribution of Recent benthic foraminifera on the northeast United States continental slope. *Geological Society of America, Bulletin*, 93, 200–206.
- Moodley, L., Heip, C. H. R., & Middelburg, J. J. (1998b). Benthic activity in sediments of the northwestern Adriatic Sea: Sediment oxygen consumption, macro- and meiofauna dynamics. *Journal of Sea Research*, 40, 263–280.
- Morigi, C., Jorissen, F. J., Gervais, A., Guichard, S., & Borsetti, A. M. (2001). Benthic foraminiferal faunas in surface sediments off NW Africa: Relationship with the organic flux to the ocean floor. *Journal of Foraminiferal Research*, 31, 350–368.
- Müller, P. J., & Suess, E. (1979). Productivity, sedimentation rate, and sedimentary organic matter in the oceans. I. Organic carbon preservation. *Deep-Sea Research*, 26, 1347–1362.
- Mullineaux, L. S., & Lohmann, G. P. (1981). Late Quaternary Stagnations and recirculation of the eastern Mediterranean: Changes in the deep water recorded by fossil benthic foraminifera. *Journal of Foraminiferal Research*, 11, 20–39.

- Murray, J. W. (1973). *Distribution and ecology of living benthic foraminiferids*. London: Heinemann, 288 pp.
- Murray, J. W. (1988). Neogene bottom water-masses and benthic foraminifera in the NE Atlantic Ocean. *Journal of the Geological Society, London*, 145, 125–132.
- Murray, J. W. (1991). *Ecology and paleoecology of benthic foraminifera* (p. 397). Harlow: Longman.
- Murray, J. W. (2001). The niche of benthic foraminifera, critical thresholds and proxies. *Marine Micropaleontology*, 41, 1–7.
- Murray, J. W., Weston, J. F., Haddon, C. A., & Powell, A. D. J. (1986). Miocene to recent bottom water masses of the north-east Atlantic: An analysis of benthic foraminifera. In: C. P. Summerhayes & N. J. Shackleton (Eds), *North Atlantic Palaeoceanography* (Vol. 21, pp. 219–230). London: Special Publication of the Geological Society.
- Murray, J., Young, J., Newton, J., Dunne, J., Chapin, T., Paul, B., & McCarthy, J. J. (1996). Export flux of particulate organic carbon from the central equatorial Pacific determined using a combined drifting trap-²³⁴Th approach. *Deep Sea Research II*, 43, 1095–1132.
- Naidu, P. D., & Malmgren, B. A. (1995). Do benthic foraminifer records represent a productivity index in oxygen minimum zone areas? An evaluation from the Oman Margin, Arabian Sea. *Marine Micropaleontology*, 26, 49–55.
- Natland, M. L. (1933). The temperature- and depth-distribution of some Recent and fossil Foraminifera in the Southern California Region. *Scripps Institute of Oceanography Bulletin, Technical Series*, 3, 225–230.
- Natland, M. L., & Kuenen, P. H. (1951). Sedimentary history of the Ventura Basin, California, and the action of turbidity currents. In: J. L. Hough (Ed.), *Turbidity currents and the transportation of coarse sediments to deep water. A symposium* (Vol. 2, pp. 76–107). Tulsa, OK: Society of Economic Paleontologists and Mineralogists, Special Publication.
- Nees, S., Altenbach, A. V., Kassens, H., & Thiede, J. (1997). High-resolution record of foraminiferal response to Late Quaternary sea-ice retreat in the Norwegian-Greenland Sea. *Geology*, 25, 659–662.
- Nees, S., Armand, L., De Deckker, P., Labracherie, M., & Passlow, V. (1999). A diatom and benthic foraminiferal record from the South Tasman Rise (southeastern Indian Ocean): Implications for palaeoceanographic changes for the last 200,000 years. *Marine Micropaleontology*, 38, 69–89.
- Nees, S., & Struck, U. (1999). Benthic foraminiferal response to major oceanographic changes. In: F. Abrantes & A. Mix (Eds), *Reconstructing ocean history: A window into the future* (pp. 195–216). Dordrecht: Kluwer Academic/Plenum Publishers.
- Nelson, D. M., Anderson, R. F., Barber, R. T., Brzezinski, M. A., Buesseler, K. O., Chase, Z., Collier, R. W., Dickson, M. L., François, R., Hiscock, M. R., Honjo, S., Marra, J., Martin, W. R., Sambrotto, R. N., Sayles, F. L., & Sigmon, D. E. (2002). Vertical budgets for organic carbon and biogenic silica in the Pacific sector of the Southern Ocean, 1996–1998. *Deep Sea Research Part II: Topical Studies in Oceanography*, 49(9–10), 1645–1674.
- Nittrouer, C. A., DeMaster, D. J., McKee, B. A., Cutshall, N. H., & Larsen, I. L. (1984). The effect of sediment mixing on Pb-210 accumulation rates for the Washington continental shelf. *Marine Geology*, 54, 201–221.
- Nomaki, H., Heinz, P., Hemleben, C., & Kitazato, H. (2005). Behavior and response of deep-sea benthic foraminifera to freshly supplied organic matter: A laboratory feeding experiment in microcosm environments. *Journal of Foraminiferal Research*, 35, 103–113.
- Ohga, T., & Kitazato, H. (1997). Seasonal changes in bathyal foraminiferal populations in response to the flux of organic matter (Sagami Bay, Japan). *Terra Nova*, 9, 33–37.
- Ohkushi, K., Thomas, E., & Kawahata, H. (2000). Abyssal benthic foraminifera from the North-western Pacific (Shatsky Rise) during the last 298 kyr. *Marine Micropaleontology*, 38, 119–147.
- Osterman, L. E., & Kellogg, T. B. (1979). Recent benthic foraminiferal distribution from the Ross Sea, Antarctica: Relation to ecological and oceanographic conditions. *Journal of Foraminiferal Research*, 9, 250–269.
- Pace, M. L., Knauer, G. A., Karl, D. M., & Martin, J. H. (1987). Primary production, new production, and vertical flux in the eastern Pacific Ocean. *Nature*, 325, 803–804.
- Parisi, E., & Cita, M. B. (1982). Late Quaternary paleoceanographic changes recorded by deep-sea benthos in the western Mediterranean Ridge. *Geografia Fisica e Dinamica Quaternaria*, 5, 102–114.

- Parker, F. L. (1958). Eastern Mediterranean Foraminifera. Reports of the Swedish deep-sea expedition 1947–1948, VIII: *Sediment cores from the Mediterranean and the Red Sea*, 4, 283.
- Peterson, L. C. (1984). Recent abyssal benthic foraminiferal biofacies of the eastern Equatorial Indian Ocean. *Marine Micropaleontology*, 8, 479–519.
- Peterson, L. C., & Lohmann, G. P. (1982). Major change in Atlantic Deep and Bottom Waters 700,000 yr Ago: Benthonic Foraminiferal Evidence from the South Atlantic. *Quaternary Research*, 17, 26–38.
- Pfannkuche, O., & Lochte, K. (1993). Open ocean pelago-benthic coupling: Cyanobacteria as tracers of sedimenting salp faeces. *Deep-Sea Research*, 40, 727–737.
- Pflum, C. E., & Frerichs, W. E. (1976). Gulf of Mexico Deep-Water Foraminifers. *Cushman Foundation for Foraminiferal Research, Special Publication*, 14, 125.
- Phleger, F. L. (1960). *Ecology and distribution of recent Foraminifera* (p. 297). Baltimore, MD: Johns Hopkins Press.
- Phleger, F. L., & Soutar, A. (1973). Production of benthic foraminifera in three east Pacific oxygen minima. *Micropaleontology*, 19, 110–115.
- Pianka, E. R. (1970). On r and K selection. *American Naturalist*, 104, 592–597.
- Poliski, W. (1959). Foraminiferal biofacies off North Asiatic Coast. *Journal of Paleontology*, 33, 569–587.
- Pujos-Lamy, A. (1973). Répartition bathymétrique des foraminifères benthiques du Golfe de Gascogne. *Revista Española De Micropaleontología*, 5, 213–234.
- Rabouille, C., & Gaillard, J. F. (1991). A coupled model representing the deep-sea organic carbon mineralization and oxygen consumption in surficial sediments. *Journal of Geophysical Research*, 96, 2761–2776.
- Rathburn, A. E., & Corliss, B. H. (1994). The ecology of living (stained) deep-sea benthic foraminifera from the Sulu Sea. *Paleoceanography*, 9, 87–150.
- Rathburn, A. E., Corliss, B. H., Tappa, K. D., & Lohmann, K. C. (1996). Comparison of the ecology and stable isotopic compositions of living (stained) benthic foraminifera from the Sulu and South China Seas. *Deep-Sea Research I*, 43, 1617–1646.
- Rathburn, A. E., Perez, M. E., & Lange, C. (2001). Benthic-pelagic coupling in The Southern California bight: Relationships between sinking organic material, diatoms and benthic foraminifera. *Marine Micropaleontology*, 43, 261–271.
- Rathburn, A. E., Perez, M. E., Martin, J. B., Day, S. E., Mahn, C., Gieskes, J., Ziebis, W., Williams, D., & Bahls, A., (2003). Relationships between the distribution and stable isotopic composition of living benthic foraminifera and cold methane seeps biogeochemistry in Monterey Bay, California. *Geochemistry, Geophysics, Geosystems*, 4(12), doi:10.1029/2003GC000595.
- Ravizza, G. E., & Zachos, J. C. (2004). Records of Cenozoic Ocean Chemistry. In: H. Elderfield (Ed.), *Treatise on geochemistry* (Vol. 6, pp. 551–581). The Oceans and Marine Geochemistry, Chapter 20. Amsterdam: Elsevier.
- Reimers, C. E. (1987). An in situ microprofiling instrument for measuring interfacial pore water gradients: Methods and oxygen profiles from the North Pacific Ocean. *Deep-Sea Research*, 34, 2019–2035.
- Robinson, B. H., Reisenbichler, K. R., & Sherlock, R. E. (2005). Giant larvacean houses: Rapid carbon transport to the deep sea floor. *Science*, 308, 1609–1611.
- Rutgers van der Loeff, M. M. (1990). Oxygen in pore waters of deep-sea sediments. *Philosophical Transactions Royal Society London, A*, 331, 69–84.
- Saidova, K. M. (1961). Ecology of the foraminifera and paleoceanography of the Far Eastern seas of the USSR and the northwestern part of the Pacific Ocean. *Akademie Nauk SSSR, Instituta Okeanologii*, 45, 65–71 (in Russian).
- Sarnthein, M., Winn, K., Duplessy, J. C., & Fontugne, M. R. (1988). Global variations of surface ocean productivity in low and mid latitudes: Influence on CO₂ reservoirs of the deep ocean and atmosphere during the last 21,000 years. *Paleoceanography*, 3, 361–399.
- Schemainda, R., Nehring, D., & Schultz, S. (1975). Ozeanologische untersuchungen zum produktionspotential der Nordwestafrikanischen wasserauftriebsregion 1970–1973. *Nationalkomitee für Geodäsie und Geophysik bei der Akademie der Wissenschaften der Deutschen Demokratischen Republik, IV/16*, 85.

- Schmiedl, G., De Bovée, F., Buscail, R., Charrière, B., Hemleben, C., Medernach, L., & Picon, P. (2000). Trophic control of benthic foraminiferal abundance and microhabitat in the bathyal Gulf of Lions, western Mediterranean Sea. *Marine Micropaleontology*, *40*, 167–188.
- Schmiedl, G., & Mackensen, A. (1997). Late Quaternary paleoproductivity and deep water circulation in the eastern South Atlantic Ocean: Evidence from benthic foraminifera. *Palaeogeography, Palaeoclimatology, Palaeoecology*, *130*, 43–80.
- Schmiedl, G., Mackensen, A., & Müller, P. J. (1997). Recent benthic foraminifera from the eastern South Atlantic Ocean: Dependence on food supply and water masses. *Marine Micropaleontology*, *32*, 249–287.
- Schmiedl, G., Mitschele, A., Beck, S., Emeis, K. C., Hemleben, C., Schulz, H., Sperling, M., & Weldeab, S. (2003). Benthic foraminiferal record of ecosystem variability in the eastern Mediterranean Sea during times of sapropel S₅ and S₆ deposition. *Palaeogeography, Palaeoclimatology, Palaeoecology*, *190*, 139–164.
- Schmitz, W. J. (1992). On the intrabasinal thermohaline circulation. *Reviews of Geophysics*, *33*, 151–173.
- Schnitker, D. (1974). West abyssal circulation during the past 120,000 years. *Nature*, *248*, 385–387.
- Schnitker, D. (1976). Structure and cycles of the western North Atlantic bottom water, 24,000 years BP to present. *EOS*, *57*, 257–258.
- Schnitker, D. (1979). The deep waters of the western North Atlantic during the past 24,000 years and the re-initiation of the Western Boundary Undercurrent. *Marine Micropaleontology*, *4*, 264–280.
- Schnitker, D. (1980). Quaternary deep-sea benthic foraminifera and bottom water masses. *Annual Review of Earth and Planetary Science*, *8*, 343–370.
- Schönfeld, J. (1997). The impact of the Mediterranean Outflow Water (MOW) on benthic foraminiferal assemblages and surface sediments at the southern Portuguese continental margin. *Marine Micropaleontology*, *29*, 211–236.
- Schönfeld, J. (2002a). Recent benthic foraminiferal assemblages in deep high-energy environments from the Gulf of Cadiz (Spain). *Micropaleontology*, *44*, 141–162.
- Schönfeld, J. (2002b). A new benthic foraminiferal proxy for near-bottom current velocities in the Gulf of Cadiz, northeastern Atlantic Ocean. *Deep-Sea Research I*, *49*, 1853–1875.
- Schönfeld, J., & Altenbach, A. V. (2005). Late Glacial to Recent distribution pattern of deep-water *Uvigerina* species in the north-eastern Atlantic. *Marine Micropaleontology*, *57*, 1–24.
- Schrader, H. J. (1974). Cenozoic marine planktonic diatom stratigraphy of the tropical Indian Ocean. In: R. L. Fisher & E. T. Bunce (Eds), *Initial Reports of the Deep Sea Drilling Project* (Vol. 24, pp. 887–968). Washington: U. S. Government Printing Office.
- Sen Gupta, B. (Ed.) (1999). *Modern foraminifera* (p. 371). Dordrecht: Kluwer Academic Publishers.
- Sen Gupta, B. K., & Machain-Castillo, M. L. (1993). Benthic foraminifera in oxygen-poor habitats. *Marine Micropaleontology*, *20*, 3–4.
- Sen Gupta, B. K., Platon, E., Bernhard, J. M., & Aharon, P. (1997). Foraminiferal colonization of hydrocarbon-seep bacterial mats and underlying sediment, Gulf of Mexico Slope. *Journal of Foraminiferal Research*, *27*, 292–300.
- Sen Gupta, B. K., Smith, L. E., & Lobegeier, M. K. (2006). Attachment of Foraminifera to vestimentiferan tubeworms at cold seeps: Refuge from sea-floor hypoxia and sulfide toxicity. *Marine Micropaleontology*, *62*(1), 1–6.
- Shirayama, Y. (1984). The abundance of deep sea meiobenthos in the Western Pacific in relation to environmental factors. *Oceanologica Acta*, *7*, 113–121.
- Sigman, D. M., & Haug, G. H. (2004). The biological pump in the past. In: H. Elderfield (Ed.), *Treatise on geochemistry* (Vol. 6, pp. 491–528) *The Oceans and Marine Geochemistry*, Chapter 18. Amsterdam: Elsevier.
- Sliter, W. V., & Baker, R. A. (1972). Cretaceous bathymetric distribution of benthic foraminifera. *Journal of Foraminiferal Research*, *2*, 167–183.
- Smart, C. W. (2002). Environmental applications of deep-sea benthic foraminifera. In: S. K. Haslett (Ed.), *Quaternary Environmental Micropalaeontology* (pp. 14–58). London: Arnold Publishers.
- Smart, C. W., King, S. C., Gooday, A. J., Murray, J. W., & Thomas, E. (1994). A benthic foraminiferal proxy of pulsed organic matter paleofluxes. *Marine Micropaleontology*, *23*, 89–99.

- Smith, P. B. (1964). Recent foraminifera off Central America. Ecology of benthic species. *USGS Professional Paper*, 429B, 1–55.
- Soetaert, K., Herman, P. M. J., & Middelburg, J. J. (1996). A model of early diagenetic processes from the shelf to abyssal depths. *Geochimica et Cosmochimica Acta*, 60, 1019–1040.
- Streeter, S. S. (1973). Bottom water and benthonic foraminifera in the North Atlantic-glacial-interglacial cycles. *Quaternary Research*, 3, 131–141.
- Streeter, S. S., Belanger, P. E., Kellogg, T. B., & Duplessy, J. C. (1982). Late Pleistocene paleo-oceanography of the Norwegian-Greenland sea: Benthic foraminiferal evidence. *Quaternary Research*, 18, 72–90.
- Streeter, S. S., & Shackleton, N. J. (1979). Paleocirculation of the deep North Atlantic: 150,000 year record of benthic foraminifera and oxygen-18. *Science*, 203, 168–171.
- Stschedrina, Z. G. (1957). Some regularities in the distribution of recent foraminifera. *Trudy Leningradskogo Obshch. Estest*, 73, 99–106.
- Suess, E. (1980). Particulate organic carbon flux in the ocean-surface productivity and oxygen utilization. *Nature*, 280, 260–263.
- Suhr, S. B., Pond, D. W., Gooday, A. J., & Smith, C. R. (2003). Selective feeding by benthic foraminifera on phytodetritus on the western Antarctic Peninsula shelf: Evidence from fatty acid biomarker analysis. *Marine Ecology Progress Series*, 262, 153–162.
- Sun, X., Corliss, B. H., Brown, C. W., & Showers, W. J. (2006). The effect of primary productivity and seasonality on the distribution of deep-sea benthic foraminifera in the North Atlantic. *Deep-Sea Research I*, 53, 28–47.
- Tabachnick, B. G., & Fidell, L.S. (1983). *Using multivariate statistics* (p. 509). New York, NY: Harper & Row Publishers.
- Tengberg, A., De Bovee, F., Hall, P., Berelson, W., Chadwick, D., Cicceri, G., Crassous, P., Devol, A., Emerson, S., Gage, J., Glud, R., Graziottin, F., Gundersen, J., Hammond, D., Helder, W., Hinga, K., Holby, O., Jahnke, R., Khrifounoff, A., Lieberman, S., Nuppenau, V., Pfannkuche, O., Reimers, C., Rowe, G., Sahami, A., Sayles, F., Schuster, M., Smallman, D., Wehrli, B., & De Wilde, P. (1995). Benthic chamber and profiling landers in oceanography—A review of design, technical solutions and functioning. *Progress in Oceanography*, 35, 253–294.
- Thomas, E. (1986). Changes in composition of Neogene benthic foraminiferal faunas in equatorial Pacific and north Atlantic. *Palaeogeography, Palaeoclimatology, Palaeoecology*, 53, 47–61.
- Thomas, E. (2007). Cenozoic mass extinctions in the deep sea: What perturbs the largest habitat on earth? In: S. Monechi, M. Rampino & R. Coccioni (Eds), *Large ecosystem perturbations: Causes and consequences*. Geological Society of America Special paper, in press.
- Thomas, E., Booth, L., Maslin, M., & Shackleton, N. J. (1995). Northeastern Atlantic benthic foraminifera during the last 45,000 years: Changes in productivity seen from the bottom up. *Paleoceanography*, 10, 545–562.
- Thomas, E., Gooday, A. J. (1996). Cenozoic deep-sea benthic foraminifera: Tracers for changes in oceanic productivity? *Geology*, 24, 355–358
- Toyofuku, T., & Kitazato, H. (2005). Micromapping of Mg/Ca values in cultured specimens of the high-magnesium benthic foraminifera. *Geochemistry, Geophysics, and Geosystems*, 6, Q11P05.
- Turley, C. (2002). *The importance of 'marine snow': Microbiology Today*, 29, 177–179.
- Tyson, R. V., & Pearson, T. H. (1991). Modern and ancient continental shelf anoxia: An overview. In: Tyson, R. V., Pearson, T. H., (Eds), *Modern and Ancient Continental Shelf Anoxia* (Vol. 58, pp. 1–24). London: Geological Society of London Special Publication.
- Van Aken, H. M. (2000). The hydrography of the mid-latitude northeast Atlantic Ocean I: The deep water masses. *Deep-Sea Research I*, 47, 757–788.
- Van der Zwaan, G. J. (1982). Paleoecology of Late Miocene Mediterranean Foraminifera. *Utrecht Micropaleontological Bulletins*, 25, 202.
- Van der Zwaan, G. J., Duijnste, I. A. P., Den Dulk, M., Ernst, S. R., & Kouwenhoven, N. T. (1999). Benthic foraminifera: proxies or problems? A review of paleoecological concepts. *Earth Sciences Reviews*, 46, 213–236.
- Van der Zwaan, G. J., Jorissen, F. J., & De Stigter, H. (1990). The depth dependency of planktonic/benthic foraminiferal ratios: Constraints and applications. *Marine Geology*, 95, 1–16.

- Van Hinsbergen, D. J. J., Kouwenhoven, T. J., & Van der Zwaan, G. J. (2005). Paleobathymetry in the backstripping procedure: Correction for oxygenation effects on depth estimates. *Palaeogeography, Palaeoclimatology, Palaeoecology*, *221*, 245–265.
- Walker, S. E., & Goldstein, S. T. (1999). Taphonomic tiering: Experimental field taphonomy of molluscs and foraminifera above and below the sediment-water interface. *Palaeogeography, Palaeoclimatology, Palaeoecology*, *149*, 227–244.
- Walton, W. R. (1952). Techniques for recognition of living Foraminifera. *Contributions Cushman Foundation for Foraminiferal Research*, *3*, 56–60.
- Wassmann, P. (1993). Regulation of vertical export of particulate organic matter from the euphotic zone by planktonic heterotrophs in eutrophicated aquatic environments. *Marine Pollution Bulletin*, *26*, 636–643.
- Wefer, G., Berger, W. H., Bijma, J., & Fischer, G. (1999). Clues to ocean history: A brief overview of proxies. In: G. Fischer & G. Wefer (Eds), *Uses of proxies in Paleoceanography: Examples from the South Atlantic* (pp. 1–68). Berlin, Heidelberg: Springer-Verlag.
- Weinelt, M., Kuhnt, W., Sarnthein, M., Altenbach, A., Costello, O., Erlenkeuser, H., Pflaumann, U., Simstich, J., Thies, A., Trauth, M. H., & Vogelsand, E. (2001). Paleoceanographic proxies in the northern North Atlantic. In: P. Schäfer, W. Ritzrau, M. Schlüter & T. Thiede (Eds), *The northern North Atlantic: A changing environment* (pp. 319–352). Berlin, Heidelberg: Springer-Verlag.
- Weston, J. F. (1982). *Distribution and ecology of Recent Deep Sea benthic foraminifera in the Northeast Atlantic Ocean*. Ph.D. thesis, University of Exeter, England.
- Wilson-Finelli, A., Chandler, G. T., & Spero, H. J. (1998). Stable isotope behavior in paleoceanographically important benthic foraminifera: Results from microcosm culture experiments. *Journal of Foraminiferal Research*, *28*, 312–320.
- Wollenburg, J. E., & Kuhnt, W. (2000). The response of benthic foraminifera to carbon flux and primary production in the Arctic Ocean. *Marine Micropaleontology*, *40*, 189–231.
- Wollenburg, J. E., Kuhnt, W., & Mackensen, A. (2001). Changes in Arctic Ocean paleoproductivity and hydrography during the last 145 kyr: The benthic foraminiferal record. *Paleoceanography*, *16*, 65–77.
- Woodruff, F. (1985). Changes in Miocene deep-sea benthic foraminiferal distribution in the Pacific Ocean: Relationship to paleoceanography. In: Kennett, J. P. (Ed.), *The Miocene Ocean: Paleoceanography and Biogeography* (Vol. 163, pp. 131–175). Boulder, Memoir: Geological Society of America.
- Woodruff, F., & Savin, S. M. (1989). Miocene deepwater oceanography. *Paleoceanography*, *4*, 87–140.